17<sup>th</sup> Annual Workshop of the National Reference Laboratories for *E. coli* Rome 10-11 October 2022

### **PT30**

# Detection of Shiga toxin-producing *E. coli* (STEC) in sprout spent irrigation water







### **Objectives and Design of the study**

The objective of PT30 consisted in the examination of artificially contaminated sprout spent irrigation water expanding the observations related with the testing of sprout spent irrigation water

The participating Labs were requested to carry out the pretreatment procedure developed by the EURL for *E. coli* and then apply ISO TS 13136:2012 for detecting the presence of STEC in sprout irrigation water, carrying out the enrichment at 41.5°C

- Detection of the main STEC virulence genes (*eae* and *stx* genes)
- Isolation and characterization of STEC strains

#### The voluntary inter-laboratory study (PT30) was organized and run in October 2021







### **PT30: Participants**







### **Test Samples**

The spent irrigation water used in the study was obtained from a local sprout producer who collected the irrigation water after 48h from the beginning of radish sprout production process

The presence of a natural background microflora has been evaluated  $\rightarrow$  4x10<sup>5</sup> CFU/ml Background microflora

➡ Water samples were negative at the Real Time PCR screening for the gene targets of STEC according to the ISO TS 13136:2012

Beside the presence of the bacterial background microflora, the water resulted heavily contaminated with protozoa







### **Stability tests:**

- Tests carried out on spent irrigation water of the same nature but collected during another cycle of sprout production
- Tests carried out using different levels of contaminations
- the STEC O157 strain could be isolated from the 50 CFU/ml spiking level after seven days from contamination.

### **PT30: Samples**

	Contamination level in:			
Contaminant (Genotype)	Sample 1	Sample 2		
C210-03 STEC O157 (stx1+, stx2+, eae+)	50 CFU/ml	-		





#### **PT30: Samples**

	Contamination level in:			
Contaminant (Genotype)	Sample 1	Sample 2		
C210-03 STEC O157 (stx1+, stx2+, eae+)	50 CFU/ml	-		

- The homogeneity test was performed on 6 randomly selected samples for each of the two sample types
- Samples labelled with randomly generated numerical codes were shipped on the 4th of October 2021
- ✓ Results submitted through an on-line form from 37 labs





Screening step: Proportion of Laboratories correctly detecting the presence or absence of virulence and serogroup-associated genes in the enrichment cultures



Two out of the three labs detecting the presence of STEC in sample 1 could isolate the STEC O157 *eae+ stx1+ stx2+* contaminating strain

#### **Concluding remarks**

- PT30 was not meant to assess the proficiency of the laboratories but it was rather a collaborative study to verify the appropriateness of the procedure for the pre-treatment and analysis of spent irrigation water samples for the presence of STEC
- This study confirmed the complexity of testing the sprout spent irrigation water and highlighted the necessity to refine the pre-treatment procedure
- The preliminary tests showed the water samples contained a large number of freeliving amoebae
- All the information coming from this study will be used to improve the testing process for spent irrigation water

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### **PT33**

#### Detection of Diarrhoeagenic *E. coli*, including STEC in cheese







### **Objectives and Design of the study**

The objective of PT33 consisted in the examination of artificially contaminated cheese samples, in order to enhance the preparedness of NRLs in testing cheese for the presence of Diarrhoeagenic *E. coli*, including STEC.

- The detection of the main virulence genes associated with pathogenic *E.coli*
- The isolation and characterization of the strains

#### The inter-laboratory study (PT33) was organized and run in June 2022







### **PT33: Participants**

#### 26 NRLs representing 22 EU MS

#### + 5 Non-EU NRLs of

Egypy Iceland Norway Switzerland UK (3 Labs)



![](_page_10_Picture_5.jpeg)

![](_page_10_Picture_7.jpeg)

### **Test Samples**

The cheese used in the study was purchased from a local retailer

The presence of a natural background microflora has been evaluated by plating on two different media (TSA and MacConkey agar) serial dilutions of 25 g of cheese in BPW  $\implies$  No growth was observed

> Two samples consisting of 25 g of cheese have been assayed for the presence of STEC and EAEC  $\implies$  Both samples were negative at the Real Time PCR screening for the gene targets of STEC and EAEC

![](_page_11_Picture_4.jpeg)

![](_page_11_Picture_6.jpeg)

### Stability tests (May 2022):

EAEC O104 Concentration	1	T0 T1 (3 days)		T2 (7	days)	T3 (10 days)			
Test	RT-PCR	Isolation	RT-PCR	Isolation	RT-PCR	Isolation	RT-PCR	Isolation	
2 CFU/g	+	+	+	+	+	+	+	+	
STEC 078 Concentration	T0 T1		T1 (3	T1 (3 days)		T2 (7 days)		T3 (10 days)	
Test	RT-PCR	Isolation	RT-PCR	Isolation	RT-PCR	Isolation	RT-PCR	Isolation	
2 CFU/g	+	+	+	+	+	+	+	+	

#### **PT33: Samples**

Cheese Samples	Contamination			
Sample 1	Negative			
Sample 2	2 CFU/g EAEC O104 aggR, aaiC*			
Sample 3	2 CFU/g STEC O78 stx1**			

![](_page_12_Picture_4.jpeg)

![](_page_12_Picture_6.jpeg)

#### **PT33: Samples**

Cheese Samples	Contamination				
Sample 1	Negative				
Sample 2	2 CFU/g EAEC O104 aggR, aaiC*				
Sample 3	2 CFU/g STEC O78 stx1**				

- ✓ Cheese Samples were contaminated on the 24<sup>th</sup> of June 2022
- The homogeneity test was performed on 6 randomly selected samples for each of the three cheese sample types
- ✓ Samples labelled with randomly generated numerical codes were shipped on the 27<sup>th</sup> of June 2022
- ✓ Results submitted through an on-line form from 33 NRLs

![](_page_13_Picture_6.jpeg)

![](_page_13_Picture_8.jpeg)

#### Real-time PCR detection of virulence genes in the enrichment cultures from EU NRLs

![](_page_14_Figure_1.jpeg)

### Isolation and genotyping of STEC and EAEC strains from cheese samples

Sample 1		Sample 2		Sample 3
NEGATIVE		aggR+ aaiC+ 0104		stx1+ stx2- eae- 078
L144	L144		L144	ONT
L222	L222		L222	0128
L226	L226		L226	ONT
L230	L230	ONT	L230	ONT
L256	L256	ONT	L256	0178
L258	L258		L258	ONT 🔨
L327	L327		L327	ONT
L337	L337		L337	ONT
L358	L358		L358	ONT
L370	L370		L370	ONT
L403	L403		L403	
L407	L407		L407	ONT
L462	L462		L462	ONT
L522	L522		L522	ONT
L615	L615		L615	ONT
L640	L640		L640	ONT
L685	L685		L685	ONT
L697	L697		L697	ONT
L705	L705		L705	ONT
L708	L708		L708	ONT
L758	L758		L758	ONT
L846	L846		L846	ONT
L896	L896	ONT	L896	ONT
L976	L976		L976	ONT
L985	L985		L985	ONT
L986	L986		L986	ONT

All the participants were able to isolate the contaminating STEC and EAEC strains. Most of the labs did not detect the O78 serogroup in the isolated STEC strain

# Real-time PCR detection of virulence genes in the enrichment cultures from Non-EU NRLs

![](_page_16_Figure_1.jpeg)

# Isolation and genotyping of STEC and EAEC strains from cheese samples

	Sample 1		Sample 2		Sample 3
	NEGATIVE		aggR+ aaiC+ 0104		stx1+ stx2- eae- 078
L563		L563	ONT aggR+	L563	ONT
L635		L635	ONT	L635	ONT
L674		L674		L674	0104
L972		L972	ONT	L972	NEGATIVE
L982		L982		L982	
L983		L983	NEGATIVE	L983	ONT
L984		L984	NEGATIVE	L984	

#### Evaluation of the NRLs performance in the Real Time PCR screening step:

- **4 penalty points** to each incorrect or missing result concerning the identification of the *stx1* and *stx2* genes
- **2 penalty points** to each result incorrectly reporting the presence of *eae* gene
- **2 penalty points** to each incorrect or missing result concerning the identification of the *aggR* and *aaiC* genes

#### **Evaluation of the NRLs performance in the isolation of STEC** and EAEC strains:

- **2 penalty points** to the lack of isolation of EAEC from sample 2 and of STEC from sample 3 or to the isolation from sample 1
- **4 penalty points** to each incorrect or missing result concerning the identification of the *stx1* and *stx2* genes
- **2 penalty points** to each result incorrectly reporting the presence of *eae* gene
- **2 penalty points** to each incorrect or missing result concerning the identification of the *aggR* and *aaiC* genes

![](_page_17_Figure_9.jpeg)

![](_page_17_Picture_10.jpeg)

#### Evaluation of the NRLs performance in the isolation of STEC and EAEC strains:

![](_page_18_Picture_1.jpeg)

- **2 penalty points** to each incorrect result concerning the identification of O78 serogroup of the STEC strain
- **No penalty points** were assigned for reporting the serogroup of the isolated STEC strain as ONT (the O78 serogroup is not in the list of the top-14 serogroups)
- The serogroup of the EAEC strain was not the subject of the performance assessment

#### **Evaluation of the EU NRLs performance in the PT procedures**

#### (screening + isolation steps)

The labs that scored higher than 8 were considered under-performant

All the labs complied the definition of satisfactory proficiency

![](_page_19_Picture_4.jpeg)

# Evaluation of the Non-EU NRLs performance in the PT procedures (screening + isolation steps)

![](_page_20_Figure_1.jpeg)

#### All the labs complied the definition of satisfactory proficiency

#### **Concluding remarks**

- A high participation was recorded for PT33. The level of participation observed confirmed the eagerness and collaboration of the network.
- No NRL was considered under-performant. The capability of the NRLs in identifying the STEC and other pathogenic *E. coli* virulence genes is highly satisfactory.
- The identification of the O78 serogroup was problematic and achieved only by three participants (one EU NRL and two Non-EU NRLs).
- The O78 serogroup is not comprised in the top-14 serogroups and thus has not been used to assess the proficiency of the laboratories in the determination of this feature.

Thanks to all the participants in the study and thank you all for your attention!