



Final report PT-07: AnMol 1/2024

PT-07: <u>"Molecular identification of Anisakid nematodes</u> <u>at the species level"</u>

Design

Purpose	Evaluation of laboratories competence in molecular identification of anisakidae nematodes species			
Scheme type	Single, simultaneous			
Participants	National reference laboratories for parasites. Public and private laboratories			
N. of participants	Depending on request			
Method	not regulated			
Test method	chosen by the participant			
	Matrix	fresh water farmed fish fillet		
PT items	Item	anisakid nematodes (DNAs or larvae fragments)		
	N. of samples	4 vials for each participant		
	Distribution Preparation and packaging can be performed bef shipment			
Subcontracted activities	NA			
Results evaluation	Qualitative			

Implementation

N. of participants	15		DNA	30
Public laboratories	/		Larvae fragments	30
Private laboratories	/	PT items	PT panel composition	2 items with a single species DNA each (<i>Contracoecum osculatum</i>), 2 items with a single larva fragment each (<i>P. decipiens sl and A. pegreffii</i>)
NRL	15		Shipping	DHL

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Istituto Superiore di Sanità

Department of Infectious Diseases Unit of Foodborne and Neglected Parasitic Diseases European Union Reference Laboratory for Parasites



Shipping dates

11/03/2024

Results

The PT final evaluation was qualitative only. The PT was considered passed if all species were correctly identified by the participant.

Laboratory code	N° of samples correctly identified	N° of samples NOT correctly identified	Method applied	Final evaluation	
AMM01	4	0	In house method;EURLP method 1 (PCR_RFLP);	Positive	
AMM02	4	0	EURLP method 2 (multiplex PCR);	Positive	
AMM03	2	2	EURLP method 2 (multiplex PCR);In house method; (PCR- RFLP)	Negative	
AMM04	4	0	Published method;	Positive	
AMM05	4	0	EURLP method 2 (multiplex PCR);	Positive	
AMM06	4	0	EURLP method 1 (PCR_RFLP);	Positive	
AMM07	4	0	EURLP method 2 (multiplex PCR);EURLP method 1 (PCR_RFLP);In house method;	Positive	
AMM08	4	0	EURLP method 1 (PCR_RFLP);	Positive	
AMM09	3	1	Published method;	Negative	
AMM10	4	0	EURLP method 1 (PCR_RFLP);	Positive	
AMM11	4	0	EURLP method 2 (multiplex PCR);	Positive	
AMM12	4	0	EURLP method 1 (PCR_RFLP);EURLP method 2 (multiplex PCR);	Positive	
AMM13	4	0	EURLP method 1 (PCR_RFLP);	Positive	
AMM14	4	0	EURLP method 2 (multiplex PCR);	Positive	
AMM15	4	0	EURLP method 1 (PCR_RFLP);	Positive	

Legend:

• Laboratories that failed the PT are marked in bold and highlighted in grey.

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Summary of results:

Total number of PT panels	60	
Number of participant laboratories	15	
Number of participants that passed the PT	13	
Number of participants that failed the PT	2	

Laboratory code	2017	2018	2019	2020	2021	2022	2023	2024
AMM01	Active of a press			Р	Р	Р	N	Р
AMM02	P	Р	N	Р	Р	Р	Р	Р
AMM03	Р	Р	Р	Р	Р	Р	P	N
AMM04			Р		Р	Р	P	Р
AMM05	Aren			NA	Р	Р	P	Р
AMM06	Р	Р	Ρ	Р	Р	Р	1. A. A.	Р
AMM07		Р		N	Р	N	N	Р
AMM08	Р	Р	Р	N	Р	Р	N	Р
AMM09			N	the weather the	Р	N	N	N
AMM10	P	Р	Р	Р	Р	P	Р	Р
AMM11	P	Р	Р	Р	Р	P	Р	Р
AMM12	P	Р	P	Р	Р	P	Р	Р
AMM13		AR - CALL		同時設備的構成	NA	Р	N	P
AMM14	東京大部分会社	Р		alight a caller			N	Р
AMM15	Р	NA		AND INTE CONTRACT	Р	P	N	Р

Overtime comparison of results

Note: P, positive; N, negative; NA, no result received; grey box, no participation

Comments:

In the PT round of 2024, all but one laboratories, received the PT items within 72 hours. Thirteen participants successfully accomplished the PT, whereas two laboratories (13%) failed. In particularly, one laboratory (AMM03) did not correctly identify both DNA samples of *Contracaecum osculatum* (they were reported as *A. typical*), whereas the other (AMM09) did not correctly identify the fragment larva of *A. pegreffii* (it was reported as *A. simplex*).

The reasons for the PT failure were analyzed and reported to the participants. AMM03 reported difficulties in sample amplification and related this to the low amount of DNA in samples. However, the same batch of DNAs were forwarded to all PT participants and no problem about amplification was reported by other participants, independently on the method applied. No problem with items storage during the shipping could be also envisaged, according to temperature check reported by the participant. Based on the information provided in the Result form and additional information provided from the participant by email, it was recommended to optimize both in-house method as well as the EURLP-multiplex PCR (e.g. test alternative Taq polymerse).

AMM09 used a published method that appears to have limitations in detecting several Anisakidae species being specific for *C.osculatum* but not for other species. It is unclear if additional method(s) were used for typing. The laboratory was invited to further improve the in-house method as well as to perform it in parallel

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with recommended EURLP methods. Over the years, this laboratory have failed four out of five times the PT (table "overtime comparison of the results").

Concerning the applied molecular method(s) (Figure 1): 5 laboratories applied only the PCR-RFLP method (EURLP 1; MI04); 5 used only the multiplex-PCR (EURLP 2; MI10); 2 used both methods; 3 used in house/published methods (one of which was PCR of the EURLP 1 followed by Sanger sequencing instead of RFLP).

Compared to the previous years the number of participant laboratories slightly increased. The number of laboratories that failed the 2024 PT round low (13%) in comparison to previous years (Figure 2), indicating that efforts were done to improve the correct application of the molecular identification tests in place.

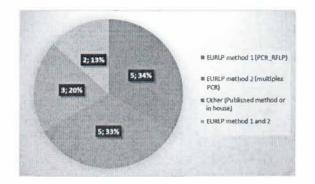
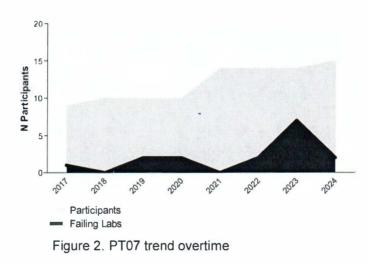


Figure 1. Percentage of Method applied for the PT2024



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Verified and issued by

The Director Dr. A. Casulli

Written and elaborated by

PTP person in charge M.L alle

Date 22-05-2024

Notes:

- To guarantee confidentiality, participant laboratories are identified by alphanumeric codes. PT participant identity is kept confidential and bound by professional secrecy. The PTP reserves itself the right to provide the laboratory PT result to the competent authority on request.
- 2. The organizer designates a qualified company for the transport and delivery of PT items.
- 3. Each participating laboratory receives a PT panel according to the PT scheme. Each PT item consists of a fish fillet sandwich spiked or not with live Anisakidae larvae. The homogeneity of PT items is ensured by an accurate control of the number of larvae spiked into each sample (item) made by two operators. PT items are stable for 7 days from the date of preparation (corresponding to the shipping date), provided that they are maintained in suitable conditions.
- 4. At the beginning of each year, the organizer draws up a PT program and makes it known by sending an email to the NRLs
- 5. The final report issue of each PT round shows the PT program implementation.

End of the report

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