

NATIONAL REFERENCE CENTRE FOR ANISAKIASIS (C.Re.N.A.)- ITALY: 2020-2021 activity

Dr. Gaetano Cammilleri

NRL for Anisakiasis, Istituto Zooprofilattico Sperimentale della Sicilia, Palermo, Italy The National Reference Centre for Anisakiasis (C.Re.N.A.) has been established at the Istituto Zooprofilattico Sperimentale of Sicily by a Health Ministry decree on August 27 2004 (G.U. N° 43 on February 22 2005) and designated NRL for Anisakiasis since November 14 2006.

Main objectives

- Development and standardization of diagnostic methods
- Confirmation of diagnosis from other laboratories
- Organization, in collaboration with the ISS, of periodic "ring tests" among Italian IZSs
- Provision of reference material
- Research activities funded by Italian Ministry of Health
- Training courses for staff of other IZSs

And also...

- Network with human health centers that can detect anisakiasis to collect epidemiological data (human cases and allergic manifestation)
- Study of allergenic potential of Anisakis
- Risk analysis

2020 activity

Inspection method

- Visual inspection
- Artificial digestion

Anisakidae identification

- Optical microscopy
- PCR-RFLP
- Multiplex PCR
- Real-time PCR assay

Specie	N. samples	N. samples infested	N. larvae	Visual inspection	Chloropeptic digestion
Engraulis encrasicolus	3373	586	586	586	-
Lepidopus caudatus	60	60	4888	60	-
Sardina pilchardus	3566	96	96	92	4
Scomber scombrus	269	87	671	72	15
Trachurus trachurus	275	133	1931	124	9



PREVALENCE OF INFESTATION



MEAN INTENSITY







Molecular analysis

Seasonal trend



Seasonal trend





Natural Product Research >

Formerly Natural Product Letters Volume 34, 2020 - Issue 1: Ethnomedicine and Foods sources: novel perspectives. Guest Editor: Nicola Cicero

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Altmetric

Short Communication Seasonal trend of Anisakidae infestation in South Mediterranean bluefish

Gaetano Cammilleri 🔄, Andrea Pulvirenti, Antonella Costa, Stefania Graci, Rosaria Collura, Maria Drussilla Buscemi,show all Pages 158-161 | Received 29 Oct 2018,Accepted 15 Jan 2019,Published online: 20 Feb 2019

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2021 Activity

Inspection method

- Visual inspection
- Artificial digestion

Anisakidae identification

- Optical microscopy
- PCR-RFLP
- Multiplex PCR
- Real-time PCR assay



Molecular analysis

- 83,3% *A. simplex s.s.*
- 16,7% A. pegreffii



Organization of Proficiency tests

Proficiency test visual method



Proficiency test PCR-RFLP method



C.Re.N.A. organized three proficiency tests in November 2020 and 2021:

- Qualitative detection (absence-presence) of *Anisakis* larvae in fish fillets (visual method)
- Detection for Anisakis
 larvae by artificial
 digestion method ISO
 23036-2/2021: dispatch n.
 3 fish fillet samples
- Identification of Anisakis larvae by PCR-RFLP: dispatch n. 3 DNA samples

The graphs show the number of participants per year



Validation of ISO 23036-2:2021



- Detection of L3 Anisakidae larvae in fish products.
- Fresh or processed
- Chloro-peptic digestion of muscle and viscera
- Determination of the vitality (37 °C)



POS CRENA 04 Comparison

POS CRENA 04

2 L water46-48 °C
Start weight 100 g
16 mL HCl 25%
vortexing
10g of pepsin (1:10.000 NF)
44 - 46°C
Filtering after 30 - 45 min



ISO 23036-2:2021
2 L of water 37± 2°C
□Start weight 25-200g
□16 mL HCl 25%
□vortexing
□10g of pepsin (1:10.000 NF)
□ 37°C
□Filtering after 45 min

Optimization of the protocol

□ temperature: 37°C (alive larvae) for 15-30′ (no more than 45′) □ weight 25-200 g

Digestion fluid: HCl 16 ml in 2 l water, pepsin (30 ml), 10 g for powder.

Over 30 trials at different conditions

- Farmed Sea bream muscle
- Superior part / ventral part
- epiaxial muscles
- Tail muscles

Optimal conditions

- 2 L of water
- 11 ml of HCl≥35%
- 10 g pepsin 1:10.000
- 50±5 g of sample
- Digestion time 40 min
- Temperature 36±1°C



Validation of the method



Larvae vitality: acid solution up to 30 min from the digestion

Motility examination by stereomicroscope









Undigested samples

Mean of 1,19 grams



LOOP-MEDIATED ISOTHERMAL AMPLIFICATION (LAMP)



LAMP assay: Primer design



Anisakis sequences reported in gene bank for the primers design



Lehgth between primeds, GC content, melting temperature, absence of complementarity. secondary structures. TTR



A set of 6 primers (2 outer (F3 and B3), 2 inner (FIP and BIP) and two loop (LF and LB)) using the Primer Explorer software

PrimerExplorer V4

Software

LAMP : DNA extraction





Real-time monitoring of time and temperature





No amplification was found specificity =100%.

No amplification was found with samples spiked with DNA and larvae of *Contracaecum* sp. *Pseudoterranova* sp. and *Hysterothylacium* sp.

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Kit	Controlfood Anisakis Glow	CAG16158 160				
Name	prova specificità	140				
Operator	Gaetano Cammilleri	120				
Instrument	mini 160044	100				
	+ k+	80		/		
	+ k+	60		/		
	— k-	40		/		
	- k-	20				
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THE OWNER AND ADDRESS	- 6					



 Sensibility 100% for every sample examined – Amplification for A. simplex s.s., A. pegreffii, A. physeteris, A. ziphidarum and A. typica DNA.





Anisakis spp. DNA detected with a dilution up to 10⁻⁴ (0.00022 ng µl⁻¹), with amplification of all the replicates.







Article

Validation of a Commercial Loop-Mediated Isothermal Amplification (LAMP) Assay for the Rapid Detection of *Anisakis* spp. DNA in Processed Fish Products

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Another publication in 2020

ORIGINAL ARTICLE

EUR ANN ALLERGY CLIN IMMUNOL

Vol. 52, N.3, 131-141, 2020

I. BRUSCA¹, S. GRACI², M. BARRALE¹, G. CAMMILLERI², M. ZARCONE³, R. ONIDA¹, A. COSTA², V. FERRANTELLI², M.D. BUSCEMI², C.G. UASUF⁴, M. GJOMARKAJ⁴, M. VAZZANA⁵, S.M. LA CHIUSA¹, G. IACOLINO¹, F. VITALE³, W. MAZZUCCO³

Use of a comprehensive diagnostic algorithm for Anisakis allergy in a high seroprevalence Mediterranean setting

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