

Shiga toxin producing *Escherichia coli* O174: A persistent STEC serogroup displaying virulence features of Uropathogenic *E. coli*

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Istituto Superiore di Sanità**



Disease caused by pathogenic *E. coli*

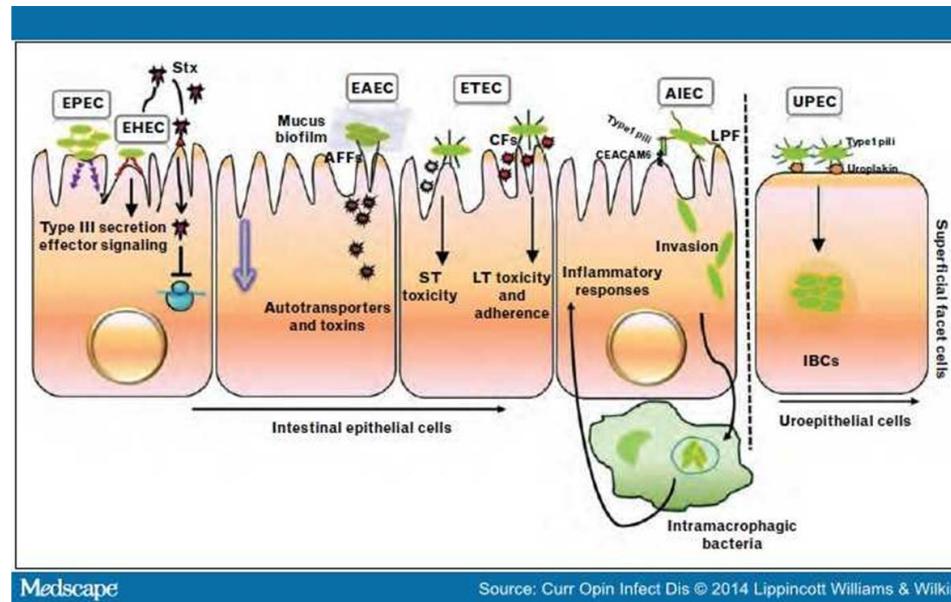
Urinary tract infections

ExPEC

Sepsis/meningitis

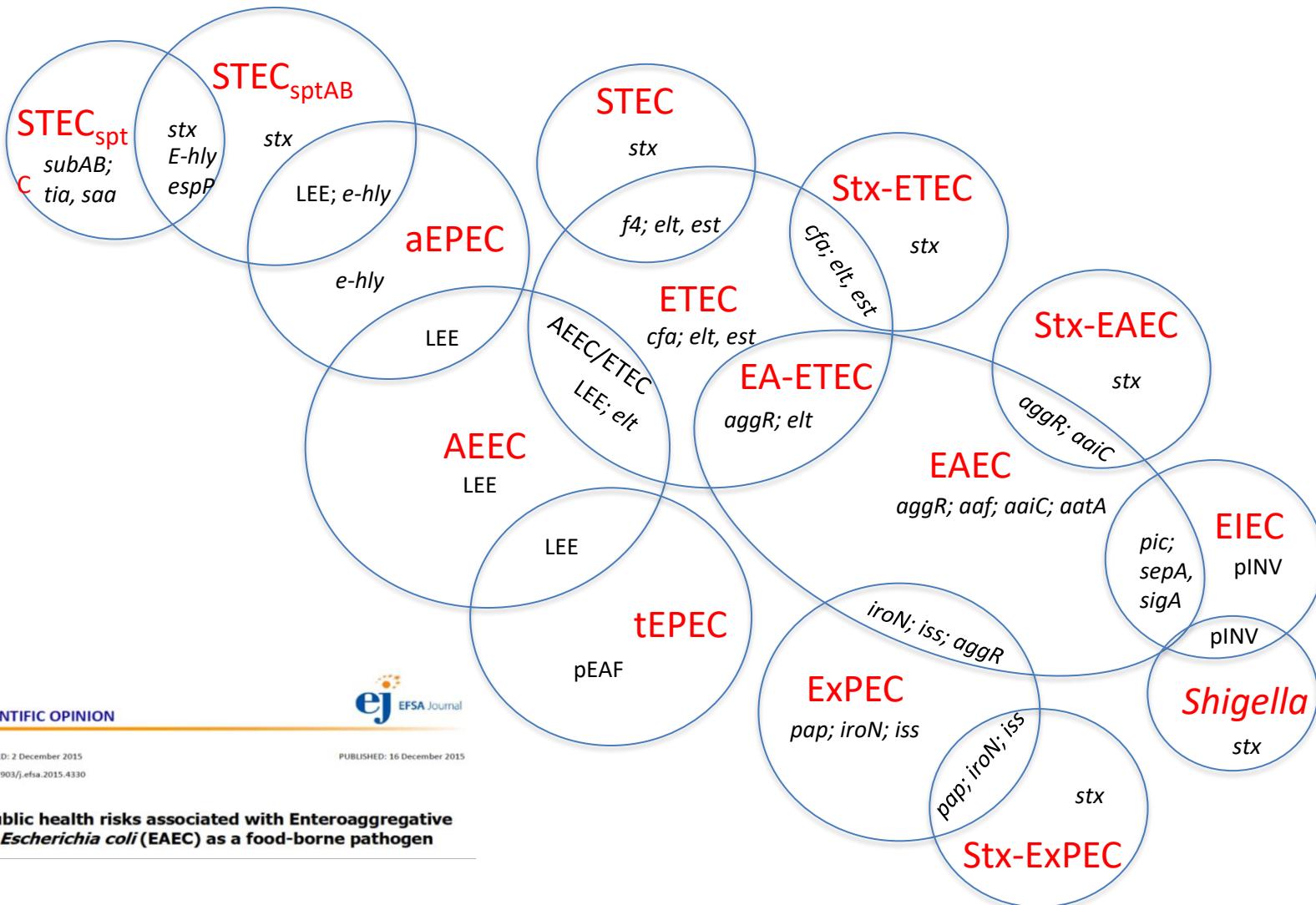
Enteric/diarrhoeal disease

DEC



Different virulence mechanisms,
according to their set of virulence determinants

The pathogenic *E. coli* Continuum



Cross-over pathotype combinations may occur

**2011 - German outbreak *E. coli* O104:H4:
4000 human cases 850 HUS 50 deaths**



Staked-brick adhesion



Stx-phage

Atypical virulence combination between EAEC and STEC

PLOS ONE

RESEARCH ARTICLE

Comparative Genomics and Characterization of Hybrid Shiga-toxigenic and Enterotoxigenic *Escherichia coli* (STEC/ETEC) Strains

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Hybrid ExPEC-STEC

EMERGING INFECTIOUS DISEASES®

ISSN: 1080-6059

RESEARCH

Enterohemorrhagic *Escherichia coli* Hybrid Pathotype O80:H2 as a New Therapeutic Challenge

Nurcan Soysal, Patricia Mariani-Kurdjian, Yasmine Smail, Sandrine Liguori, Malika Gouali, Estelle Loukiadis, Patrick Fach, Mathias Bruyand, Jorge Blanco, Philippe Bidet, Stéphane Bonacorsi

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Comparative phenotypic characterization of hybrid Shiga toxin-producing / uropathogenic *Escherichia coli*, canonical uropathogenic and Shiga toxin-producing *Escherichia coli*

Noble Selasi Gati ^{a,1}, Imke Johanna Temme ^{a,1}, Barbara Middendorf-Bauchart ^a, Alexander Kehl ^a, Ulrich Dobrindt ^b, Alexander Mellmann ^{a,*}



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- Discussion

STEC: Zoonotic origin



Natural reservoir

Routes of transmission:

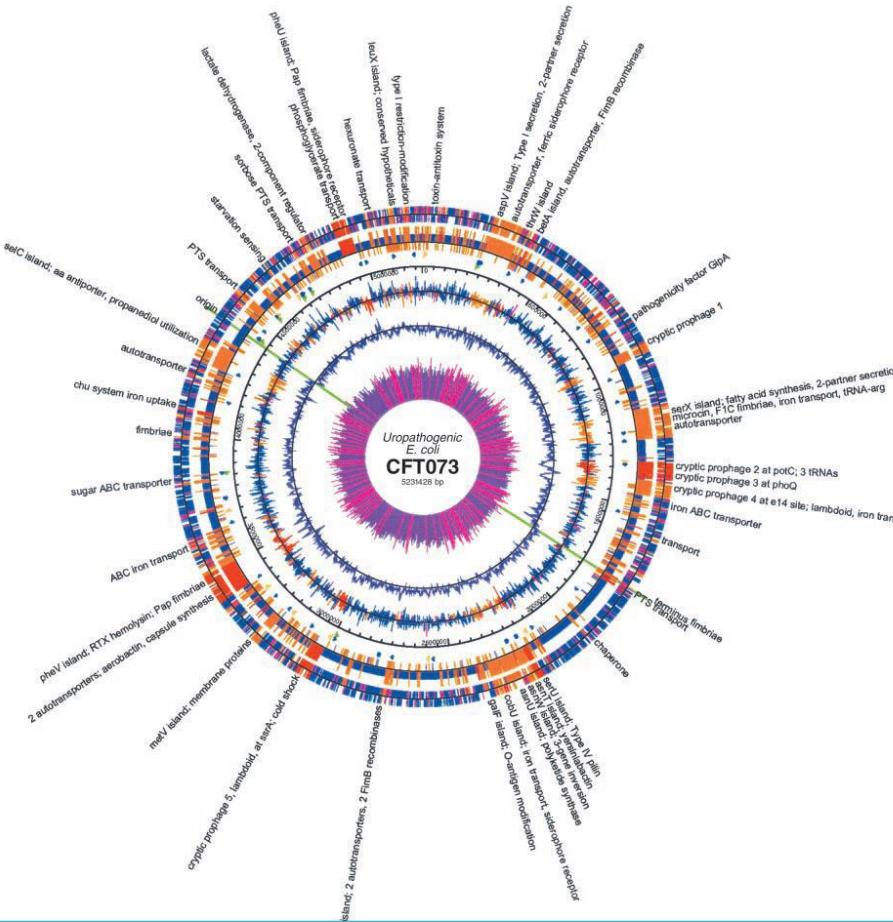
Foodborne - Primary contamination of meat and milk

Environmental contamination

Waterborne: Drinking and recreational water

Direct contact with infected animals

Uropathogenic *E. coli* (UPEC)



The common source of UPEC is the host's own intestinal flora

Table 1 Virulence genes encoding siderophores, toxins, adhesins, and protectins/serum resistance in UPEC.

Descriptions	Virulence gene (s)	Functions	References
Toxins			
Cytotoxic necrotizing factor	cnf	Involving in cells necrosis	Davis et al., 2005
Hemolysin A	hlyA	Cell lysis, inducing of pore in membrane of host cell	Bien et al., 2012
Vacuolating autotransporter toxin	var	Inducing vacuolization in host cell	Parham et al., 2005
Secreted autotransporter toxin	sar	Inducing cytotoxic effects on cells vacuolization	Guyer et al., 2002
Serine protease autotransporter	pic	Damaging the cell membrane, facilitating colonization epithelium, degrading mucus	Parham et al., 2005
Cytolytic distending toxin	cdt	Cytolytic factor, human cell apoptotic factor	Hinenoya et al., 2014
Collabactin	cib	PK-NRP compounds including DNA damage resulting apoptosis, and premature cellular destruction in epithelial cells	Morgan et al., 2019
Shigella enterotoxin-1	set-1	Invasion, inflammation	Mirzrazi et al., 2015
Arginine succinyltransferase	astA	Invasion, cytotoxic, inflammation	Jahandeh et al., 2015
Toll/interleukin receptor domain containing protein (Tcp)	tcpC	Bacterial survival, human avoidance system, cytopathic effect on kidney	He et al., 2017
Contact-dependent growth inhibition	cdiAB	Growth inhibition of the bacterial target	Hayes et al., 2014
Protectins/serum resistance			
Outer membrane protein	Omp	Enabling intracellular survival, porin	
Serum resistance proteins	iss, traT	Neutralization of anti-bactericidal effect of serum	Desloges et al., 2019;
D-serine deaminase	dsdCXA	Excite growth of bacteria using D-serine as carbon, and energy source, and inhibit the bacteriostatic activity of D-serine in the urine.	Nicholson et al., 2009
Capsule	kps	The K1 polysaccharide, a linear α2-8-linked sialic acid homopolymer, has a very important role in IBC development as well as in the multiple stages of UTI pathogenesis	Derakhshandeh et al., 2015
Moritz and Welch, 2006			
			Anderson et al., 2010
Iron acquisition systems			
SiABCD	siAB,C,D	Transporting Mn, Fe	
ChiA, Hma	chiA, hma	Enabling utilization of iron from hemoglobin in the host systems	Sabri et al., 2008
Salmochelin	iroA gene cluster	A siderophore,a modified form of enterobactin	Hagan and Mobley, 2009
	iroN	A siderophore receptor, utilizing Fe ions derived from the host bodies	Garcia et al., 2011
Aerobactin	fuc, aer	A siderophore, acquiring of Fe^{2+}/Fe^{3+} in the host systems	
Yersiniabactin	ybe	A high iron affinity siderophore, secreted alongside enterobactin	Garcia et al., 2011
Enterobactin	ent, fepA	A catecholate siderophore, higher iron affinity compared to aerobactin UPEC colonization in the urinary tract with restricted iron	Garcia et al., 2011
IreA	ireA	Iron regulated siderophore receptors involve in UTIS	Li et al., 2016
Adhesins			
Antigen43	csgn43(fhu)	A autotransporter protein, biofilm as well as adhesion development	Ulett et al., 2007
Curli fiber gene	Curli fiber (crl, csgB)	Promoting pathogenicity and enabling biofilm production	Luna-Pineda et al., 2019
Iha	tha	Fe-regulated homolog adhesion	
F1C fimbriae	foc	Adhesion to endothelial cells of kidneys and bladder, and epithelial cells of renal	Terlizzi et al., 2017
S fimbriae	sfa	Facilitating the penetration of bacteria into tissues, Adhesion to lower urinary tract, kidney cells along with cells of intestinal epithelial	Behzadi, 2020
P fimbriae	pap	Stimulating the cytokines formation via T lymphocytes, colonization agent in extra-intestinal infection	Pobiega et al., 2013
Dr fimbriae	dru	Mediating internalization bacteria to the host cell, binding to the DAF receptor on the cells of surface epithelial	Lane and Mobley, 2007
			Alvarez-Fraga et al., 2022
Afimbril adhesin	afu	Binding to the DAF receptor on the surface epithelial cells, hemagglutination capacity	
Type 1 fimbriae	fim	As colonization factor in extra-intestinal infection, biofilm production	Nowicki, 2002
Type 3 fimbriae	mrk	As colonization factor, mostly in catheter associated UTIs, biofilm formation	Stark et al., 2016
F9 fimbriae	c	Biofilm formation, mostly pyelonephritis	Wurpel et al., 2013
Auf fimbriae	auf	Biofilm formation	Wurpel et al., 2013
RTX protein ToxA	tosA	Adhesion to upper urinary tract epithelial cells, enhances survival in disseminated infections	Vigil et al., 2011; Xicohtencatl-Cortes et al., 2019
			Kathyayat et al., 2021
Hemagglutinin	hek/hra	Promote autoagglutination, hemagglutination, epithelial cell invasion	

Genomic Surveillance of STEC in Italy

Isolation of STEC strains at ISS (NRL for *E. coli*) or delivery to ISS of STEC isolates



Sequencing at ISS and upload in the ISS webplatform

OR

Upload of STEC sequence from regional centres



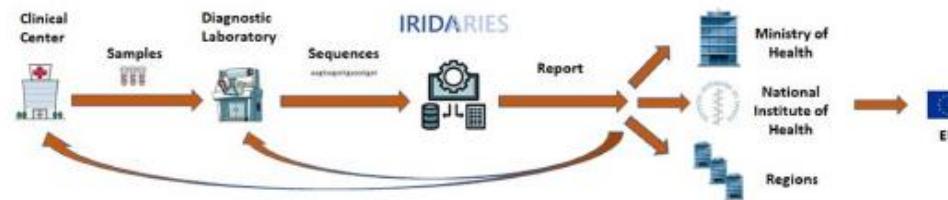
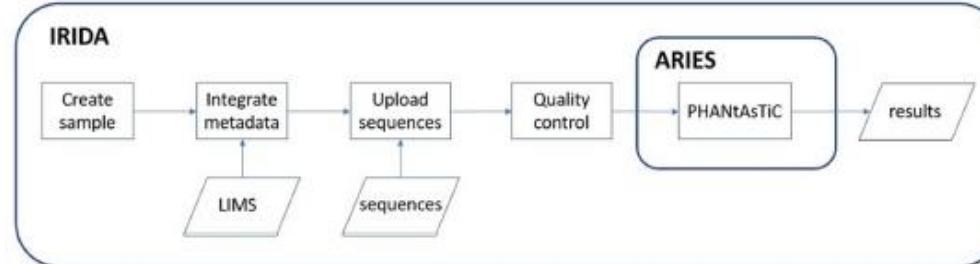
IRIDA



STEC strains characterization and comparison with isolates from different sources



IRIDA-ARIES system



Knjin A. et al. Frontiers in Public Health, 2023

Phantastic Pipeline:

- ✓ Serotyping
- ✓ MLST-7 genes
- ✓ Virulotyping
- ✓ Stx genes subtyping
- ✓ cgMLST



cgMLST analysis among strains of the same serogroup -
In case of cluster identification: Immediate report to
concerned IRIDA-ARIES users and a report describing the
cluster is prepared for further assessment

In February 2024 a genomic cluster of O174 STEC strains was identified

Cluster 394

Consisting in 4 STEC strains *stx1 stx2* from:

- ✓ Bovine faeces (ED1796, ED1798)
- ✓ Cheese samples (ED1802, ED1803)

Isolated from two Vet Institutes in one region in the same period

Looking further in our STEC genomes collection:

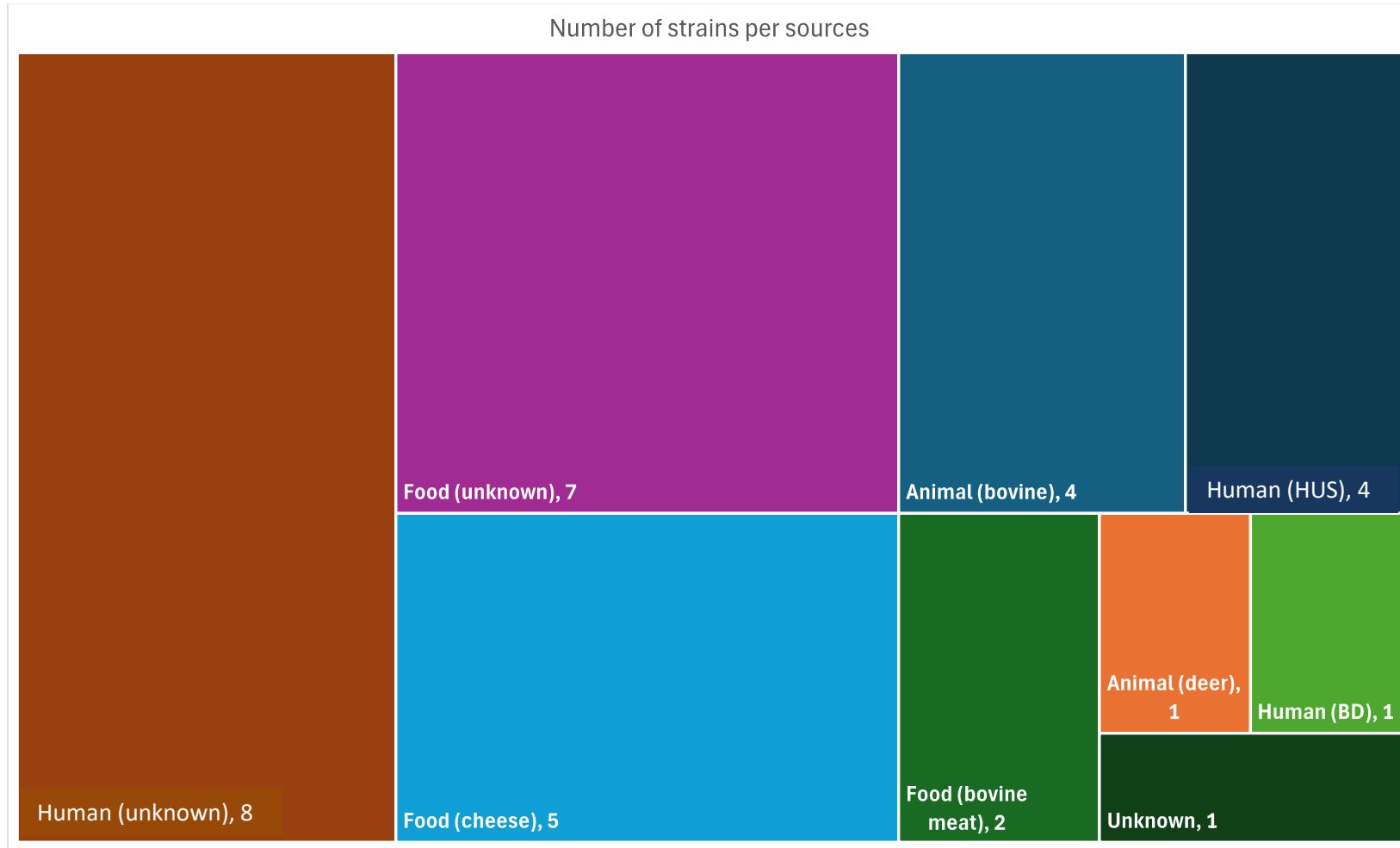
33 STEC strains belonging to O174 serogroup

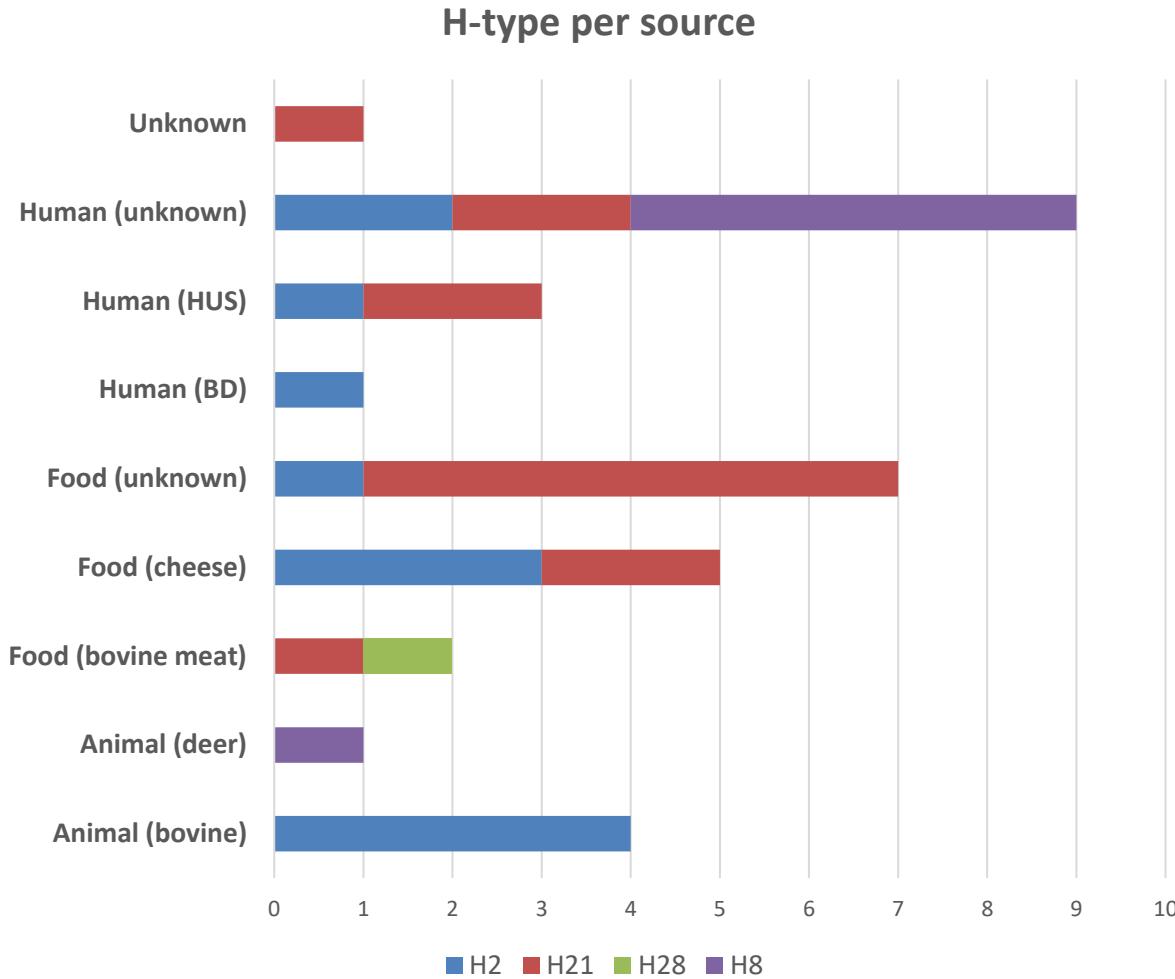
13 from humans

20 from non-human source

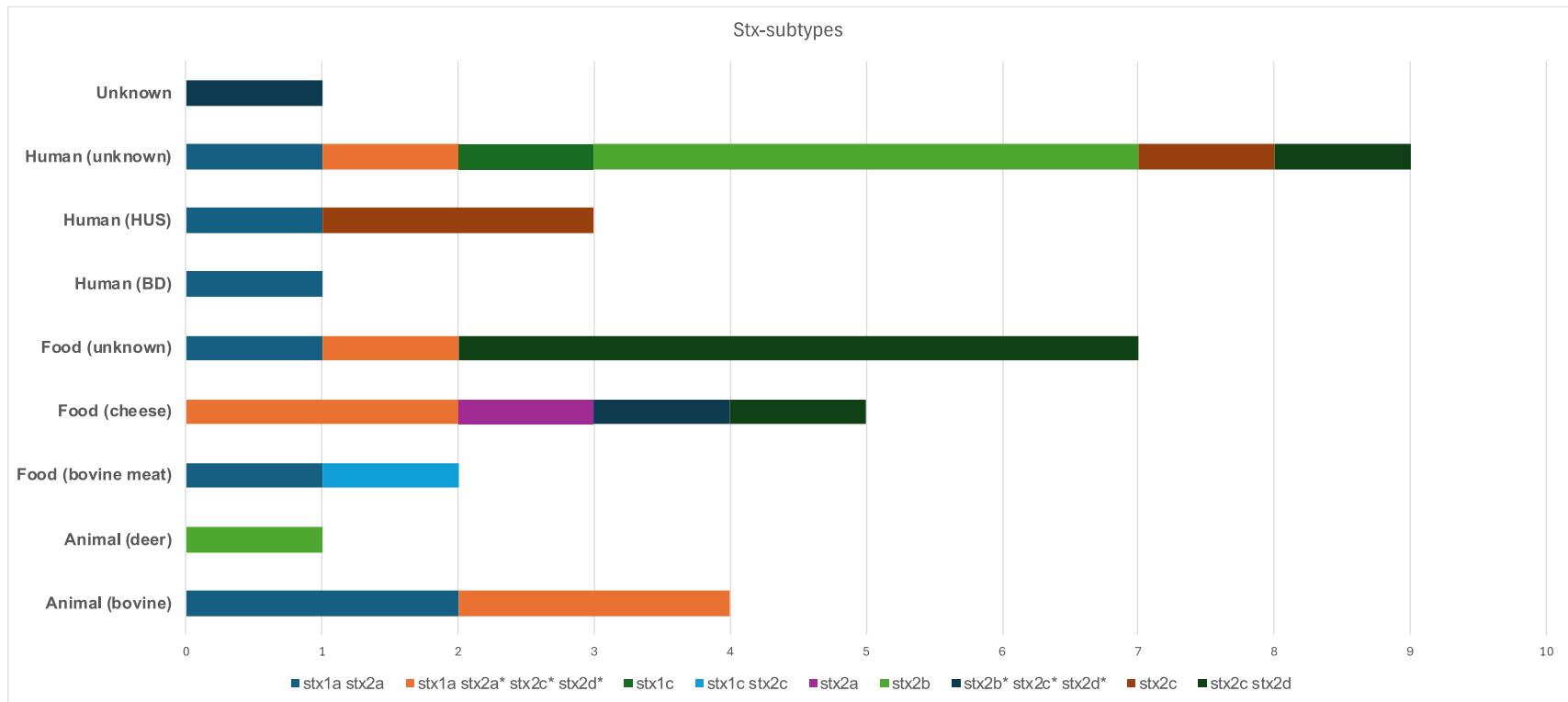
Isolated in the time span 1998-2024

Distribution of O174 strains per source





Stx-coding genes subtypes possessed by O174 strains



Strains possessing ExPEC-UPEC virulence genes

cdt: one strain (food)

sen: 5 strains (4 from humans and 1 from deer)

ompT: 16 strains (4 from humans -1 HUS, 7 from food, 5 animals)

AstA: 3 strains (food)

iss: 32 strains (only missing in one strain from bovine meat)

traT: 31 strains (missing in two strains from humans)

iha: 33 strains

papC: 17 strains (2 strains from HUS, 1 from BD, 3 from humans, 3 from cheese, 4 food, 4 from bovine animals)

afa: 1 strain (non human)

hra: 20 strains (1 from HUS, 1 from BD, 3 from humans, 3 from cheese, 5 food-including 1 from bovine meat, 4 from bovine animals, 1 deer)

ehxA: 23 strains

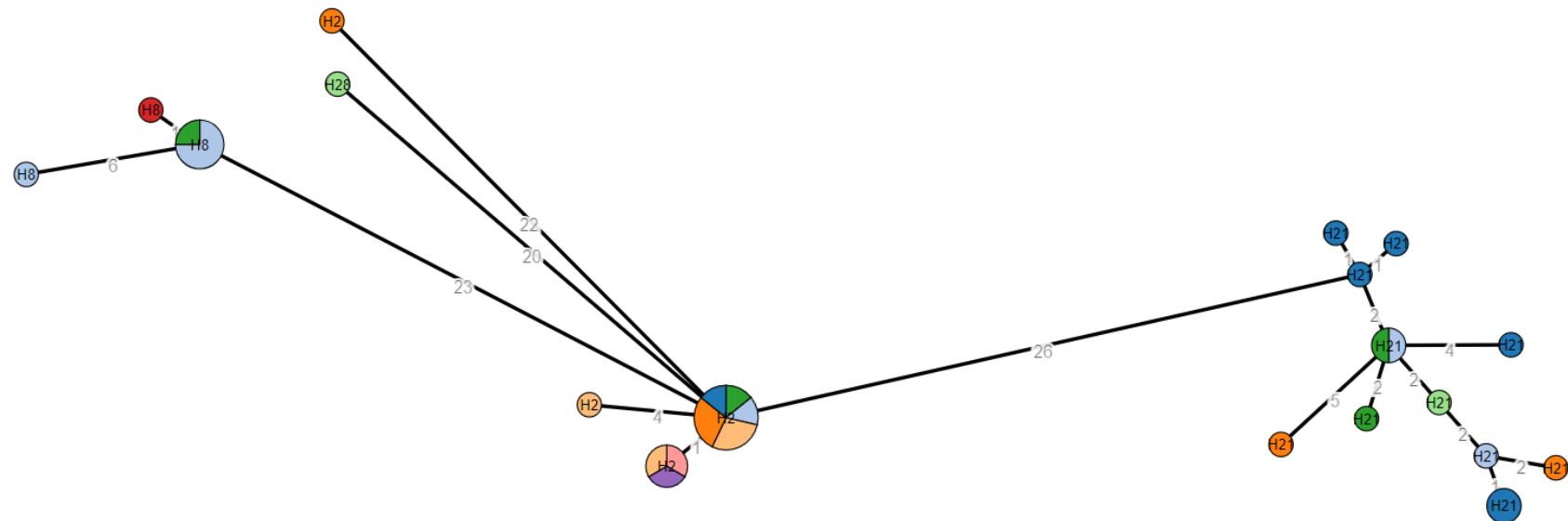
subA: 11 strains

tia: 16 strains

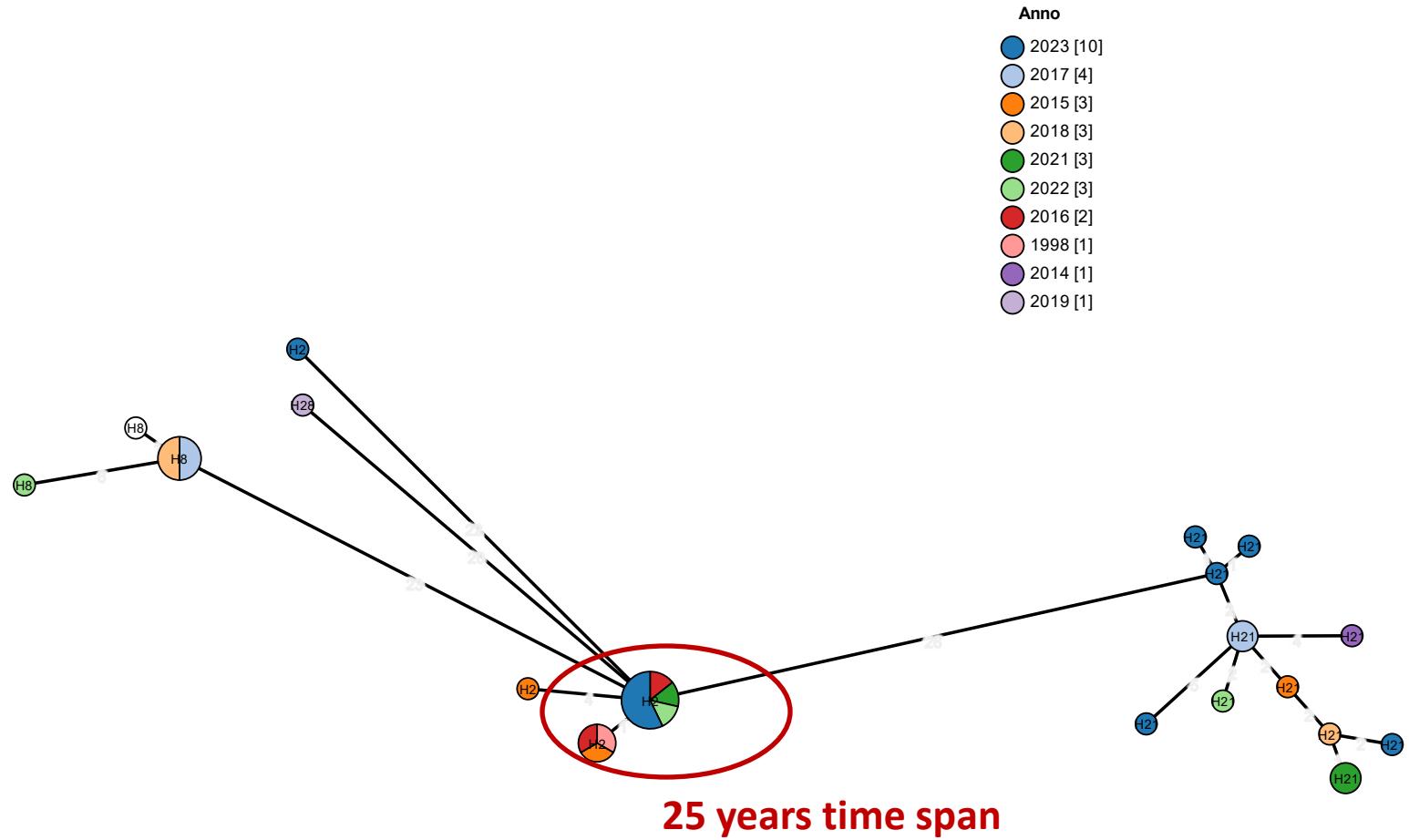
lpfA: 16 strains

cgMLST analysis

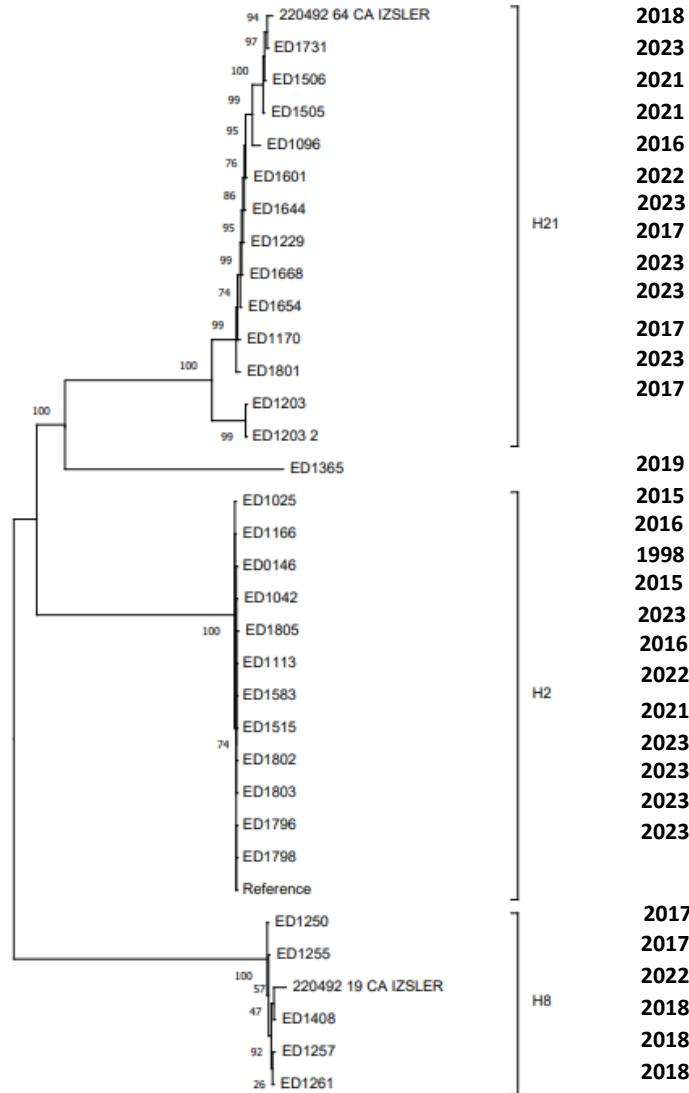
- Food Unk [7]
- Human Unk [7]
- Food Cheese [5]
- Animal, Bovine [4]
- HUS [4]
- Food Bovine [2]
- Animal, Deer [1]
- Human BD [1]
- Human Diarrhea [1]



cgMLST analysis



SNPs analysis (core genome)



Concluding remarks

STEC O174 strains isolated in a time-span 30 years in Italy and seem to be persistent (at least H2)

STEC O174 strains isolated from different sources, including severe disease and food

Different *stx* gene subtypes

They possess mosaic virulotypes

Three phylogenetically related populations can be identified, each one consisting of strains possessing the same *fliC*

All the three groups contain strains from different origin



Zoonotic origin

Thank you for
your attention!

Sample	Source	Year	O-group	H-type	MLST	Virulotype		Cluster ID
						Stx-genes subtypes		
ED0146	Animal (bovine)	1998	O174	H2	ST661	stx1a stx2a	cba, ehxa, espp, gad, hra, iha, iss, lpfa, ompt, papc, saa, stx1a, stx1b, stx2a, stx2b, terc, trat	
ED1025	Animal (bovine)	2015	O174	H2	ST661	stx1a stx2a	cea, ehxa, espp, gad, hra, iha, iss, lpfa, papc, saa, stx1a, stx1b, stx2a, stx2b, terc, trat	
ED1042	Human (unknown)	2015	O174	H2	ST661	stx1a stx2a* stx2c* stx2d*	celb, ehxa, espp, gad, hra, iha, iss, lpfa, papc, saa, stx1a, stx1b, stx2a, stx2b, terc, trat	
ED1096	Food (unknown)	2016	O174	H21	ST677	stx1a stx2a* stx2c* stx2d*	asta, gad, hra, iha, iss, lpfa, stx2a, stx2b, terc, trat	
ED1113	Human (HUS)	2016	O174	H2	ST661	stx1a stx2a	ehxa, espp, gad, hra, iha, iss, lpfa, papc, saa, stx1a, stx1b, stx2a, stx2b, terc, trat	
ED1166	Human (BD)	2016	O174	H2	ST661	stx1a stx2a	celb, ehxa, espp, gad, hra, iha, iss, lpfa, papc, saa, stx1a, stx1b, stx2b, terc, trat	
ED1170	Human (unknown)	2017	O174	H21	ST677	stx2c	espi, gad, iha, iss, lpfa, ompt, papc, stx2b, terc, tia	
ED1203	Food (bovine meat)	2017	O174	H21	ST442	stx1c stx2c	gad, hra, iha, irea, lpfa, stx1a, stx1b, stx2a, stx2b, terc, trat	
ED1229	Human (HUS)	2017	O174	H21	ST677	stx2c	cea, ehxa, espi, espp, gad, iha, iss, lpfa, papc, saa, stx2a, stx2b, suba, terc, tia, trat	
ED1250	Human (unknown)	2017	O174	H8	ST13	stx2b	cba, cea, cia, cma, ehxa, espi, gad, iha, irea, iss, lpfa, senb, stx2a, stx2b, suba, terc, tia, trat	173
ED1255	Human (unknown)	2017	O174	H8	ST13	stx2b	cba, cea, celb, cia, cma, ehxa, espi, gad, iha, irea, iss, lpfa, senb, stx2a, stx2b, suba, terc, tia, trat	
ED1257	Human (unknown)	2018	O174	H8	ST13	stx2b	cba, cea, celb, cia, cma, ehxa, espi, gad, iha, irea, iss, lpfa, senb, stx2a, stx2b, suba, terc, tia, trat	
ED1261	Human (unknown)	2018	O174	H8	ST13	stx2b	cba, cea, cia, cma, ehxa, espi, gad, iha, irea, iss, lpfa, ompt, senb, stx2a, stx2b, suba, terc, tia, trat	173
ED1408	Animal (deer)	2018	O174	H8	ST13	stx2b	cba, cea, cia, cma, ehxa, espi, gad, hra, iha, irea, iss, lpfa, ompt, senb, stx2a, stx2b, terc, tia, trat	
220492-64_CA_IZSLER	Human (unknown)	2018	O174	H21	ST677	stx2c stx2d	gad, hra, iha, iss, lpfa, stx2a, stx2b, terc, tia, trat	
ED1365	Food (bovine meat)	2019	O174	H28	ST?	stx1a stx2a	cdtb, cea, celb, cia, cib, cvac, ehxa, epea, espp, gad, iha, iss, lpfa, ompt, saa, stx1a, stx1b, stx2a, stx2b, suba, terc, trat	
ED1515	Food (unknown)	2021	O174	H2	ST661	stx1a stx2a	ehxa, espp, gad, hra, iha, iss, lpfa, papc, saa, stx1a, stx1b, stx2a, stx2b, terc, trat	
ED1506	Food (unknown)	2021	O174	H21	ST677	stx2c stx2d	asta, gad, hra, iha, iss, lpfa, stx2a, stx2b, terc, tia, trat	58
ED1505	Food (unknown)	2021	O174	H21	ST677	stx2c stx2d	asta, gad, hra, iha, iss, lpfa, stx2a, stx2b, terc, tia, trat	58
220492-19_CA_IZSLER	Human (unknown)	2022	O174	H8	ST13	stx1c	celb, espi, gad, iha, irea, iss, lpfa, ompt, stx1a, stx1b, suba, terc, tia	
ED1601	Human (HUS)	2022	O174	H21	ST677	stx2c	cea, cia, cib, epea, espi, gad, iha, iss, lpfa, ompt, stx2a, stx2b, terc, tia, trat	
ED1583	Human (unknown)	2022	O174	H2	ST661	stx1a stx2a	ehxa, espp, gad, hra, iha, iss, lpfa, papc, saa, stx1a, stx1b, stx2a, stx2b, terc, trat	
ED1644	Food (unknown)	2023	O174	H21	ST677	stx2c stx2d	ehxa, espi, espp, gad, iha, iss, lpfa, ompt, papc, saa, stx2a, stx2b, suba, terc, tia, trat	
ED1654	Food (unknown)	2023	O174	H21	ST677	stx2c stx2d	ehxa, espi, espp, gad, iha, iss, lpfa, papc, saa, stx2a, stx2b, suba, terc, tia, trat	