

Metal ions and oxidative protein modification in neurological disease

Lawrence M. SAYRE (a), Paula I. MOREIRA (b), Mark A. SMITH (b) and George PERRY (b)

(a) Department of Chemistry;

(b) Department of Pathology, Case Western Reserve University, Cleveland (OH), USA

Summary. - This review highlights the role of oxidative stress and imbalances in metal ion homeostasis in the neurodegenerative diseases Alzheimer's disease and Parkinson's disease and in the progressive demyelinating disease multiple sclerosis. The chemistry and biochemistry of oxidative stress-induced protein damage are first described, followed by the evidence for a pathological role of oxidative stress in these disease states. It is tempting to speculate that free radical oxygen chemistry contributes to pathogenesis in all these conditions, though it is as yet undetermined what types of oxidative changes occur early in the disease, and what types are secondary manifestations of neuronal degeneration.

Key words: neurodegeneration, Alzheimer's disease, Parkinson's disease, multiple sclerosis, metals, oxidative stress.

Riassunto (*Ioni metallici e ossidazione proteica nella patologia neurologica*). - Questa rassegna evidenzia il ruolo dello stress ossidativo e l'alterazione dell'omeostasi di ioni metallici nelle malattie neurodegenerative e da demielinizzazione progressiva (Alzheimer, Parkinson e sclerosi multipla). Vengono descritte in primo luogo la chimica e la biochimica del danno proteico indotto da stress ossidativo, seguite dall'evidenza del ruolo patologico dello stress ossidativo in queste malattie. Si tenta di valutare il contributo dei radicali liberi dell'ossigeno alla patogenesi di tali disordini, sebbene non sia ancora chiaro quali tipi di cambiamenti ossidativi avvengano nei primi stadi e quali costituiscano manifestazioni secondarie della degenerazione neuronale.

Parole chiave: neurodegenerazione, malattia di Alzheimer, morbo di Parkinson, sclerosi multipla, metalli, stress ossidativo.

Introduction

Recent advances in molecular genetics have furthered knowledge of the hereditary basis of familial forms of Alzheimer's disease (AD) and Parkinson's disease (PD). The spontaneously occurring forms of these age-related neurodegenerative diseases, as well as the progressive autoimmune demyelinating disease multiple sclerosis (MS), are of unknown origin, but are thought to reflect a complex combination of hereditary, environmental, and lifestyle factors. In AD and PD, there is compelling evidence for a role of a metabolic imbalance and resulting oxidative stress that is superimposed on hereditary factors, and likely play a larger role in the spontaneously occurring forms of these two diseases [1, 2]. The link between oxidative stress and neuronal death is complex, but there is support for a contributory role of oxidative stress-induced neurotoxicity in both diseases [3]. The fluctuating aspect of MS between periods of

exacerbation and remission would suggest that this disease has little in common with age-related neurodegenerative diseases. However, there is growing awareness that the progression of MS is associated with Wallerian degeneration ("dying back" of axons from terminals), and oxidative stress may play a pathological role here as well.

The condition of oxidative stress in each of these disease states is accompanied to a varying degree by a dyshomeostasis of metal ions, including the redox-active transition metals iron and copper as well as redox inactive metal ions such as zinc. The central nervous system is particularly vulnerable to oxidative stress on account of the high rate of dioxygen utilization, the relatively poor concentrations of antioxidants and related enzymes, and the high content of polyunsaturated lipids, the most vulnerable biomacromolecule to oxidation. There is also an accumulation of iron in the brain as a function of age, which can be a potent catalyst for oxidative species formation.

A characteristic hallmark of AD and PD is the deposition of abnormal forms of specific proteins in the brain, which are immunoreactive to antibodies recognizing protein side-chains modified either directly by reactive oxygen or nitrogen species, or by products of lipid peroxidation or glycooxidation. Although the source(s) of oxidative damage are not entirely clear, this damage may reflect the findings of increased localization of redox-active transition metals in the brain regions most affected. This review describes evidence that suggests a role of oxidative stress and/or metal imbalances in these neurodegenerative conditions as well as in MS.

Definition of oxidative stress

Oxidative stress is defined as the imbalance between biochemical processes leading to production of reactive oxygen species (ROS) and those responsible for the removal of ROS, the so-called cellular antioxidant cascade. Tissues that become subject to oxidative stress witness steady state levels of ROS-mediated damage to all biomacromolecules (polynucleotides, proteins, lipids, and sugars) that can lead to a critical failure of biological functions and ultimately cell death. Several neurodegenerative disorders are associated with oxidative stress that is manifested by lipid peroxidation, protein oxidation and other markers.

Mitochondria are essential organelles for neuronal function because the high-energy requirement makes them highly dependent on aerobic oxidative phosphorylation. However, oxidative phosphorylation is a major source of ROS. All aerobic organisms produce at least minimal levels of ROS, mostly arising from the side-production of superoxide by reaction of molecular oxygen with sites in the electron-transport chain where reducing equivalents accumulate. Superoxide, which is also generated from the respiratory burst of neutrophils, can be subsequently transformed to the other classical ROS species H_2O_2 and hydroxyl radical. ROS can also arise from mutationally-altered or damaged metallo-enzymes involved in oxidative metabolism. Reactive species generated by mitochondria have several cellular targets including mitochondrial components themselves (lipids, proteins and DNA). The lack of histones in mitochondrial DNA (mtDNA) and diminished capacity for DNA repair render mitochondria an easy target to oxidative stress events. Mitochondrial dysfunction and free radical-induced oxidative damage have been implicated in the pathogenesis of PD and AD, as well as other neurodegenerative disorders.

Usually considered as the chief instigator of oxidative stress damage, the hydroxyl radical reacts non-discriminately with all biomacromolecules at

diffusion-controlled rates, i.e., within nm distances from its site of generation. Hydroxyl radical can be produced by gamma radiation, but is most commonly generated physiologically by the Fenton reaction between reduced transition metals (usually iron(II) or copper(I)) and H_2O_2 . Re-reduction of the resulting oxidized transition metal ions (iron(III) or copper(II)) can be effected by cellular reductants such as vitamin C or thiols. In contrast to hydroxyl radical, superoxide radical is chemically unreactive, except at lower pH, where it exists as the hydroperoxy radical. However, superoxide can serve as the reductant of oxidized metal ions for the production of hydroxyl radical from H_2O_2 , the so-called Haber-Weiss reaction. Under normal conditions, damage by ROS is kept in check by an efficient antioxidant cascade, including both enzymatic and non-enzymatic entities. Important in the former regard are cytosolic copper-zinc superoxide dismutase (CuZnSOD) and mitochondrial manganese superoxide dismutase (MnSOD), which convert superoxide to O_2 and H_2O_2 . The latter, also the normal by-product of oxygen reduction by oxidases such as monoamine oxidase, is removed by catalase and peroxidases, which have ubiquitous tissue distribution, but can result in greater oxidative stress susceptibility in tissues lacking sufficient activity of these enzymes.

In recent years, the upsurge in research on nitric oxide (NO) as a common second messenger in neurotransmission and signaling has resulted in recognition of its enzymatic release from activated microglia (macrophages in the central nervous system), along with superoxide. Accumulating levels of diffusible NO and superoxide give rise to peroxynitrite. Peroxynitrite and related reactive nitrogen species (RNS) are capable of both oxidation chemistry and nitration of the aromatic side-chains of tyrosine and tryptophan [4], resulting in a condition known as "nitrosative stress". There seems to be growing acceptance that ROS- and RNS-mediated damage seen in the central nervous system may reflect underlying neuroinflammatory processes [5].

Metal ions and oxidative stress

Copper, manganese, iron, and other trace redox-active transition metals are essential in most biological reactions, e.g., in the synthesis of DNA, RNA, and proteins, and as cofactors of numerous enzymes, particularly those involved in respiration. Thus their deficiency can lead to disturbances in central nervous system and other organ function. However, accumulation of redox-active transition metals in tissues in excess of the capacity of the cellular complement of metalloproteins (catalytic, transport,

storage) can be cytotoxic as a consequence of their participation in an array of cellular disturbances characterized by oxidative stress and increased free radical production [6, 7]. Only trace levels of such circulating excess of redox-active transition metal ions are required for ROS generation, generally reflecting reaction of the reduced metal oxidation states with dioxygen or hydrogen peroxide (Fenton reaction). Although the metal ions are thereby converted to their oxidized forms, these can be re-reduced by superoxide ion or cellular reducing agents such as ascorbate.

Common to both AD and PD is a dyshomeostasis of both redox-active and redox-inactive metal ions. In general, the loss of homeostasis of iron and copper in the brain is accompanied by severe neurological consequences. A role of oxidative stress in AD and PD is consistent with the finding that the areas of the brain affected by these diseases contain abnormally high levels of redox-active metals, particularly iron. However, it is not known whether the metal excesses are a cause of oxidative stress and neurodegeneration or a by-product of the neuronal cell loss. Alterations in the levels of "anti-oxidant" metalloenzymes likely also contribute to altered redox homeostasis in neurodegenerative diseases [8], though it is unclear whether this reflects altered enzyme activities or, indirectly, a disturbance in transition metal homeostasis.

Metal ions that become separated from specific storage and transport proteins still readily coordinate adventitiously to proteins, nucleic acids, and circulating amino acids, so that the concentrations of "free" aqueous metal ions are very low. This does not mean, however, that only the "free" forms are active in ROS generation. As long as the coordination sphere of the metal ion is not saturated (allowing for interaction with O_2 or H_2O_2) and supports the cycling between the oxidized and reduced states of the metal at biologically accessible potentials, the coordinated metal may serve as a catalytic center for generating ROS. For example, it is well known that the iron-EDTA complex is more active than free iron in mediating H_2O_2 -dependent damage to DNA *in vitro* [9]. Because the capacity for ROS generation depends heavily on metal coordination environments, redox *inactive* metal ions such as zinc may be pathogenic by virtue of their ability to displace redox-active metal ions from sites where redox activity of the latter is held in check. Transition metals, along with redox-inactive metal ions, may additionally contribute to neurodegeneration through their deleterious effects on protein and peptide structure, such as a pathological aggregation phenomenon. In these cases, transition metals can sometimes exert dual neurotoxic properties.

More than any other transition metal, free iron has been implicated in ROS generation *in vivo* in tissues

suffering oxidative stress. Abnormally high levels of iron have been demonstrated in a number of neurodegenerative disorders (*vide infra*). Importantly, experimental findings of increased *total* iron do not necessarily implicate increased oxidative stress if there are concomitant increases in proteins that store iron in redox inert forms. For example, ferritin (which may be upregulated in PD and AD) contains a core of insoluble, unreactive ferrihydrate. However, the entry and release of iron from ferritin occurs via its more coordinatively labile ferrous state, active in Fenton generation of hydroxyl radical. Microglia are the major sites of ferritin bound iron and are thought to be partly responsible for oxidative damage in PD and other neurodegenerative disorders. Microglia stimulated *in vivo* with phorbol ester show increased lipid peroxidation resulting from a superoxide-dependent release of iron from ferritin [10]. Besides superoxide, ferritin iron can be released by 6-hydroxydopamine, a neurotoxin implicated in PD, and by other easily oxidized catechols [11]. These studies suggest that ferritin iron release contributes to free-radical-induced cell damage *in vivo*.

In recent years, it has become evident that the regulation/management of iron at the cellular level, although primarily by the transferrin receptor and ferritin, is also under the control of the lactotransferrin receptor, melanotransferrin, ceruloplasmin, and divalent cation transporter. Thus, disruption in the expression of these latter proteins in the brain can contribute to altered brain iron metabolism in neurodegenerative disorders [12]. Control of the aforementioned proteins and therefore overall regulation of cellular iron metabolism involves the action of two iron regulatory proteins, IRP-1 and IRP-2 [13]. Under conditions of iron starvation, IRPs stabilize the transferrin receptor and inhibit the translation of ferritin mRNAs by binding to "iron responsive elements" (IREs) within their untranslated regions. In iron-replete cells, IRP1 assembles a cubane iron-sulfur cluster, which prevents IRE binding, while IRP-2 undergoes proteasomal degradation. IRP-1, but not IRP-2, is rapidly activated by extracellular H_2O_2 , establishing a regulatory connection between the control of iron metabolism and response to oxidative stress.

In addition to oxidative damage to proteins (see next section), oxidative stress conditions and the occurrence of iron- or copper-mediated Fenton chemistry results also in oxidative damage to nucleic acids, in particular RNA. 8-Hydroxyguanosine (8-OHG), a marker of nucleic acid oxidation, is commonly observed in the cytoplasm of the neurons that are particularly vulnerable to degeneration in AD [14, 15]. 8-OHG is likely to form at the site of

hydroxyl radical production, most likely by the reaction of H_2O_2 with reduced copper or iron bound to nucleic acid bases (the oxidized metal ion thereby generated is re-reduced by cellular reductants such as ascorbate or superoxide). RNA oxidation is also seen in vulnerable neurons in Parkinson disease as well as in dementia with Lewy bodies [16], suggesting that it might represent one of the fundamental abnormalities in age-associated neurodegenerative diseases.

Chemistry of oxidative protein modification occurring in oxidative stress

Protein damage that occurs under conditions of oxidative stress may represent (i) direct oxidation of protein side-chains by ROS [17] and/or RNS [18] or (ii) adduction of secondary products of oxidation of sugars, termed glycooxidation, or of polyunsaturated lipids, termed lipoxidation [2]. In addition to traditional ROS and RNS species such as peroxynitrite, oxidative damage to proteins can occur due to alternate oxidants (e.g., HOCl) and circulating oxidized amino acids such as tyrosine radical generated by metalloenzymes such as myeloperoxidase [19]. The accumulation of oxidized protein is a complex function of the rates of ROS formation, antioxidant levels, and the ability to proteolytically eliminate oxidized forms of proteins. At least in the case of AD, proteomics approaches to identify proteins that suffer oxidative modification in the disease are now yielding information on details of the oxidative stress cascade [20].

Direct protein oxidation

ROS-mediated oxidation of protein side-chains has been reviewed [21], and usually results either in introduction of hydroxyl groups or in generation of protein-based carbonyls detectable by 2,4-dinitrophenylhydrazine (DNPH), usually from oxidation of Ser and Thr side-chains or from oxidative deamination of Lys and Arg side-chains. At least some of the carbonyls seen may represent deterioration of side-chain hydroperoxides, since reductive workup reveals hydroxylated Val and Leu [22]. In addition to protein side-chain oxidation by ROS, carbonyl groups can also result from the hydroxyl radical-like reactivity of peroxynitrite [23]. Use of radical initiators accomplishes oxidation of Met and aromatic amino acid side-chains (His, Tyr, Trp), with little carbonyl generation [24]. Met sulfoxide, which represents the most common type of protein oxidation observed in proteomics studies [18], reflects oxidation of Met by H_2O_2 , peroxides, or peroxynitrite. In contrast,

oxidation of Met by hydroxyl radical and transition metals generates a cation radical which usually decomposes to give alternate products [25]. Since Met sulfoxide can be reduced to Met by methionine sulfoxide reductases [26, 27], it has become clear in recent years that cyclic oxidation/reduction of Met residues might serve as an antioxidant mechanism to scavenge ROS [28]. Although there is substantial discussion of oxidative crosslinking of proteins that accompanies oxidative stress, the only crosslink structure arising from *direct protein oxidation* identified so far is the oxidative coupling of tyrosine (giving, e.g., dityrosine) [29].

The short diffusion distance of the main damaging ROS species, hydroxyl radical, suggests that most metal-catalyzed oxidative damage to proteins occurs via reaction of H_2O_2 (or perhaps O_2) with the sites of metals coordinated adventitiously to the proteins. Examples of such "site-specific" oxidations, which have been modeled using an exogenous reducing agent and O_2 or H_2O_2 as terminal oxidant include (i) conversion of His to 2-oxohistidine [30, 31], (ii) oxidative coupling of Tyr [32], and (iii) oxidation of aromatic amino-acid side chains [33]. Adventitious binding of transition metals to proteins can in many respects mimic redox metalloenzymes, though the rates are expected to be significantly slower. In this regard, although most scientists would consider adventitious protein-bound metals to exert pro-oxidant activity, anti-oxidant effects are thus also possible if these protein-metal systems mimic anti-oxidant metalloenzymes such as SOD or catalase.

Proteins that serve to scavenge adventitious metal ions have been genetically engineered to *abrogate* the redox properties of the metal, such as is the case for the copper-binding site at the amino terminus of serum albumin. Proteins not normally functioning as metal-binding proteins may sometimes act as neuro-protectants by sequestering the metal ions in redox inactive forms. However, adventitiously-bound metals tend to have at least some redox catalytic activity, and it should be possible in some cases for adventitiously-bound metals to possess *greater* redox catalytic activity than the free metal ion. Furthermore, modification of proteins by carbonyl products of glycooxidation and lipoxidation can increase the capacity of the protein to bind copper and iron in a redox-active manner. One readily understood example is the conversion of the lysine ϵ -amino group to ϵ -(carboxymethyl)lysine (CML), which creates the well-known bidentate ligand glycinate [34, 35]. Modification of proteins by 4-hydroxy-2-nonenal (HNE) also increases binding of redox-active copper and iron [36], though this is not yet understood on a structural level. Nonetheless,

since pro-oxidant effects of such bound metals can result in further oxidation of lipids and sugars, these events provide for an autocatalytic mechanism of exacerbated oxidative damage to the proteins and the cell that contain them.

Glycooxidation

The term advanced glycation end-product (AGE) describes either protein damage that results from adduction of reducing sugars and subsequent oxidative evolution, or adduction of more reactive sugar oxidation products, termed glycooxidation. On the basis mostly of the importance of elucidating the structure and presence of AGEs that are implicated in the late-stage pathology of diabetes and diabetic complications, immunochemical tools to detect AGEs became available many years ago. AGEs represent a heterogeneous array of structures that arise mainly from the "Maillard reaction" of reducing sugars with principally the ϵ -amino groups of lysine residues, though the guanidino group of arginine also becomes heavily modified. This chemistry is related to the "browning" of foods, and complex chemical changes can occur over a long period of time. A significant research effort has been directed at elucidating particularly stable AGE structures, and obtaining antibodies specific for such structures [37].

In recent years, it has become evident that the main epitope recognized by anti-AGE antibodies is CML [38]. How CML arises during the Maillard reaction is not yet entirely clear, though it is presumed that the main pathway involves oxidative cleavage of the so-called Amadori tautomers (Lys ϵ -NHCH₂C(=O)R) of initially formed Schiff bases (Lys ϵ -N=CHCH(OH)R). Model studies demonstrate that CML can arise from essentially a Cannizzaro reaction of glyoxal with the ϵ -amino group of lysine, and glyoxal is a known product of glycooxidation as well as lipid peroxidation [39]. Although anti-CML antibody recognition can thus not distinguish between sugar- and lipid-derived modifications of proteins, CML has been popularized as an important marker of oxidative stress.

In addition to CML, condensation of glyoxal with two amine groups leads to a bis-Schiff base that has been implicated as an intermediate to the AGE termed "GOLD" (glyoxal-lysine-dimer), requiring an additional carbon that must arise from cleavage following condensation with a second molecule of glyoxal [40]. The glyoxal bis-Schiff base can also undergo an intramolecular Cannizzaro reaction, leading to a stable glycinamide lysine-lysine crosslink [41, 42]. Our model studies (unpublished) have indicated both GOLD and glycinamide crosslinks appear to form in parallel.

The glycooxidation product methylglyoxal can, in parallel to glyoxal, form N ϵ -(1-carboxyethyl)lysine (CEL) [43], and the imidazolium compound "MOLD" [44]. Although methylglyoxal is a normal intermediary metabolite in man, arising either enzymatically (e.g., from dihydroxyacetone phosphate (DHAP) by methylglyoxal synthase) or non-enzymatically by elimination of phosphate from glyceraldehyde phosphate or DHAP, its levels in blood are increased in diabetes.

Lipoxidation

Polyunsaturated lipids in lipoproteins and membranes are highly susceptible to oxidative stress damage, and suffer an ensuing radical chain autoxidation process known as lipid peroxidation. Some of the chemically and metabolically stable lipid oxidation products have been used as *in vivo* biomarkers. For example, the levels of isoprostanes, derived from arachidonic acid, and neuroprostanes, derived from docosahexaenoic acid, have been found to be increased in CSF and in diseased regions of autopsy brain tissue in neurodegenerative disease [45-48]. The unsaturated hydroperoxides generated from peroxidation of polyunsaturated lipids can break down, usually in the presence of reduced metals or ascorbate [49], to a host of mono- and bi-functional reactive aldehydes. During the past few decades, there has been an intensive effort directed at ascertaining the nature of protein and DNA modification by these aldehydes, and evidence is accumulating for their being causally involved in many pathophysiological effects associated with oxidative stress in cells and tissues *in vivo* [50].

Initially, most attention focused on malondialdehyde (MDA, 1,3-propanedial), a hydrophilic mediator that serves as the principal aldehyde detected in the TBARS (thiobarbituric acid reactive substances) assay for lipid peroxidation [51]. Although other lipid-derived aldehydes give a positive TBARS signal, MDA partitions so efficiently into the water phase relative to more lipophilic lipoxidation products, that the TBARS measurement constitutes a reasonable definition of released MDA. Although the ability of MDA to modify proteins should be limited by its existence at physiological pH mainly in the form of its "inert" resonance-stabilized enolate anion conjugate base, MDA nonetheless appears capable of forming an array of protein adducts, associated with crosslinking and a characteristic fluorescence (ex/em 390-400/460-470 nm). The most obvious crosslink would be the Lys-Lys bis-Schiff base, which would exist as its resonance-stabilized 1-amino-3-iminopropene tautomer. Although the presence of this crosslink *reversibly formed in solution* is supported by the isolation of the

corresponding propano di-lysine derivative following borohydride reduction [52], the extent to which it contributes to *long-lived* crosslinking of protein by MDA has been questioned [53], and its independent synthesis confirmed its inability to rationalize MDA-derived fluorescence [54]. The 1-amino-3-iminopropene crosslink would have to arise from initial *mono*-Schiff base adducts. Considerable amounts of the latter do form on protein Lys residues, but exist in their resonance-stabilized enamino forms (N^{ϵ} -(2-propenal)lysine adducts) [55], which would not be expected to react readily with a second amine molecule.

Work by others to determine the nature of the MDA-derived fluorophore in model studies under physiologic conditions had led to the isolation of strongly fluorescent 4-methyl-1,4-dihydropyridine-3,5-dicarboxaldehydes [56, 57]. A retro-synthetic analysis of this structure indicates the need for two MDA molecules along either side of the ring, tied together by a C_2 fragment that must result from breakdown of a third MDA molecule [53]. Consistent with this proposal, the same structures are generated in higher yield from a mixture of MDA and acetaldehyde [58]. The mentioned fluorophore would not explain the protein crosslinking potential of MDA, and further studies using a model lysine-containing peptide led to the characterization of another fluorescent MDA adduct, in this case a dihydropyridinyl pyridinium lysine-lysine crosslink derived from four molecules of MDA [59]. The relevance of this type of adduct has been questioned by workers who identified a non-fluorescent imidazole-based Lys-Arg crosslink derived from a single MDA molecule [60].

Recent studies have suggested that acrolein, another reactive three-carbon aldehyde, is also produced from peroxidation of lipids during oxidative stress. Protein Lys residues appear to constitute the main target of modification by acrolein, and a 3-formyldihydropyridine, constituting condensation of two acrolein molecules with a single lysine, was initially thought to represent the structure of the major adduct based on model studies [61]. Although this adduct was additionally reported to be detected immunochemically in oxidative stress, including AD [62], recent studies determined that the mature adduct present and detected is actually a 3-methylpyridinium structure [63]. In addition to acrolein, lipoxidation-derived α -hydroxyaldehydes have been found to modify protein Lys residues to generate fluorescent 3-hydroxypyridinium adducts [64, 65].

Our lab has focused mainly on the highly reactive bifunctional aldehyde, 4-hydroxy-2-nonenal (HNE), a major product of oxidation of ω -6 polyunsaturated (linoleoyl and arachidonyl) chains that has become the

most thoroughly studied biologically-active lipoxidation-derived aldehyde [66]. The propensity of HNE to form

Michael adducts with nucleophilic amino acids represents the dominant *initial* reaction pathway of HNE with proteins [51, 67-69]. Unlike simple α,β -unsaturated aldehydes, the Michael adducts in this case are stabilized in the form of cyclic hemiacetals. However, although the Michael adducts formed with Cys and His are stable to isolation, Michael adducts to Lys ϵ -amino groups are formed reversibly [70], and can be isolated only following reductive trapping with $NaBH_4$. Our efforts to characterize an *irreversibly-formed* HNE-lysine adduct led to the discovery of HNE-derived 2-pentylpyrroles [71, 72], that we termed the first example of an "advanced lipoxidation end-product" (ALE).

The bifunctional aspect of HNE allows it in theory to crosslink proteins by Michael addition of Cys, His, or Lys at C3 and Schiff base condensation with Lys at the C1 carbonyl [73, 74]. However, Schiff bases are hydrolytically labile, and our efforts to characterize *stable* adducts that would rationalize the known protein cross-linking activity of HNE [75-77], led to the discovery of a fluorescent four-electron oxidation product, 2-hydroxy-2-pentyl-1,2-dihydropyrrol-3-one iminium, that represents a Lys-Lys crosslink [78-80]. Recent studies in our lab indicate that the fluorescent

Lys-Lys crosslink may well constitute the major entity underlying crosslinking of proteins by HNE [81, 82]. Antibodies to the HNE-derived 2-pentylpyrrole or fluorescent crosslink have implicated HNE modification in Alzheimer's disease [77, 83-87], Parkinson's disease [88, 89], ALS [90], systemic amyloidoses [91], and Alexander's disease [92].

In the effort to confirm the structure of the HNE-derived fluorophore by independent synthesis, we found that the 4-keto cousin of HNE, 4-oxo-2-nonenal (ONE), gives rise to the fluorophore in much higher yield [78]. Soon thereafter, ONE was demonstrated to be a direct product of lipid oxidation in its own right [93, 94], suggesting that the fluorescent crosslink may arise more from ONE than from HNE. In addition to the fluorescent Lys-Lys crosslink, ONE forms simple Michael adducts of Cys, His, or Lys as well as pyrrole crosslinks that arise from Paal-Knorr condensation of these initial Michael adducts with protein Lys residues [95]. Recent mass spectroscopic studies have confirmed the formation on proteins of HNE and ONE adducts characterized in model studies [96].

When one examines the bulk of data on the role of AGEs and ALEs in oxidative stress, including the finding that at least some AGE modifications can also result from lipid peroxidation, it seems reasonable to conclude that lipoxidative damage to proteins may be

much more prominent than glycoxidative damage in the complement of neurodegenerative disease conditions. In addition, although MCO events [97] and radical-initiated events [24] can lead to 2,4-dinitrophenylhydrazine (DNPH)-detectable protein-based carbonyls, as mentioned above, the main fraction of DNPH reactivity seen in aged and diseased tissues appears to represent modification of protein side-chains by bifunctional carbonyl-containing products of lipoxidation [98] and/or glycoxidation.

Neuropathology of Alzheimer's disease

AD is characterized pathologically by the presence of two hallmark lesions in the brain, extracellular amyloid plaques, known as senile plaques (SP) and intraneuronal neurofibrillary tangles (NFT), as well as neuropil threads and a selective loss of neurons. SP contain mainly the A β peptide, whereas NFT are composed mainly of the microtubule-associated protein (MAP) tau present as paired helical filaments (PHF). Much excitement in recent years has come from the discovery of a multiplicity of mutations in the genes encoding the amyloid- β precursor protein (A β PP) and/or the presenilins, that account for the bulk of the familial cases of AD on the basis of overproduction of A β PP and/or altered A β PP proteolytic processing, both leading to increased Ab. Transgenic mice overexpressing A β PP or other human mutant AD-related proteins, as well as animals expressing more than one of these mutations, exhibit many of the neuropathologic and behavioral features of the human disease, including the development of senile plaques and some neurotoxicity. In addition, NFT can be produced in mice expressing mutant tau protein, and tangle formation is further enhanced in animals that also express mutant A β PP [99]. The animal models of AD have been used to develop and test treatments that reduce brain levels of the A β , neuritic plaque load and glial activation, and some have been found to restore learning and memory function [100]. Although the relationship between these genetic models and neuronal loss in AD are tentative, the mechanisms involved may be partially shared with sporadic AD.

There is considerable debate as to whether neuronal loss in AD reflects appearance of either SP or NFT and if so, which one plays a more important role. A majority of workers in the field still favor the amyloid cascade hypothesis, specifying that the onset and progression of AD is initiated by aggregation of A β into toxic fibrillar deposits within the extracellular space of the brain, thereby disrupting neuronal and synaptic function and eventually leading to neuronal degeneration and

dementia [101]. However, A β , formed upon proteolytic processing of A β PP by β - and γ -secretases, has a widespread distribution through the brain and body in healthy individuals throughout life. Moreover, *soluble* A β appears to serve a variety of physiological functions, including modulation of synaptic function, facilitation of neuronal growth and survival, protection against oxidative stress, and surveillance against neuroactive compounds, toxins and pathogens [102]. At the other extreme, the mature SP are apparently also non-toxic because neuronal loss in AD does not occur in the vicinity of SP deposition. Also, A β deposition is inversely correlated with the levels of 8OHG since this oxidative marker is found physically distant from the A β deposits [14]. Thus, recent focus on A β toxicity has been on fibrillar polymers that are toxic to cultured neurons, and especially on smaller oligomers [103], particularly those containing more A β (1-42) than A β (1-40) [104]. The mechanism of neurotoxicity of A β (1-42), wherein Met-35 appears to play a critical role, is usually considered to involve induction of oxidative stress and lipid peroxidation [105].

Despite the genetic evidence implicating a pathologic role of A β , only tau pathology and NFT have been found to correlate with symptom presentation in patients [106]. In normal adult brain, the microtubule-associated protein tau binds microtubules through its tubulin-binding domains, and this assembly depends partially upon the degree of phosphorylation. By regulating microtubule assembly, τ has a role in modulating the functional organization of the neuron, particularly in axonal morphology, growth, and polarity [107]. Besides the role in microtubule stabilization, tau has other functions such as membrane interactions or anchoring of enzymes [108]. The microtubule-binding domains in tau have been identified, and it is found that phosphorylation of Ser/Thr-Pro moieties within these domains abrogates the association of tau with microtubules [109, 110].

In AD, tau is "hyperphosphorylated", at up to 22 different sites, at the stage where it loses its microtubule-binding and stabilizing function and aggregates into PHF [111]. Tau hyperphosphorylation reflects both an abnormal action of kinases, as well as decreased phosphatase activity [112]. The pool of soluble tau that is incapable of interacting with microtubules becomes prone toward self-aggregation at high concentration. There is evidence also that tau undergoes a hierarchical series of truncations at both N- and C-termini, that affect its conformationally-dependent folding, as it transforms from an unfolded monomer to the structured polymer characteristic of NFT [113].

It seems clear that tau hyperphosphorylation is a critical factor underlying neurofibrillary pathology. Supporting evidence is the appearance of epitopes

recognized by anti-PHF antibodies, as a result of kinase stimulation in neuronal cells, that precedes death of these cells [114]. Not only does hyperphosphorylated tau fail in its normal function in stabilizing microtubules, but it reflects a “gain of toxic function” due to its sequestering normal tau and other MAPs, resulting in the disruption of microtubules [115, 116]. Phosphorylation of tau induces its binding by chaperones such as hsp 27, possibly a cellular survival response aimed at preventing more toxic conformations of the protein [117]. In addition, packaging of dysfunctional tau into PHF of NFT may reflect the futile effort of the cell to reduce the effect of soluble “toxic” tau [116]. A pseudohyperphosphorylated form of tau (substituting Glu for Ser residues in the sequence known to be phosphorylated), expressed in neural differentiated PC12 cells, was shown to exhibit reduced microtubule interaction, to fail to stabilize the microtubule network, to sensitize the cells to other apoptotic stimuli, and, after longer culture, to be cytotoxic [118]. Recent research has focused on evidence that dysfunctional proteasome activity, arising from inhibitory binding of PHF-tau, contributes to the intraneuronal accumulation of oxidatively damaged proteins in the brains of patients with AD, which may be sufficient to induce neuronal degeneration and death [119].

NFT display resistance to proteolysis, and this may reflect in part the presence of transaminase-mediated γ -glutamyl- ϵ -lysine crosslinks [120] among proteins identified in NFT (hsp27, SN, ubiquitin, parkin) [121]. It has been our contention that the *persistent insolubilization* and *permanency* of NFT aggregates probably represents, at least in part, cementing of the aggregates by processes associated with oxidative stress. In particular, the finding that most of the covalent, including crosslinking, modifications induced by products of oxidative stress (see above) are seen in apparently normal neurons in AD and at pre-NFT PHF-tau stages [86, 122, 123] suggests that these modifications play at least partially a causative rather than by-stander role in the neurofibrillary pathology in AD. We recently showed that seven distinct antibodies raised against NFT that recognize unique epitopes of tau in AD, recognize phosphorylated tau more strongly after treatment with HNE [124]. These findings support the idea that HNE modifications of tau promote and contribute to the generation of the major conformational properties defining NFT.

Metals in Alzheimer’s disease

The likely role of an imbalance of redox-active metals in rationalizing the pervasive oxidative stress damage seen in AD brain has been mentioned above.

Several studies have indicated imbalances of many trace elements in AD, including Al, Si, Pb, Hg, Zn, Cu, and Fe, with only the latter two being redox active. Micro particle-induced X-ray emission data show that Zn(II), Fe(III), and Cu(II) are significantly elevated in AD neuropil and that these metals are significantly further concentrated within the core and periphery of senile plaques [125]. Over-accumulation of iron in AD has been found in the hippocampus, cerebral cortex and basal nucleus of Meynert, and colocalizes with SP and NFT [126]. These results extend earlier studies reporting increased levels of iron, transferrin, and ferritin in AD. The association of iron with NFT may be, in part, related to iron binding to their primary protein constituent, tau [127]. Dysregulation of iron homeostasis in AD, possibly involving ceruloplasmin [128], is also indicated by the finding that IRP-2 is specifically co-localized in AD with redox-active iron in NFT, SP neurites, and neuropil threads [129]. These results suggest that alterations in IRP-2 may be directly linked to impaired iron homeostasis in AD.

Increased iron in AD may also be explained by data from our laboratory and others indicating that heme oxygenase-1 (HO-1) is induced in AD brains [130, 131]. HO-1 catalyzes the conversion of heme to iron and biliverdin, which, in turn, is reduced to bilirubin, an antioxidant. Since HO-1 is induced in proportion to the level of heme, the induction of HO-1 is suggestive of abnormal turnover of heme in AD. This may be connected to the findings in AD of abnormalities in mitochondria, which contain numerous heme proteins. In turn, the increase in heme induces synthesis of more HO-1 suggesting that mitochondrial turnover promotes oxidative stress via increase of redox-active iron [132].

An assay for *redox-active* iron shows it to be increased in NFT as well as in A β deposits [133], and aluminum, which also accumulates in NFT-containing neurons [133], can stimulate iron-induced lipid peroxidation [134] by displacing it from inert redox sites. Also, using an *in situ* iron detection method, we found a marked association of redox-active iron with both NFT and SP in AD [135]. Further studies revealed the presence of non-enzymatic redox activity in both SP and NFT using a more general detection method that responds to redox-active copper as well as iron [36]. These results suggest that metal accumulations are major producers of the ROS responsible not only for the numerous oxidative stress markers that appear on NFT and SP, but also for the more global oxidative stress parameters observed in AD.

Evidence for imbalances of trace metal homeostasis in AD have led over the years to efforts to identify possible interactions of metal ions with both A β PP and A β . It has been found that Al, Fe, Zn, and Cu accelerate aggregation of A β , with the

relative effectiveness being pH-dependent [136, 137]. The finding that chelators can at least partially solubilize β -amyloid deposits from AD [138] suggests a possible physiological role of Cu(II), Zn(II), and/or Fe(II) in A β deposition, thereby explaining the enrichment of these metals with SP in AD [139, 140]. Although metal-induced aggregation of A β may not reflect redox chemistry, copper and iron bound to soluble or aggregated forms of the peptide do seem to be redox active and thus capable of mediating ROS production [36, 141-143]. At the same time, whereas A β complexes of iron and zinc are toxic to cultured neurons, A β -copper complexes are not neurotoxic and the neurotoxicity of both iron and copper was reduced by binding to A β [140]. A recent study examining the effects of Cu(II), Zn(II), Fe(III), and Al(III) on A β (1-42) aggregation that took into account contaminating metals in "A β (1-42) only" preparations, revealed a complex interplay between these metals, finding that only the trivalent metal ions induced β -pleated sheet formation [144]. It was concluded that chelation therapy directed at Cu and Zn that does not also chelate Al and Fe, might exacerbate amyloid fibril formation.

There is always the question of whether increased tissue burden of metals reflects at least in part dietary factors. There is mixed evidence for a role of dietary copper in experimental models of AD, one study in transgenic mice suggesting a stabilization of SOD and reduced A β production [145], and another study in rabbits showing induction of A β plaques and learning deficits [146].

Current research has led to the characterization of A β PP as a copper-binding protein involved in regulating copper uptake and efflux, and copper regulates A β PP expression [147]. Previous work found that binding of A β PP can reduce Cu(II) to Cu(I) concomitant with production of a disulfide linkage [148]. Subsequent exposure to H₂O₂ results in reoxidation of Cu(I) and concomitant site-specific cleavage of A β PP [148, 149]. Redox chemistry associated with A β - or A β PP-bound metals could contribute to a perturbation of free radical homeostasis and resulting ROS-mediated neuronal toxicity in AD. An iron responsive element on A β PP points to a role for iron in the metabolism of A β PP and offers another mechanism whereby iron chelators can reduce Abeta peptide burden during Alzheimer's disease [150].

Taken together, the studies by us and others indicating the presence of redox-active iron and copper in AD pathology suggest that these metal accumulations could be major contributors to not only local ROS-mediated damage but also more global oxidative stress

parameters tied to neuronal toxicity in AD.

Role of oxidative stress in Alzheimer's disease

Oxidative damage of biomacromolecules [151] has been described in association with neuronal degeneration in AD brains: 1) DNA and RNA oxidation is marked by increased levels of 8-hydroxy-2-deoxyguanosine (8OHdG) and 8-hydroxyguanosine (8OHG) [14, 15] and increased DNA oxidation and decreased repair in CSF [152, 153]; 2) protein oxidation is marked by elevated levels of protein carbonyls and nitrotyrosine [154-156]; 3) lipid peroxidation is marked by higher levels of TBARS and isoprostanes, as well as protein modification by HNE [48, 86, 157]; 4) sugar oxidation is marked by increased protein glycation and glycooxidation [158-164]. Moreover, crosslinking of proteins, such as by bifunctional lipoxidation-derived aldehydes, may lead to the resistance of the lesions to intracellular and extracellular removal even though they are extensively ubiquitinated [165], and this resistance of NFT to proteolysis might play an important role in the progression of AD [166].

Investigations into possible sources of oxidative stress and metal ion excesses that underlie increased ROS generation in AD have focused recently on mitochondria, not only because of their propensity for ROS generation [167], but also because mitochondrial turnover and associated degradation of heme proteins can release iron [168]. Heme oxygenase-1, which is increased in AD [130, 131, 169], catalyzes the conversion of heme to the antioxidant biliverdin, but the other product is iron. There is substantial evidence for a down-regulation of the mitochondrial respiratory chain in AD [170], and evidence for increased turnover of mitochondria is the finding of mtDNA and cytochrome oxidase in the neuronal cytoplasm in AD. Mitochondrial structural abnormalities and mtDNA deletions have also been observed, in each case occurring before neuronal degeneration and amyloid deposition [171]. These findings together suggest that mitochondrial dysfunction, acting in concert with cytoskeletal pathology, serves to increase redox-active metals and initiates a cascade of abnormal events in the early stages of AD pathogenesis.

A state of pervasive oxidative stress underlying the sites of damage in AD is further provided by immunocytochemical evidence for the upregulation of the "antioxidant enzyme" superoxide dismutase [172, 173], glucose-6-phosphate dehydrogenase [174, 175], and increased levels of reduced sulfhydryls [175, 176] in vulnerable neurons. Increased oxidative damage in AD may thus represent a failure of adequate antioxidant defenses [177]. Notwithstanding, there is growing support for the hypothesis that oxidative markers are apparent mainly in initial stages of the disease, and that compensating cellular defense mechanisms eventually

result in diminished oxidative damage [178, 179].

In fact, there is now considerable evidence that the deposition of tau in NFT and A β in SP may represent responses to the oxidative stress condition, whereby these protein aggregates serve to trap reactive products of oxidative stress. It has been shown that NFT enables neurons to survive decades [180]. Tau and the related neurofilament proteins, especially NFH, appear adapted to oxidative stress due to their high content of lysine [181], suggesting that they can serve as a "buffer" for reactive carbonyl products of lipid and sugar oxidation. Tau also binds iron, and can contribute to iron homeostasis [36]. Also, an antioxidant role for A β *in vivo* is supported by the finding that an increase in A β deposition in AD cortex is associated with a decrease in neuronal levels of 8OHG, i.e., with decreased oxidative damage [14, 182]. Similar negative correlation between A β deposition and oxidative damage is found in patients with Down syndrome [182, 183]. A β deposits observed in both studies mainly consist of early diffuse plaques, meaning that these diffuse amyloid plaques may be considered as a compensatory response that reduces oxidative stress [184].

The strong chelating properties of A β for zinc, iron, and copper explain the reported enrichment of these metals in amyloid plaques in AD discussed above, but also suggest that one function of A β is to sequester these metal ions (though the chelation would have to lower redox activity for this to be protective). Also, Met-35 in A β can scavenge peroxides, with the resulting methionine sulfoxide being re-reduced by methionine sulfoxide reductase. Thus, although there is substantial evidence of a pro-oxidant role of especially A β (1-42), the *overall* net effect of A β on oxidative stress is a complex question. While more studies are required to understand the role of NFT and SP deposition in neuronal and extra-neuronal homeostasis, the results obtained to date clearly implicate a regulated process that, at least through mid-stages of the disease, might have a neuroprotective role.

Role of oxidative stress in the pathogenesis of Parkinson's disease

Parkinson's disease involves the selective destruction of the dopamine-producing cells of the substantia nigra in the midbrain. The cause and pathogenesis of PD remain unknown: mitochondrial dysfunction, oxidative damage, environmental factors, and genetic predisposition might all be involved. Because oxidative stress is intimately linked to other components of the degenerative process, it is difficult

to determine whether oxidative stress leads to, or is a consequence of, these events [185]. The principal histopathologic hallmark of PD are Lewy bodies, eosinophilic intraneuronal filamentous inclusions found in magnocellular neurons of brain stem nuclei, predominantly substantia nigra and locus coeruleus. Like PHF, Lewy bodies are insoluble in detergent and are resistant to removal from the extracellular space following neuronal death [186]. Structurally similar Lewy bodies are also found in cortical neurons in PD and diffuse Lewy body disease. The principal protein constituent of Lewy bodies is α -synuclein, originally identified as the precursor of a peptide known as non-A β component (NAC) that is tightly associated with amyloid deposits in AD.

There is substantial evidence that mitochondrial complex I inhibition may be the central cause of sporadic PD [187]. Evidence centers on the observed 30-40% decrease in complex I activity in the substantia nigra. The decrease in activity seems to be matched by a decrease in protein content that appears to reflect a mtDNA defect and underproduction of certain complex I subunits [188, 189]. Support for a causative role of the complex I deficiency in PD is that chronic infusion of the complex I inhibitor rotenone in rat brain results in a selective loss of nigral DA neurons and the appearance of cytoplasmic α -synuclein inclusions resembling Lewy bodies, the underlying mechanisms for which appear to involve oxidative stress. Evidence for oxidative stress in PD is the finding of oxidative damage to DNA [190, 191] and protein [192, 193] observed in the nigro-striatal region of PD brain, as well as immunocytochemical evidence for protein nitration [194], glycation [195], and HNE modification [88, 89].

Notwithstanding, with the discovery of missense mutations in two genes, α -synuclein and parkin, which are responsible for rare cases of heritable early-onset PD, there has recently been a major shift in emphasis in PD research [196]. Overexpression of normal α -synuclein only modestly affects cell viability, but most studies show that overexpression of mutant α -synuclein proteins is neurotoxic, most commonly by induction of apoptosis [197, 198]. Although the physiological functioning of α -synuclein remains to be fully defined, recent data are suggestive of a role in regulating membrane stability and neuronal plasticity [199].

Numerous proteins have been shown to associate with soluble α -synuclein [200], and the peptide has been shown to bind to tau and stimulate its phosphorylation [201]. At the same time, when α -synuclein and tau coexist, they tend to form homopolymers rather than heteropolymers. This does not preclude α -synuclein-tau interaction exerting a physiological role,

because α -synuclein induces fibrillization of tau, and coincubation of tau and α -synuclein synergistically promotes fibrillization of both proteins [202]. NFT frequently coexist with Lewy bodies in the same neurons of patients with diffuse Lewy body disease [203]. Moreover, though there is wide recognition of a disease class known as tauopathies (e.g., NFT formation in AD) and another class known as synucleinopathies (e.g., Lewy body formation in PD), emerging evidence indicates that there is frequent overlap of the pathological and clinical features of all these patients [204].

A connection between α -synuclein aggregation and oxidative stress parameters seen in PD is suggested by the finding that amyloid-like aggregates of α -synuclein similar to those seen *in vivo* can be induced by coincubation with Cu(II) [205], Fe/H₂O₂ [206], or cytochrome *c*/H₂O₂ [207]. Also, studies using anti-nitrotyrosine antibodies have shown the presence of nitrated α -synuclein in human Lewy bodies and other α -synuclein inclusions [208]. Nitrated α -synuclein monomers and dimers but not oligomers were found to accelerate the rate of fibril formation of unmodified α -synuclein when present at low concentrations [209]. This suggests that post-translational modification of α -synuclein could promote the formation of hallmark intracytoplasmic inclusions found in PD and other synucleinopathies.

It seems clear that dysfunction of α -synuclein is a common feature of all forms of PD. However, exacerbating the fact that mutations in or oxidative modifications of α -synuclein can promote aggregation, is the increased evidence for dysfunction of the ubiquitin-proteasome system, which relates to the second set of familial PD genetic defects discovered, namely mutations in two critical enzymes: parkin, a ubiquitin ligase [210], and ubiquitin C-terminal hydrolase L1. Defects of the ubiquitin-proteasome system lead to a condition known as “*proteolytic stress*” [211], where the cell accumulates misfolded or abnormal (e.g., oxidatively modified) proteins that cannot be cleared. The inability of neurons to degrade abnormal protein is now a guiding principle affecting numerous neurodegenerative conditions, and can reflect interference with polyubiquitination [212] as well as inhibition of the proteasomal pathway. It is not surprising that mutant forms of α -synuclein found in familial PD cannot be easily cleared, but proteolytic stress appears also to underlie nigral pathology in *sporadic* forms of PD [213]. Oxidative stress can impair the ubiquitin-proteasome process directly, and products of oxidative damage, such as HNE, can damage the 26S proteasome [185]. Furthermore, proteasome binding to insoluble α -synuclein filaments and soluble α -synuclein oligomers results in marked

inhibition of its chymotrypsin-like hydrolytic activity [214]. At the same time, α -synuclein is required for the fibrillar nature of ubiquitinated inclusions induced by proteasomal inhibition in primary neurons [215]. The finding that Lewy bodies in PD are immunoreactive for ubiquitin supports a link between defects in energy metabolism and the disposal of damaged proteins in the development of PD [216].

Metals in Parkinson's disease

Abnormally high levels of iron and oxidative stress have been demonstrated in a number of neurodegenerative disorders characterized by nigral degeneration such as PD, multiple system atrophy, and progressive supranuclear palsy. Selective dopaminergic cell loss in PD is correlated with increased levels of cellular iron. Iron accumulates in astrocytes in the SN pars compacta of old rats [217, 218]. At the same time, there is an increase in the Fe(III)/Fe(II) ratio and a decrease in GSH [219]. One interpretation is that mitochondrial sequestration of redox-active iron in aging nigral astroglia may be one factor predisposing the senescent nervous system to PD. Moreover, there is circumstantial evidence that the intracellular redox imbalance results in aberrant oxidation of dopamine to 6-hydroxydopamine, which in turn can undergo autoxidation to the corresponding quinone concomitant with generation of superoxide. This reaction cascade, either by itself, or as amplified by redox cycling of this quinone leading to further generation of ROS at the expense of cellular reductants, can serve to explain the ultimate demise of these neurons. Studies to clarify the mechanism of dopamine oxidation *in vitro* have demonstrated conversion to 6-hydroxydopamine in the presence of Fe(II) and either H₂O₂ or alkyl peroxides [220]. Since the non-ferritin-bound labile iron pool is presumed responsible for dopaminergic degeneration via Fenton chemistry, efforts have focused on developing brain-permeable iron chelators [221].

An epidemiological study revealed that a dietary iron intake in the highest quartile compared with those in the lowest quartile had an increased risk of PD [222]. Also, there was an apparent joint effect of iron and manganese, with dietary intake above median levels of both together conferring a nearly doubled risk of PD. There is little evidence for a similar effect of dietary iron on AD prevalence. A series of recent studies suggest that iron regulatory proteins (IRPs) coordinate both cellular iron levels and energy metabolism, both of which are disrupted in Parkinson's disease (PD) and may in turn contribute to increased levels of oxidative stress associated with the disease.

Iron has also been recently implicated in promotion of α -synuclein aggregation either directly or via increasing levels of oxidative stress suggesting an important role for it in Lewy body formation [223]. There is substantial support for the hypothesis that oxidative stress, augmented iron deposition, and mitochondrial insufficiency constitute a single neuropathologic "lesion", where any one component in this triad obligates the other two [224].

On the basis of evidence for increased metals seen in a PD autopsy tissue, there has been a significant effort to ascertain if there are altered levels of metal ions in bodily fluids of live patients. One study looking at serum levels of 12 elements found significant decreases in Al and S in both early and severe PD compared to controls, Fe and Zn were lower in severe PD, and several other elements (K, Mg, Cu, P) were higher in early and severe PD [225]. In another study, quantification of 8 elements in whole blood, serum, urine, and cerebrospinal fluid (CSF) found a complex interplay between redox-active and redox-inactive elements [226]. Interestingly, the finding that the oxidant status was increased and the anti-oxidant capacity decreased in serum from PD patients, compared to controls, was correlated best with Fe, which along with Zn was increased in blood, whereas serum levels of Cu were lower.

Chronic exposure to Mn results in extrapyramidal syndromes resembling PD, and Mn has therefore been labeled as an environmental toxic factor that induces brain dysfunction. However, Mn(II) itself is inactive in Fenton chemistry, and there is no convincing *in vivo* evidence for a prooxidant role of Mn in the brain. Although Mn was once considered as a possible contributor to idiopathic PD, it is now clear that the PD-like syndrome induced by Mn poisoning may have little connection to nigrostriatal damage occurring in idiopathic PD. Recent data indicates that Mn-induced parkinsonism (termed manganism) can be differentiated from PD because accumulation of Mn and damage occur mainly in the basal ganglia (pallidum and striatum), rather than in the pars compacta of the SN [227, 228].

The role of oxidative stress and metals in multiple sclerosis

Multiple sclerosis (MS) is a chronic inflammatory demyelinating disease of the central nervous system that is generally believed to be of autoimmune origin [229, 230]. This disabling disorder is characterized pathologically by selective and coordinated inflammatory destruction of the myelin sheath, with ensuing damage to the underlying axon [231].

Although it is generally accepted that the immune system contributes to tissue damage, the cause of MS is still unclear. However, accumulating data indicate that oxidative stress plays a major role in the pathogenesis of MS.

It is well known that inflammation might raise reactive oxygen and nitrogen species levels leading to oxidative stress. One of the most abundant sources of reactive species, apart from the electron-transport chain of mitochondria, is the respiratory burst system of activated microglia. ROS and RNS generated by macrophages have been implicated as mediators of demyelination and axonal injury in both experimental autoimmune encephalomyelitis (EAE - the generally accepted animal model for the study of MS) and MS [232, 233]. In addition, free radicals can activate certain transcription factors, such as nuclear transcription factor-kappa B, which up-regulates the expression of many genes involved in EAE and MS, such as tumor necrosis factor- α , inducible nitric oxide synthase (iNOS), intracellular adhesion molecule 1 and vascular cell adhesion molecule 1 [234, 235].

An analysis of the blood (plasma, erythrocytes and lymphocytes) of 28 MS patients compared to 30 healthy age-matched controls revealed that MS patients displayed significantly reduced plasma levels of ubiquinone and vitamin E, and depressed erythrocyte glutathione peroxidase [236]. It was concluded that the blood of patients with multiple sclerosis shows signs of significant oxidative stress. This conclusion supports an earlier study that found a significant decrease in glutathione peroxidase activity in the erythrocytes of 24 MS patients, compared to controls [237]. Abnormal catalase activity has been reported also in the granulocytes and erythrocytes of MS patients [238] being decreased in the former and increased in the latter, compared to normal controls. In addition, a 38% increase in lipid peroxidation, significantly elevated levels of oxidized glutathione, and a reduction in the plasma vitamin E:lipid ratio has been reported during the active phase of MS [239]. Examination of CSF showed significantly higher concentrations of isoprostanes [240], an increase MDA and glutathione reductase activity, and a decrease of glutathione peroxidase activity [241]. Moreover, direct examination of MS plaques has revealed increased free radical activity, along with decreased levels of glutathione, α -tocopherol and uric acid [242]. Furthermore, it has been shown that activated mononuclear cells of MS patients produce high amounts of reactive oxygen and nitrogen species and that oxidative damage to DNA, including mitochondrial DNA [243], develops in association

with inflammation in chronic active plaques [244].

Because commonly activated immune cells release glutamate, glutamate excitotoxicity is thought to be involved in MS lesion formation and axonal damage. Demyelinating lesions caused by excitotoxins can be similar to those observed in MS, causing histologically similar damage. Excessive release of glutamate from damaged neurons can lead to the over-excitation of surrounding neurons and subsequent apoptotic cell death mediated by several types of glutamate receptors [245]. It was found that oligodendrocytes are highly susceptible to glutamate excitotoxicity mainly via AMPA/kainate receptors [246]. Recent experimental studies showed that the treatment with AMPA/kainate antagonists resulted in substantial amelioration of experimental EAE [247]. The AMPA/kainate antagonists also increased oligodendrocyte survival and reduced dephosphorylation of neurofilament H, an indicator of axonal damage [248, 249]. A recent study showed that riluzole, an antagonist of glutamate neurotransmission, dramatically reduces the clinical severity, inflammation, demyelination and axonal damage of EAE mice [250]. Consistent with this possibility, the neurological deficit resulting from EAE has generally been reduced by trial therapies to diminish the concentration of ROS [251]. There is also evidence in MS of an increase in CSF levels of glutamate in association with the severity and course of the disease [252, 253] and that glutamate production by macrophages may underlie axonal damage and oligodendrocytes death in MS lesions [254].

Several investigators have shown that inhibition of iNOS, as well as the use of anti-sense knockdown of iNOS, suppress EAE in mice and rats [255-258]. Furthermore, it has been shown that inhibition of iNOS, or scavenging nitric oxide or peroxynitrite by uric acid, inhibited neurological deficits in mice with EAE, while withdrawal of iNOS inhibitor resulted in the appearance of neurological signs within 24 hours [259, 260].

Evidence for a pathogenic role of ROS in MS pathology has led to the employment of several antioxidant strategies in an effort to ameliorate EAE. It has been shown that oral administration of the oxidant-scavenger N-acetyl-L-cysteine, which can effectively raise intracellular glutathione levels, inhibited the induction of acute EAE [261]. Additionally, the intraperitoneal administration of catalase before the onset of neurological deficit delayed the onset of EAE and reduced its severity and duration [262]. A salen-manganese complex, Euk-8, that may be regarded as a prototype molecule of a new class of synthetic catalytic scavengers with combined superoxide dismutase and catalase activity, was also shown to be effective in EAE mice [263]. It was also shown that α -

lipoic acid inhibits the symptoms in EAE mice [264]. However, other workers have suggested that α -lipoic acid can effectively interfere with the autoimmune reaction associated with EAE through non-antioxidant mechanisms [265]. Recently, bilirubin has been found to prevent both acute and chronic EAE in mice [266]. More significantly, bilirubin suppressed ongoing clinical EAE and halted EAE progression when given after disease onset.

Following the encouraging findings in the EAE models, many researchers have suggested that dietary antioxidant intake may help to stop or delay disease evolution. An older study showed that supplementation of the diet of MS patients with antioxidants (selenium, vitamin C and vitamin E) could increase and normalize the glutathione peroxidase activity and the cellular content of linoleic acid in erythrocytes and hematogenous cells within 3 weeks [267]. Also, in a large study of 144 MS patients, adherence to a low-fat diet enriched with vitamin A was shown to delay the progression of symptoms and to lower death rates [268]. Indeed, high dose antioxidant supplementation is now recommended for such patients and has been shown to help normalize the otherwise low glutathione peroxidase activity in MS [269]. It has been shown that low serum uric acid levels of MS patients can be raised and maintained by oral administration of its precursor inosine [270]. Despite the encouraging results obtained with antioxidants, more epidemiological and clinical trials should be performed to clarify the real efficacy of antioxidant strategies in the fight against MS progression.

Although there were early conflicting reports of abnormal iron deposition in the brain in MS, a study using an enhanced staining method revealed that in addition to its normal deposition in oligodendrocytes and myelin, iron was detected in reactive microglia, amoeboid microglia and macrophages in MS brains [271]. It is thought that the localization of iron deposition in MS indicates potential sites where iron could promote oxidative damage. The concentrations of ferritin, transferrin and iron were measured in the CSF of MS and control patients [272]. Ferritin levels were significantly elevated in the CSF of chronic progressive active MS patients, but not in patients with relapsing-remitting disease, and levels of transferrin and total iron were similar in all groups. However, in brain tissue from MS patients, disruptions were found in the normal pattern of binding distributions for both transferrin and ferritin [273]. In addition, compared to controls, levels of manganese are significantly decreased and levels of copper significantly elevated in the CSF of MS patients, whereas no differences were found for zinc [274]. Evidence for the role of disrupted iron metabolism and iron-mediated oxidative stress in the pathogenesis of

MS and EAE has been recently reviewed [275].

Conclusions

Although demyelination in MS is normally considered an inflammatory disease, evidence for a neurodegenerative component in MS brings this disease process into a common discussion of the role of oxidative stress in age-related neurodegenerative disorders such as AD and PD. In AD and PD, the degree of cognitive and/or clinical impairment is correlated to the degeneration and subsequent loss of specific neuronal populations, presumed to correlate in turn with the pathological lesions. The proteins depositing in these lesions are selectively vulnerable to and, at least sometimes participate in, oxidative stress, and it is tempting to speculate that free radical oxygen chemistry and a dyshomeostasis of metal ions plays a pathogenetic role in all these neurodegenerative conditions. However, it is as yet undetermined what types of oxidative changes occur early in the disease, and what types are secondary manifestations of neuronal degeneration, as well as how the effects of oxidative stress are differentially manifested in the specific neuronal populations affected by the individual disease conditions. Thus, since most antioxidants considered for therapeutic intervention also have metal-reducing capacity, devising a successful regimen of antioxidant action to retard the progression of these diseases remains a complicated goal. Nonetheless, at least in the case of AD, recent clinical data indicate that the use of a combination of various antioxidants might indeed be effective in preventing the disease [276].

Work carried out in our lab and elsewhere has focused on oxidative modifications to biomacromolecules that provide a window to view the disease process. Since these oxidative markers show up, at least in the case of AD, are early stages of development of the classical pathological lesions, it is our contention that oxidative stress-induced modifications contribute to disease pathogenesis. The role of metal ions in mediating oxidative stress reflects the complex intertwining between effects of both redox-active and redox-inactive metals in competing with each other for available metal-binding proteins and adventitious protein binding sites that may either enhance or reduce redox activity. Thus, when considering chelation therapy as an adjunct to possible antioxidant intervention in these disease states, it will be important to recognize the dualistic action of metals in catalyzing both prooxidant and antioxidant reactions. Certainly, developing a more complete understanding of the biochemical bases underlying these disease states will continue to lead to improved animal and cellular model systems that can be used to test pharmacological

agents as prospective therapies.

Acknowledgement

Work in the author's laboratories have been supported by the National Institutes of Health (AG14249, HL53315, NS38648) and by the Alzheimer's Association.

Submitted on invitation.

Accepted on 19 May 2005.

REFERENCES

1. Andersen JK. Oxidative stress in neurodegeneration: cause or consequence? *Nat Med* 2004;10 Suppl:S18-25.
2. Sayre LM, Smith MA, Perry G. Chemistry and biochemistry of oxidative stress in neurodegenerative disease. *Curr Med Chem* 2001;8(7):721-38.
3. Facheris M, Beretta S, Ferrarese C. Peripheral markers of oxidative stress and excitotoxicity in neurodegenerative disorders: tools for diagnosis and therapy? *J Alzheimers Dis* 2004;6:177-84.
4. Alvarez B, Radi R. Peroxynitrite reactivity with amino acids and proteins. *Amino Acids* 2003;25:295-311.
5. Floyd RA. Neuroinflammatory processes are important in neurodegenerative diseases: an hypothesis to explain the increased formation of reactive oxygen and nitrogen species as major factors involved in neurodegenerative disease development. *Free Radic Biol Med* 1999;26:1346-55.
6. Sayre LM, Perry G, Smith MA. Redox metals and neurodegenerative disease. *Curr Opin Chem Biol* 1999;3(2):220-5.
7. Sayre LM, Perry G, Atwood CS, Smith MA. The role of metals in neurodegenerative diseases. *Cell Mol Biol* 2000;46(4):731-41.
8. Corson LB, Folmer J, Strain JJ, Culotta VC, Cleveland DW. Oxidative stress and iron are implicated in fragmenting vacuoles of *Saccharomyces cerevisiae* lacking Cu,Zn-superoxide dismutase. *J Biol Chem* 1999;274:27590-6.
9. Papavassiliou AG. Chemical nucleases as probes for studying DNA-protein interactions. *Biochem J* 1995;305:345-57.
10. Yoshida T, Tanaka M, Sotomatsu A, Hirai S, Okamoto K. Activated microglia cause iron-dependent lipid peroxidation in the presence of ferritin. *NeuroReport* 1998;9:1929-33.
11. Double KL, Maywald M, Schmittel M, Riederer P, Gerlach M. In vitro studies of ferritin iron release and neurotoxicity. *J Neurochem* 1998;70:2492-9.
12. Qian ZM, Wang Q. Expression of iron transport proteins and excessive iron accumulation in the brain in neurodegenerative disorders. *Brain Res Rev* 1998;27:257-67.
13. Pantopoulos K. Iron metabolism and the IRE/IRP regulatory system: an update. *Ann NY Acad Sci* 2004;1012:1-13.
14. Nunomura A, Perry G, Pappolla MA, Wade R, Hirai K, Chiba S, Smith MA. RNA oxidation is a prominent feature of vulnerable neurons in Alzheimer disease. *J Neuroscience*

- 1999;19:1959-64.
15. Nunomura A, Perry G, Aliev G, Hirai K, Takeda A, Balraj EK, Jones PK, Ghanbari H, Wataya T, Shimohama S, Chiba S, Atwood CS, Petersen RB, Smith MA. Oxidative damage is the earliest event in Alzheimer disease. *J Neuropathol Exp Neurol* 2001;60:759-67.
 16. Nunomura A, Chiba S, Kosaka K, Takeda A, Castellani RJ, Smith MA, Perry G. Neuronal RNA oxidation is a prominent feature of dementia with Lewy bodies. *Neuroreport* 2002;13(16):2035-9.
 17. Stadtman ER. Protein oxidation and aging. *Science* 1992;257:1220-4.
 18. Dalle-Donne I, Scaloni A, Giustarini D, Cavarra E, Tell G, Lungarella G, Colombo R, Rossi R, Milzani A. Proteins as biomarkers of oxidative/nitrosative stress in diseases: the contribution of redox proteomics. *Mass Spectrom Rev* 2005;24(1):55-99.
 19. Hazen SL, Hsu FF, Gaut JP, Crowley JR, Heinecke JW. Modification of proteins and lipids by myeloperoxidase. *Methods Enzymol* 1999;300:88-105.
 20. Butterfield DA. Proteomics: a new approach to investigate oxidative stress in Alzheimer's disease brain. *Brain Res* 2004;1000:1-7.
 21. Davies MJ. The oxidative environment and protein damage. *Biochim Biophys Acta* 2005;1703(2):93-109.
 22. Fu S-L, Dean RT. Structural characterization of the products of hydroxyl-radical damage to leucine and their detection on proteins. *Biochem J* 1997;324:41-8.
 23. Beckman JS. Oxidative damage and tyrosine nitration from peroxynitrite. *Chem Res Toxicol* 1996;9:836-44.
 24. Ma Y-S, Chao C-C, Stadtman ER. Oxidative modification of glutamine synthetase by 2,2'-azobis(2-amidinopropane) dihydrochloride. *Arch Biochem Biophys* 1999;363:129-34.
 25. Schoneich C. Methionine oxidation by reactive oxygen species: reaction mechanisms and relevance to Alzheimer's disease. *Biochim Biophys Acta* 2005;1703(2):111-9.
 26. Boschi-Muller S, Olry A, Antoine M, Branlant G. The enzymology and biochemistry of methionine sulfoxide reductases. *Biochim Biophys Acta* 2005;1703(2):231-8.
 27. Moskovitz J. Methionine sulfoxide reductases: ubiquitous enzymes involved in antioxidant defense, protein regulation, and prevention of aging-associated diseases. *Biochim Biophys Acta* 2005;1703(2):213-9.
 28. Stadtman ER, Moskovitz J, Levine RL. Oxidation of methionine residues of proteins: biological consequences. *Antioxid Redox Signal* 2003;5(5):577-82.
 29. Malencik DA, Anderson SR. Dityrosine as a product of oxidative stress and fluorescent probe. *Amino Acids* 2003;25(3-4):233-47.
 30. Uchida K, Kawakishi S. Reactions of a histidyl residue analogue with hydrogen peroxide in the presence of copper(II) ion. *J Agric Food Chem* 1990;38:660-4.
 31. Zhao F, Ghezzi-Schoneich E, Aced GL, Hong J, Milby T, Schoneich C. Metal-catalyzed oxidation of histidine in human growth hormone. *J Biol Chem* 1997;272:9019-29.
 32. Huggins TG, Wells-Knecht MC, Detorie NA, Baynes JW, Thorpe SR. Formation of *o*-tyrosine and dityrosine in proteins during radiolytic and metal-catalyzed oxidation. *J Biol Chem* 1003;268:12341-7.
 33. Buranaprapuk A, Leach SP, Kumar CV, Bocarsly JR. Protein cleavage by transition metal complexes bearing amino acid substituents. *Biochim Biophys Acta* 1998;1387:309-16.
 34. Saxena AK, Saxena P, Wu X, Obrenovich M, Weiss MF, Monnier VM. Protein aging by carboxymethylation of lysines generates sites for divalent metal and redox active copper binding: relevance to diseases of glycoxidative stress. *Biochem Biophys Res Commun* 1999;260:332-8.
 35. Requena JR, Stadtman ER. Conversion of lysine to N^ε-(carboxymethyl)lysine increases susceptibility of proteins to metal-catalyzed oxidation. *Biochem Biophys Res Commun* 1999;264:207-11.
 36. Sayre LM, Perry G, Harris PLR, Liu Y, Schubert KA, Smith MA. In situ oxidative catalysis by neurofibrillary tangles and senile plaques in Alzheimer disease: a central role for bound transition metals. *J Neurochem* 2000;74:270-9.
 37. Nagai R, Horiuchi S, Unno Y. Application of monoclonal antibody libraries for the measurement of glycation adducts. *Biochem Soc Trans* 2003;31(Pt 6):1438-40.
 38. Ikeda K, Higashi T, Sano H, Jinnouchi Y, Yoshida M, Araki T, Ueda S, Horiuchi S. N^ε-(Carboxymethyl)lysine protein adduct is a major immunological epitope in proteins modified with advanced glycation end products of the Maillard reaction. *Biochemistry* 1996;35:8075-83.
 39. Fu M-X, Requena JR, Jenkins AJ, Lyons TJ, Baynes JW, Thorpe SR. The advanced glycation end product, N^ε-(carboxymethyl)lysine, is a product of both lipid peroxidation and glycoxidation reactions. *J Biol Chem* 1996;271:9982-6.
 40. Wells-Knecht KJ, Brinkmann E, Baynes JW. Characterization of an imidazolium salt formed from glyoxal and N^ε-hippuryllysine: a model for Maillard reaction crosslinks in proteins. *J Org Chem* 1995;60:6246-7.
 41. Glomb MA, Pfahler C. Amides are novel protein modifications formed by physiological sugars. *J Biol Chem* 2001;276(45):41638-47.
 42. Liu Y. *Modification of proteins by lysine-directed bifunctional aldehydes derived from lipid peroxidation*. Ph.D. Thesis, Case Western Reserve University, Cleveland OH, USA, 2003.
 43. Ahmed MU, Brinkmann Frye E, Degenhardt TP, Thorpe SR, Baynes JW. N-epsilon-(carboxyethyl)lysine, a product of the chemical modification of proteins by methylglyoxal, increases with age in human lens proteins. *Biochem J* 1997;324:565-70.
 44. Brinkmann E, Wells-Knecht KJ, Thorpe SR, Baynes JW. Characterization of an imidazolium compound formed by reaction of methylglyoxal and N^ε-hippuryllysine. *J Chem Soc Perkin Trans 1* 1995;2817-8.
 45. Pratico D, Lee VMY, Trojanowski JQ, Rokach J, Fitzgerald GA. Increased F2-isoprostanes in Alzheimer's disease: evidence for enhanced lipid peroxidation *in vivo*. *FASEB J* 1998;12:1777-83.
 46. Noorooz-Zadeh J, Liu EHC, Yhlen B, Anggard EE, Halliwell B. F₂-isoprostanes as specific marker of docosahexaenoic acid peroxidation in Alzheimer's disease. *J Neurochem* 1999;72: 734-40.

47. Greco A, Minghetti L, Levi G. Isoprostanes, novel markers of oxidative injury, help understanding the pathogenesis of neurodegenerative diseases. *Neurochem Res* 2000;25(9-10):1357-64.
48. Montine KS, Quinn JF, Zhang J, Fessel JP, Roberts LJ 2nd, Morrow JD, Montine TJ. Isoprostanes and related products of lipid peroxidation in neurodegenerative diseases. *Chem Phys Lipids* 2004;128:117-24.
49. Lee SH, Oe T, Blair IA. Vitamin C-induced decomposition of lipid hydroperoxides to endogenous genotoxins. *Science* 2001;292(5524):2083-6.
50. Petersen DR, Doorn JA. Reactions of 4-hydroxynonenal with proteins and cellular targets. *Free Radic Biol Med* 2004;37(7):937-45.
51. Esterbauer H, Schaur RJ, Zollner H. Chemistry and biochemistry of 4-hydroxynonenal, malondialdehyde and related aldehydes. *Free Radic Biol Med* 1991;11:81-128.
52. Requena JR, Fu MX, Ahmed MU, Jenkins AJ, Lyons TJ, Baynes JW, Thorpe SR. Quantification of malondialdehyde and 4-hydroxynonenal adducts to lysine residues in native and oxidized human low-density lipoprotein. *Biochem J* 1997;322:317-25.
53. Slatter DA, Murray M, Bailey AJ. Formation of a dihydropyridine derivative as a potential cross-link derived from malondialdehyde in physiological systems. *FEBS Lett* 1998;421:180-4.
54. Itakura K, Uchida K. Evidence that malondialdehyde-derived aminoenimine is not a fluorescent age pigment. *Chem Res Toxicol* 2001;14(5):473-5.
55. Uchida K, Sakai K, Itakura K, Osawa T, Toyokuni S. Protein modification by lipid peroxidation products: formation of malondialdehyde-derived N^ε-(2-propenal)lysine in proteins. *Arch Biochem Biophys* 1997;346:45-52.
56. Kikugawa K, Beppu M. Involvement of lipid oxidation products in the formation of fluorescent and cross-linked proteins. *Chem Phys Lipids* 1987;44:277-6.
57. Nair V, Offerman RJ, Turner GA, Pryor AN, Baenziger NC. Fluorescent 1,4-dihydropyridines: the malondialdehyde connection. *Tetrahedron* 1988;44:2793-803.
58. Xu D, Thiele GM, Kearley ML, Haugen MD, Klassen LW, Sorrell MF, Tuma DJ. Epitope characterization of malondialdehyde-acetaldehyde adducts using an enzyme-linked immunosorbent assay. *Chem Res Toxicol* 1997;10:978-86.
59. Itakura K, Uchida K, Osawa T. A novel fluorescent malondialdehyde-lysine adduct. *Chem Phys Lipids* 1996;84:75-9.
60. Slatter DA, Avery NC, Bailey AJ. Identification of a new cross-link and unique histidine adduct from bovine serum albumin incubated with malondialdehyde. *J Biol Chem* 2004;279(1):61-9.
61. Uchida K, Kanematsu M, Sakai K, Matsuda T, Hattori N, Mizuno Y, Suzuki D, Miyata T, Noguchi N, Niki E, Osawa T. Protein-bound acrolein: potential markers for oxidative stress. *Proc Natl Acad Sci U S A* 1998;95:4882-7.
62. Calingasan NY, Uchida K, Gibson GE. Protein-bound acrolein: a novel marker of oxidative stress in Alzheimer's disease. *J Neurochem* 1999;72:751-6.
63. Furuhashi A, Ishii T, Kumazawa S, Yamada T, Nakayama T, Uchida K. N^ε-(3-methylpyridinium)lysine, a major antigenic adduct generated in acrolein-modified protein. *J Biol Chem* 2003;278:48658-65.
64. Liu Z, Sayre LM. Model studies on the modification of proteins by lipoxidation-derived 2-hydroxyaldehydes. *Chem Res Toxicol* 2003;16:232-41.
65. Itakura K, Furuhashi A, Shibata N, Kobayashi M, Uchida K. Maillard reaction-like lysine modification by a lipid peroxidation product: immunochemical detection of protein-bound 2-hydroxyheptanal *in vivo*. *Biochem Biophys Res Commun* 2003;308:452-7.
66. Uchida K. 4-Hydroxy-2-nonenal: a product and mediator of oxidative stress. *Prog Lipid Res* 2003;42:318-43.
67. Uchida K, Stadtman ER. Modification of histidine residues in proteins by reaction with 4-hydroxynonenal. *Proc Natl Acad Sci U S A* 1992;89:4544-8.
68. Uchida K, Szveda LI, Chae H-Z, Stadtman ER. Immunochemical detection of 4-hydroxynonenal protein adducts in oxidized hepatocytes. *Proc Natl Acad Sci U S A* 1993;90:8742-6.
69. Uchida K, Itakura K, Kawakishi S, Hiai H, Toyokuni S, Stadtman ER. Characterization of epitopes recognized by 4-hydroxy-2-nonenal specific antibodies. *Arch Biochem Biophys* 1995;324:241-8.
70. Nadkarni DV, Sayre LM. Structural definition of early lysine and histidine adduction chemistry of 4-hydroxynonenal. *Chem Res Toxicol* 1995;8:284-91.
71. Sayre LM, Arora PK, Iyer RS, Salomon RG. Pyrrole formation from 4-hydroxynonenal and primary amines. *Chem Res Toxicol* 1993;6:19-22.
72. Sayre LM, Sha W, Xu G, Kaur K, Nadkarni D, Subbanagounder G, Salomon RG. Immunochemical evidence supporting 2-pentylpyrrole formation on proteins exposed to 4-hydroxy-2-nonenal. *Chem Res Toxicol* 1996;9:1194-201.
73. Uchida K, Stadtman ER. Covalent attachment of 4-hydroxynonenal to glyceraldehyde-3-phosphate dehydrogenase. *J Biol Chem* 1993;268:6388-93.
74. Friguet B, Stadtman ER, Szveda LI. Modification of glucose-6-phosphate dehydrogenase by 4-hydroxynonenal. *J Biol Chem* 1994;269:21639-43.
75. Cohn JA, Tsai L, Friguet B, Szveda LI. Chemical characterization of a protein-4-hydroxy-2-nonenal cross-link: immunochemical detection in mitochondria exposed to oxidative stress. *Arch Biochem Biophys* 1996;328:158-64.
76. Montine TJ, Amarnath V, Martin ME, Strittmatter WJ, Graham DG. E-4-Hydroxy-2-nonenal is cytotoxic and crosslinks cytoskeletal proteins in P19 neuroglial cultures. *Am J Pathol* 1996;148:89-93.
77. Montine TJ, Huang DY, Valentine WM, Amarnath V, Saunders A, Weisgraber KH, Graham DG, Strittmatter WJ. Crosslinking of apolipoprotein E by products of lipid peroxidation. *J Neuropathol Exp Neurol* 1996;55:202-10.
78. Xu G, Sayre LM. Structural characterization of a 4-hydroxy-2-alkenal-derived fluorophore that contributes to lipoperoxidation-dependent protein crosslinking in aging and degenerative disease. *Chem Res Toxicol* 1998;11:247-51.

79. Itakura K, Osawa T, Uchida K. Structure of a fluorescent compound formed from 4-hydroxy-2-nonenal and *N*^ε-hippuryllysine: A model for fluorophores derived from protein modification by lipid peroxidation. *J. Org. Chem* 1998;63:185-7.
80. Tsai L, Szveda PA, Vinogradova O, Szveda LI. Structural characterization and immunochemical detection of a fluorophore derived from 4-hydroxy-2-nonenal and lysine. *Proc Natl Acad Sci U S A* 1998;95:7975-80.
81. Xu G, Liu Y, Sayre LM. Independent synthesis, solution behavior, and studies on the mechanism of formation of a primary amine-derived fluorophore representing crosslinking of proteins by (*E*)-4-hydroxy-2-nonenal (HNE). *J Org Chem* 1999;64:5732-45.
82. Xu G, Liu Y, Kansal MM, Sayre LM. Rapid crosslinking of proteins by 4-ketoaldehydes and 4-hydroxy-2-alkenals does not arise from the lysine-derived monoalkylpyrroles. *Chem Res Toxicol* 1999;12:855-61.
83. Montine KS, Kim PJ, Olson SJ, Markesbery WR, Montine TJ. 4-Hydroxy-2-nonenal pyrrole adducts in human neurodegenerative disease. *J Neuropathol Exp Neurol* 1997;56:866-71.
84. Montine KS, Olson SJ, Amarnath V, Whetsell WO Jr, Graham DG, Montine TJ. Immunohistochemical detection of 4-hydroxy-2-nonenal adducts in Alzheimer's disease is associated with inheritance of *APOE4*. *Am J Pathol* 1997;150:437-43.
85. Montine KS, Reich E, Neely MD, Sidell KR, Olson SJ, Markesbery WR, Montine TJ. Distribution of reducible 4-hydroxynonenal adduct immunoreactivity in Alzheimer disease is associated with *APOE* genotype. *J Neuropathol Exp Neurol* 1998;57:415-25.
86. Sayre LM, Zelasko DA, Harris PL, Perry G, Salomon RG, Smith MA. 4-Hydroxynonenal-derived advanced lipid peroxidation end products are increased in Alzheimer's disease. *J Neurochem* 1997;68:2092-7.
87. Ando Y, Brannstrom T, Uchida K, Nyhlin N, Nasman B, Suhr O, Yamashita T, Olsson T, El Sahly M, Uchino M, Ando M. Histochemical detection of 4-hydroxynonenal protein in Alzheimer amyloid. *J Neurol Sci* 1998;156:172-76.
88. Yoritaka A, Hattori N, Uchida K, Tanaka M, Stadtman ER, Mizuno Y. Immunohistochemical detection of 4-hydroxynonenal protein adducts in Parkinson disease. *Proc Natl Acad Sci U S A* 1996;93:2696-701.
89. Castellani RJ, Perry G, Siedlak SL, Nunomura A, Shimohama S, Zhang J, Montine T, Sayre LM, Smith MA. Hydroxynonenal adducts indicate a role for lipid peroxidation in neocortical and brainstem Lewy bodies in humans. *Neurosci Lett* 2002;319:25-8.
90. Pedersen WA, Fu W, Keller JN, Markesbery WR, Appel S, Smith RG, Kasarskis E, Mattson MP. Protein modification by the lipid peroxidation product 4-hydroxynonenal in the spinal cords of amyotrophic lateral sclerosis patients. *Ann Neurol* 1998;44:819-24.
91. Ando Y, Nyhlin N, Suhr O, Holmgren G, Uchida K, El Sahly M, Yamashita T, Terasaki H, Nakamura M, Uchino M, Ando M. Oxidative stress is found in amyloid deposits in systemic amyloidosis. *Biochem Biophys Res Commun* 1997;232:497-502.
92. Castellani RJ, Perry G, Harris PL, Cohen ML, Sayre LM, Salomon RG, Smith MA. Advanced lipid peroxidation end-products in Alexander's disease. *Brain Res* 1998;787:15-8.
93. Lee SH, Blair IA. Characterization of 4-oxo-2-nonenal as a novel product of lipid peroxidation. *Chem Res Toxicol* 2000;13:698-702.
94. Spiteller P, Kern W, Reiner J, Spiteller G. Aldehydic lipid peroxidation products derived from linoleic acid. *Biochim Biophys Acta* 2001;1531:188-208.
95. Zhang WH, Liu J, Xu G, Yuan Q, Sayre LM. Model studies on protein side chain modification by 4-oxo-2-nonenal. *Chem Res Toxicol* 2003;16:512-23.
96. Liu Z, Minkler PE, Sayre LM. Mass spectroscopic characterization of protein modification by 4-hydroxy-2-(*E*)-nonenal and 4-oxo-2-(*E*)-nonenal. *Chem Res Toxicol* 2003;16(7):901-11.
97. Mattana J, Margiloff L, Singhal PC. Metal-catalyzed oxidation of extracellular matrix proteins disrupts integrin-mediated adhesion of mesangial cells. *Biochem Biophys Res Commun* 1997;233:50-55.
98. Yan L-J, Lodge JK, Traber MG, Packer L. Apolipoprotein B carbonyl formation is enhanced by lipid peroxidation during copper-mediated oxidation of human low-density lipoprotein. *Arch Biochem Biophys* 1997;339:165-71.
99. Gotz J, Schild A, Hoernkli F, Pennanen L. Amyloid-induced neurofibrillary tangle formation in Alzheimer's disease: insight from transgenic mouse and tissue-culture models. *Int J Dev Neurosci* 2004;22(7):453-65.
100. German DC, Eisch AJ. Mouse models of Alzheimer's disease: insight into treatment. *Rev Neurosci* 2004;15(5):353-69.
101. Selkoe DJ. Defining molecular targets to prevent Alzheimer disease. *Arch Neurol* 2005;62(2):192-5.
102. Bishop GM, Robinson SR. Physiological roles of amyloid-beta and implications for its removal in Alzheimer's disease. *Drugs Aging* 2004;21(10):621-30.
103. Kim HJ, Chae SC, Lee DK, Chromy B, Lee SC, Park YC, Klein WL, Krafft GA, Hong ST. Selective neuronal degeneration induced by soluble oligomeric amyloid beta protein. *FASEB J* 2003;17:118-20.
104. Kirkitadze MD, Bitan G, Teplow DB. Paradigm shifts in Alzheimer's disease and other neurodegenerative disorders: the emerging role of oligomeric assemblies. *J Neurosci Res* 2002;69(5):567-77.
105. Butterfield DA, Boyd-Kimball D. Amyloid beta-peptide(1-42) contributes to the oxidative stress and neurodegeneration found in Alzheimer disease brain. *Brain Pathol* 2004;14(4):426-32.
106. King ME. Can tau filaments be both physiologically beneficial and toxic? *Biochim Biophys Acta* 2005;1739(2-3):260-7.
107. Buee L, Bussiere T, Buee-Scherrer V, Delacourte A, Hof PR. Tau protein isoforms, phosphorylation and role in neurodegenerative disorders. *Brain Res Brain Res Rev* 2000;33:95-130.
108. Brandt R, Leger J, Lee G. Interaction of tau with the neural plasma membrane mediated by tau's amino-terminal projection domain. *J Cell Biol* 1995;131:1327-40.
109. Xie H-q, Litersky JM, Hartigan JA, Jope RS, Johnson GVW. The interrelationship between selective tau phosphorylation and microtubule association. *Brian Res* 1998;798:173-83.
110. Sengupta A, Kabat J, Novak M, Wu Q, Grundke-Iqbal I, Iqbal K. Phosphorylation of tau at both Thr 231 and Ser 262 is required for maximal inhibition of its binding to microtubules. *Arch Biochem Biophys* 1998;357:299-309.

111. Hanger DP, Betts JC, Loviny TLF, Blackstock WP, Anderton BH. New phosphorylation sites identified in hyperphosphorylated tau (paired helical filament-tau) from Alzheimer's disease brain using nanoelectrospray mass spectrometry. *J Neurochem* 1998;71:2465-76.
112. Stoothoff WH, Johnson GV. Tau phosphorylation: physiological and pathological consequences. *Biochim Biophys Acta* 2005;1739:280-97.
113. Binder LI, Guillozet-Bongaarts AL, Garcia-Sierra F, Berry RW. Tau, tangles, and Alzheimer's disease. *Biochim Biophys Acta* 2005;1739:216-23.
114. Nuydens R, De Jong M, Nuyens R, Cornelissen F, Geerts H. Neuronal kinase stimulation leads to aberrant tau phosphorylation and neurotoxicity. *Neurobiol Aging* 1995;16:465-75.
115. Iqbal K, Zaidi T, Bancher C, Grundke-Iqbal I. Alzheimer paired helical filaments. Restoration of the biological activity by dephosphorylation. *FEBS Lett* 1994;349:104-8.
116. Iqbal K, Alonso Adel C, Chen S, Chohan MO, El-Akkad E, Gong CX, Khatoon S, Li B, Liu F, Rahman A, Tanimukai H, Grundke-Iqbal I. Tau pathology in Alzheimer disease and other tauopathies. *Biochim Biophys Acta* 2005;1739:198-210.
117. Kosik KS, Shimura H. Phosphorylated tau and the neurodegenerative foldopathies. *Biochim Biophys Acta* 2005;1739:298-310.
118. Fath T, Eidenmuller J, Brandt RB. Tau-mediated cytotoxicity in a pseudohyperphosphorylation model of Alzheimer's disease. *J Neurosci* 2002;22:9733-41.
119. Keck S, Nitsch R, Grune T, Ullrich O. Proteasome inhibition by paired helical filament-tau in brains of patients with Alzheimer's disease. *J Neurochem* 2003;85:115-22.
120. Miller ML, Johnson GVW. Transglutaminase cross-linking of the τ protein. *J Neurochem* 1995;65:1760-70.
121. Nemes Z, Devreese B, Steinert PM, Van Beeumen J, Fesus L. Cross-linking of ubiquitin, HSP27, parkin, and alpha-synuclein by gamma-glutamyl-epsilon-lysine bonds in Alzheimer's neurofibrillary tangles. *FASEB J* 2004;18:1135-7.
122. Perry G, Raina AK, Nunomura A, Wataya T, Sayre LM, Smith MA. How important is oxidative damage? Lessons from Alzheimer's disease. *Free Radic Biol Med* 2000;28:831-4.
123. Ko L-w, Ko EC, Nacharaju P, Liu W-K, Chang E, Kenessey A, Yen S-HC. An immunochemical study on tau glycation in paired helical filaments. *Brain Res* 1999;830:301-13.
124. Liu Q, Smith MA, Avila J, DeBernardis J, Kansal M, Takeda A, Zhu X, Nunomura A, Honda K, Moreira PI, Oliveira CR, Santos MS, Shimohama S, Aliev G, de la Torre J, Ghanbari HA, Siedlak SL, Harris PL, Sayre LM, Perry G. Alzheimer-specific epitopes of tau represent lipid peroxidation-induced conformations. *Free Radic Biol Med* 2005;38(6):746-54.
125. Lovell MA, Robertson JD, Teesdale WJ, Campbell JL, Markesbery WR. Copper, iron and zinc in Alzheimer's disease senile plaques. *J Neurol Sci* 1998;158:47-52.
126. Connor JR, Milward EA, Moalem S, Sampietro M, Boyer P, Percy ME, Vergani C, Scott RJ, Chorney M. Is hemochromatosis a risk factor for Alzheimer's disease? *J Alzheimers Dis* 2001;3:471-7.
127. Perez M, Valpuesta JM, de Garcini EM, Quintana C, Arrasate M, Lopez Carrascosa JL, Rabano A, Garcia de Yébenes J, Avila J. Ferritin is associated with the aberrant tau filaments present in progressive supranuclear palsy. *Am J Pathol* 1998;152:1531-9.
128. Castellani RJ, Smith MA, Nunomura A, Harris PLR, Perry G. Is increased redox-active iron in Alzheimer disease a failure of the copper-binding protein ceruloplasmin? *Free Radic Biol Med* 1999;26:1508-12.
129. Smith MA, Wehr K, Harris PLR, Siedlak SL, Connor JR, Perry G. Abnormal localization of iron regulatory protein (IRP) in Alzheimer disease. *Brain Res* 1998;788:232-6.
130. Smith MA, Kutty RK, Richey PL, Yan S-D, Stern D, Chader GJ, Wiggert B, Petersen RB, Perry G. Heme oxygenase-1 is associated with the neurofibrillary pathology of Alzheimer disease. *Am J Pathol* 1994;145:42-7.
131. Premkumar DRD, Smith MA, Richey PL, Petersen RB, Castellani R, Kutty RK, Wiggert B, Perry G, Kalara RN. Induction of heme oxygenase-1 mRNA and protein in neocortex and cerebral vessels in Alzheimer's disease. *J Neurochem* 1995;65:1399-402.
132. Honda K, Casadesus G, Petersen RB, Perry G, Smith MA. Oxidative stress and redox-active iron in Alzheimer's disease. *Ann NY Acad Sci* 2004;1012:179-82.
133. Good PF, Perl DP, Bierer LM, Schmeidler J. Selective accumulation of aluminum and iron in the neurofibrillary tangles of Alzheimer's disease: a laser microprobe (LAMMA) study. *Ann Neurol* 1992;31:286-92.
134. Oteiza PI. A mechanism for the stimulatory effect of aluminum on iron-induced lipid peroxidation. *Arch Biochem Biophys* 1994;308:374-9.
135. Smith MA, Harris PLR, Sayre LM, Perry G. Iron accumulation in Alzheimer disease is a source of redox-generated free radicals. *Proc Natl Acad Sci USA* 1997;94:9866-8.
136. Mantyh PW, Ghilardi JR, Rogers S, DeMaster E, Allen CJ, Stimson ER, Maggio JE. Aluminum, iron, and zinc ions promote aggregation of physiological concentrations of beta-amyloid peptide. *J Neurochem* 1993;61:1171-4.
137. Atwood CS, Moir RD, Huang X, Scarpa RC, Bacarra NME, Romano DM, Hartshorn MA, Tanzi RE, Bush AI. Dramatic aggregation of Alzheimer A β by Cu(II) is induced by conditions representing physiological acidosis. *J Biol Chem* 1998;273:12817-26.
138. Cherny RA, Legg JT, McLean CA, Fairlic DP, Huang X, Atwood CS, Beyreuther K, Tanzi RE, Masters CL, Bush AI. Aqueous dissolution of Alzheimer's disease A β amyloid deposits by biometal depletion. *J Biol Chem* 1999;274:23223-8.
139. Dong J, Atwood CS, Anderson VE, Siedlak SL, Smith MA, Perry G, Carey PR. Metal binding and oxidation of amyloid- β within isolated senile plaque cores: Raman microscopic evidence. *Biochemistry* 2003;42:2768-73.
140. Bishop GM, Robinson SR. The amyloid paradox: amyloid-beta-metal complexes can be neurotoxic and neuroprotective. *Brain Pathol* 2004;14(4):448-52.
141. Bondy SC, Guo-Ross SX, Truong AT. Promotion of transition metal-induced reactive oxygen species formation by β -amyloid. *Brain Res* 1998;799:91-6.

142. Yang EY, Guo-Ross SX, Bondy SC. The stabilization of ferrous iron by a toxic β -amyloid fragment and by an aluminum salt. *Brain Res* 1999;839:221-6.
143. Huang X, Atwood CS, Hartshorn MA, Multhaup G, Goldstein LE, Scarpa RC, Cuajungco MP, Gray DN, Lim J, Moir RD, Tanzi RE, Bush AI. The A β peptide of Alzheimer's disease directly produces hydrogen peroxide through metal ion reduction. *Biochemistry* 1999;38:7609-16.
144. House E, Collingwood J, Khan A, Korchazkina O, Berthon G, Exley C. Aluminium, iron, zinc and copper influence the in vitro formation of amyloid fibrils of Abeta42 in a manner which may have consequences for metal chelation therapy in Alzheimer's disease. *J Alzheimer Dis* 2004;6(3):291-301.
145. Bayer TA, Schafer S, Simons A, Kemmling A, Kamer T, Tepest R, Eckert A, Schussel K, Eikenberg O, Sturchler-Pierrat C, Abramowski D, Staufenbiel M, Multhaup G. Dietary Cu stabilizes brain superoxide dismutase 1 activity and reduces amyloid Abeta production in APP23 transgenic mice. *Proc Natl Acad Sci U S A* 2003;100:14187-92.
146. Sparks DL, Schreurs BG. Trace amounts of copper in water induce beta-amyloid plaques and learning deficits in a rabbit model of Alzheimer's disease. *Proc Natl Acad Sci U S A* 2003;100:11065-9.
147. Bellingham SA, Lahiri DK, Maloney B, La Fontaine S, Multhaup G, Camakaris J. Copper depletion down-regulates expression of the Alzheimer's disease amyloid-beta precursor protein gene. *J Biol Chem* 2004;279(19):20378-86.
148. Multhaup G, Ruppert T, Schlicksupp A, Hesse L, Bill E, Pipkorn R, Masters CL, Beyreuther K. Copper-binding amyloid precursor undergoes a site-specific fragmentation in the reduction of hydrogen peroxide. *Biochemistry* 1998;37:7224-30.
149. Multhaup G, Ruppert T, Schlicksupp A, Hesse L, Behr D, Masters CL, Beyreuther K. Reactive oxygen species and Alzheimer's disease. *Biochem Pharmacol* 1997;54:533-59.
150. Rogers JT, Randall JD, Cahill CM, Eder PS, Huang X, Gunshin H, Leiter L, McPhee J, Sarang SS, Utsuki T, Greig NH, Lahiri DK, Tanzi RE, Bush AI, Giordano T, Gullans SR. An iron-responsive element type II in the 5'-untranslated region of the Alzheimer's amyloid precursor protein transcript. *J Biol Chem* 2002;277:45518-28.
151. Sayre LM, Perry G, Smith MA. *In situ* methods for detection and localization of markers of oxidative stress: application in neurodegenerative disorders. *Methods Enzymol* 1999;309:133-52.
152. Gabbita SP, Lovell MA, Markesbery WR. Increased nuclear DNA oxidation in the brain in Alzheimer's disease. *J Neurochem* 1998;71:2034-40.
153. Lovell MA, Gabbita SP, Markesbery WR. Increased DNA oxidation and decreased levels of repair products in Alzheimer's disease ventricular CSF. *J Neurochem* 1999;72:771-6.
154. Smith MA, Perry G, Richey PL, Sayre LM, Anderson VE, Beal MF, Kowall N. Oxidative damage in Alzheimer's. *Nature* 1996;382:120-1.
155. Smith MA, Harris PLR, Sayre LM, Beckman JS, Perry G. Widespread peroxynitrite-mediated damage in Alzheimer's disease. *J Neurosci* 1997;17:2653-7.
156. Hensley K, Maitt ML, Yu Z, Sang H, Markesbery WR, Floyd RA. Electrochemical analysis of protein nitrotyrosine and dityrosine in the Alzheimer brain indicates region-specific accumulation. *J Neurosci* 1998;18:8126-32.
157. Keller JN, Schmitt FA, Scheff SW, Ding Q, Chen Q, Butterfield DA, Markesbery WR. Evidence of increased oxidative damage in subjects with mild cognitive impairment. *Neurology* 2005;64(7):1152-6.
158. Smith MA, Taneda S, Richey PL, Miyata S, Yan S-D, Stern D, Sayre LM, Monnier VM, Perry G. Advanced Maillard reaction end products are associated with Alzheimer disease pathology. *Proc Natl Acad Sci U S A* 1994;91:5710-4.
159. Smith MA, Rudnicka-Nawrot M, Richey PL, Praprotnik D, Mulvihill P, Miller CA, Sayre LM, Perry G. Carbonyl-related posttranslational modification of neurofilament protein in the neurofibrillary pathology of Alzheimer's disease. *J Neurochem* 1995;64:2660-6.
160. Smith MA, Sayre LM, Monnier VM, Perry G. Radical AGEing in Alzheimer's disease. *Trends Neurosci* 1995;18:172-6.
161. Vitek MP, Bhattacharya K, Glendening JM, Stopa E, Vlassara H, Bucala R, Manogue K, Cerami A. Advanced glycation end products contribute to amyloidosis in Alzheimer disease. *Proc Natl Acad Sci U S A* 1994;91:4766-70.
162. Yan S-D, Chen X, Schmidt A-M, Brett J, Godman G, Zou Y-S, Scott C W, Caputo C, Frappier T, Smith M A, Perry G, Yen S-H, Stern D. Glycated τ protein in Alzheimer disease: a mechanism for induction of oxidant stress. *Proc Natl Acad Sci U S A* 1994;91:7787-91.
163. Ledesma MD, Bonay P, Colaco C, Avila J. Analysis of microtubule-associated protein τ glycation in paired helical filaments. *J Biol Chem* 1994;269:21614-9.
164. Castellani RJ, Harris PL, Sayre LM, Fujii J, Taniguchi N, Vitek MP, Founds H, Atwood CS, Perry G, Smith MA. Active glycation in neurofibrillary pathology of Alzheimer disease: N(epsilon)-(carboxymethyl) lysine and hexitol-lysine. *Free Radic Biol Med* 2001;31:175-80.
165. Cras P, Smith MA, Richey PL, Siedlak SL, Mulvihill P, Perry G. Extracellular neurofibrillary tangles reflect neuronal loss and provide further evidence of extensive protein cross-linking in Alzheimer disease. *Acta Neuropathol (Berl)* 1995;89:291-5.
166. Smith MA. Alzheimer disease. *Int Rev Neurobiol* 1998;42:1-54.
167. Sullivan PG, Brown MR. Mitochondrial aging and dysfunction in Alzheimer's disease. *Prog Neuropsychopharmacol Biol Psychiatry* 2005;29(3):407-10.
168. Castellani RJ, Honda K, Zhu X, Cash AD, Nunomura A, Perry G, Smith MA. Contribution of redox-active iron and copper to oxidative damage in Alzheimer disease. *Ageing Res Rev* 2004;3(3):319-26.
169. Schipper HM, Cisse S, Stopa EG. Expression of heme oxygenase-1 in the senescent and Alzheimer-diseased brain. *Ann Neurol* 1995;37:758-68.
170. Bonilla E, Tanji K, Hirano M, Vu TH, DiMauro S, Schon EA. Mitochondrial involvement in Alzheimer's disease. *Biochim Biophys Acta* 1999;1410:171-82.
171. Aliev G, Seyidova D, Neal ML, Shi J, Lamb BT, Siedlak SL, Vinters HV, Head E, Perry G, Lamanna JC, Friedland RP, Cotman CW. Atherosclerotic lesions and mitochondria DNA deletions in brain microvessels as a central target for the development of human AD and AD-like pathology in aged transgenic mice. *Ann N Y Acad Sci* 2002;977:45-64.

172. Pappolla MA, Omar RA, Kim KS, Robakis NK. Immunohistochemical evidence of oxidative stress in Alzheimer's disease. *Am J Pathol* 1992;140:621-8.
173. De Leo ME, Borrello S, Passantino M, Palazzotti B, Mordente A, Daniele A, Filippini V, Galeotta T, Masullo C. Oxidative stress and overexpression of manganese superoxide dismutase in patients with Alzheimer's disease. *Neurosci Lett* 1998;250:173-6.
174. Martins RN, Harper CG, Stokes GB, Masters CL. Increased cerebral glucose-6-phosphate dehydrogenase activity in Alzheimer's disease may reflect oxidative stress. *J Neurochem* 1986;46:1042-5.
175. Russell RL, Siedlak SL, Raina AK, Bautista JM, Smith MA, Perry G. Increased neuronal glucose-6-phosphate dehydrogenase and sulfhydryl levels indicate reductive compensation to oxidative stress in Alzheimer disease. *Arch Biochem Biophys* 1999;370:236-9.
176. Raina AK, Templeton DJ, Deak JC, Perry G, Smith MA. Quinone reductase (NQO1), a sensitive redox indicator, is increased in Alzheimer's disease. *Redox Report* 1999;4:23-7.
177. Marcus DL, Thomas C, Rodriguez C, Simberkoff K, Tsai JS, Strafaci JA, Freedman ML. Increased peroxidation and reduced antioxidant enzyme activity in Alzheimer's disease. *Exp Neurol* 1989;150:40-4.
178. Perry G, Taddeo MA, Nunomura A, Zhu X, Zenteno-Savin T, Drew KL, Shimohama S, Avila J, Castellani RJ, Smith MA. Comparative biology and pathology of oxidative stress in Alzheimer and other neurodegenerative diseases: beyond damage and response. *Comp Biochem Physiol C Toxicol Pharmacol* 2002;133:507-13.
179. Zhu X, Raina AK, Lee HG, Casadesus G, Smith MA, Perry G. Oxidative stress signalling in Alzheimer's disease. *Brain Res* 2004;1000:32-9.
180. Morsch R, Simon W, Coleman PD. Neurons may live for decades with neurofibrillary tangles. *J Neuropathol Exp Neurol* 1999;58:188-97.
181. Wataya T, Nunomura A, Smith MA, Siedlak SL, Harris PL, Shimohama S, Swzeda LI, Kaminski MA, Avila J, Price DL, Cleveland DW, Sayre LM, Perry G. High molecular weight neurofilament proteins are physiological substrates of adduction by the lipid peroxidation product hydroxynonenal. *J Biol Chem* 2002;277:4644-8.
182. Nunomura A, Perry G, Hirai K, Aliev G, Takeda A, Chiba S, Smith MA. Neuronal RNA oxidation in Alzheimer's disease and Down's syndrome. *Ann NY Acad Sci* 1999;893:362-4.
183. Nunomura A, Perry G, Pappolla MA, Friedland RP, Hirai K, Chiba S, Smith MA. Neuronal oxidative stress precedes amyloid-beta deposition in Down syndrome. *J Neuropathol Exp Neurol* 2000;59(11):1011-7.
184. Smith MA, Nunomura A, Zhu X, Takeda A, Perry G. Metabolic, metallic, and mitotic sources of oxidative stress in Alzheimer disease. *Antioxid Redox Signal* 2000;2:413-20.
185. Jenner P. Oxidative stress in Parkinson's disease. *Ann Neurol* 2003;53(Suppl 3):S26-36.
186. Galloway PG, Mulvihill P, Perry G. Filaments of Lewy bodies contain insoluble cytoskeletal elements. *Am J Pathol* 1992;140:809-22.
187. Dawson TM, Dawson VL. Molecular pathways of neurodegeneration in Parkinson's disease. *Science* 2003;302:819-22.
188. Schapira AH. Causes of neuronal death in Parkinson's disease. *Adv Neurol* 2001;86:155-62.
189. Beal MF. Mitochondrial dysfunction and oxidative damage in Alzheimer's and Parkinson's diseases and coenzyme Q10 as a potential treatment. *J Bioenerg Biomembr* 2004;36(4):381-6.
190. Sanchez-Ramos J, Overvik E, Ames B. A marker of oxyradical-mediated DNA damage (8-hydroxy-2'-deoxyguanosine) is increased in nigro-striatum of Parkinson's disease brain. *Neurodegeneration* 1994;3:197-204.
191. Zhang J, Perry G, Smith MA, Robertson D, Olson SJ, Graham DG, Montine TJ. Parkinson's disease is associated with oxidative damage to cytoplasmic DNA and RNA in substantia nigra neurons. *Am J Pathol* 1999;154:1423-9.
192. Alam ZI, Daniel SE, Lees AJ, Marsden DC, Jenner P, Halliwell B. A generalized increase in protein carbonyls in the brain in Parkinson's but not incidental Lewy body disease. *J Neurochem* 1997;69:1326-9.
193. Floor E, Wetzel MG. Increased protein oxidation in human substantia nigra pars compacta in comparison with basal ganglia and prefrontal cortex measured with an improved dinitrophenylhydrazine assay. *J Neurochem* 1998;70:268-75.
194. Good PF, Hsu A, Werner P, Perl DP, Olanow CW. Protein nitration in Parkinson's disease. *J Neuropathol Exp Neurol* 1998;57:338-42.
195. Castellani R, Smith MA, Richey PL, Perry G. Glycooxidation and oxidative stress in Parkinson disease and diffuse Lewy body disease. *Brain Res* 1996;737:195-200.
196. Burke RE. Recent advances in research on Parkinson disease: synuclein and parkin. *Neurologist* 2004;10:75-81.
197. Zhou W, Hurlbert MS, Schaack J, Prasad KN, Freed CR. Overexpression of human α -synuclein causes dopamine neuron death in rat primary culture and immortalized mesencephalon-derived cells. *Brain Res* 2000;866:33-43.
198. Lee M, Hyun D, Halliwell B, Jenner P. Effect of the overexpression of wild-type or mutant alpha-synuclein on cell susceptibility to insult. *J Neurochem* 2001;76:998-1009.
199. Recchia A, Debetto P, Negro A, Guidolin D, Skaper SD, Giusti P. Alpha-synuclein and Parkinson's disease. *FASEB J* 2004;18:617-26.
200. Zhou W, Freed CR. Tyrosine-to-cysteine modification of human alpha-synuclein enhances protein aggregation and cellular toxicity. *J Biol Chem* 2004;279:10128-35.
201. Jensen PH, Hager H, Nielsen MS, Hojrup P, Gliemann J, Jakes R. α -Synuclein binds to tau and stimulates the protein kinase A-catalyzed tau phosphorylation of Ser 262 and 356. *J Biol Chem* 1999;274(36):25481-9.
202. Giasson BI, Forman MS, Higuchi M, Golbe LI, Graves CL, Kottbauer PT, Trojanowski JQ, Lee VM. Initiation and synergistic fibrillization of tau and alpha-synuclein. *Science* 2003;300:636-40.
203. Iseki E, Marui W, Kosaka K, Ueda K. Frequent coexistence of Lewy bodies and neurofibrillary tangles in the same neurons of patients with diffuse Lewy body disease. *Neurosci Lett* 1999;265:9-12.
204. Lee VM, Giasson BI, Trojanowski JQ. More than just two peas in a pod: common amyloidogenic properties of tau and alpha-synuclein in neurodegenerative diseases. *Trends Neurosci* 2004;27:129-34.
205. Paik SR, Shin H-J, Lee J-H, Chang C-S, Kim J. Copper(II)-induced self-oligomerization of α -synuclein. *Biochem J* 1999;340:821-8.

206. Hashimoto M, Hsu LJ, Xia Y, Takeda A, Sisk A, Sundsmo M, Masliah E. Oxidative stress induces amyloid-like aggregate formation of NACP/alpha-synuclein *in vitro*. *NeuroReport* 1999;10:717-21.
207. Hashimoto M, Takeda A, Hsu LJ, Takenouchi T, Masliah E. Role of cytochrome *c* as a stimulator of alpha-synuclein aggregation in Lewy body disease. *J Biol Chem* 1996;274:28849-52.
208. Giasson BI, Duda JE, Murray IV, Chen Q, Souza JM, Hurtig HI, Ischiropoulos H, Trojanowski JQ, Lee VM. Oxidative damage linked to neurodegeneration by selective alpha-synuclein nitration in synucleinopathy lesions. *Science* 2000;290:985-9.
209. Hodara R, Norris EH, Giasson BI, Mishizen-Eberz AJ, Lynch DR, Lee VM, Ischiropoulos H. Functional consequences of alpha-synuclein tyrosine nitration: diminished binding to lipid vesicles and increased fibril formation. *J Biol Chem* 2004;279:47746-53.
210. Hattori N, Mizuno Y. Pathogenetic mechanisms of parkin in Parkinson's disease. *Lancet* 2004;364:722-4.
211. McNaught KS, Olanow CW. Proteolytic stress: a unifying concept for the etiopathogenesis of Parkinson's disease. *Ann Neurol* 2003;53(Suppl 3):S73-84.
212. Hyun DH, Gray DA, Halliwell B, Jenner P. Interference with ubiquitination causes oxidative damage and increased protein nitration: implications for neurodegenerative diseases. *J Neurochem* 2004;90:422-30.
213. McNaught KS, Belzaira R, Isacson O, Jenner P, Olanow CW. Altered proteasomal function in sporadic Parkinson's disease. *Exp Neurol* 2003;179, 38-46.
214. Lindersson E, Beedholm R, Hojrup P, Moos T, Gai W, Hendil KB, Jensen PH. Proteasomal inhibition by alpha-synuclein filaments and oligomers. *J Biol Chem* 2004;279:12924-34.
215. Rideout HJ, Dietrich P, Wang Q, Dauer WT, Stefanis L. alpha-Synuclein is required for the fibrillar nature of ubiquitinated inclusions induced by proteasomal inhibition in primary neurons. *J Biol Chem* 2004;279:46915-20.
216. Eriksen JL, Wszolek Z, Petrucelli L. Molecular pathogenesis of Parkinson disease. *Arch Neurol* 2005;62(3):353-7.
217. Schipper HM. Astrocytes, brain aging, and neurodegeneration. *Neurobiol Aging* 1996;17:467-80.
218. Schipper HM, Vininsky R, Brull R, Small L, Brawer JR. Astrocyte mitochondria: a substrate for iron deposition in the aging rat substantia nigra. *Exp Neurol* 1998;152:188-96.
219. Riederer P, Sofic E, Rausch W-D, Schmidt B, Reynold GP, Jellinger K, Youdim MBH. Transition metals, ferritin, glutathione, and ascorbic acid in parkinsonian brains. *J Neurochem* 1989;52:515-20.
220. Pezzella A, d'Ischia M, Napolitano A, Misuraca G, Protta G. Iron-mediated generation of the neurotoxin 6-hydroxydopamine quinone by reaction of fatty acid hydroperoxides with dopamine: a possible contributory mechanism for neuronal degeneration in Parkinson's disease. *J Med Chem* 1997;40:2211-6.
221. Youdim MB, Stephenson G, Ben Shachar D. Ironing iron out in Parkinson's disease and other neurodegenerative diseases with iron chelators: a lesson from 6-hydroxydopamine and iron chelators, desferal and VK-28. *Ann NY Acad Sci* 2004;1012:306-25.
222. Powers KM, Smith-Weller T, Franklin GM, Longstreth WT Jr, Swanson PD, Checkoway H. Parkinson's disease risks associated with dietary iron, manganese, and other nutrient intakes. *Neurology* 2003;60(11):1761-6.
223. Kaur D, Andersen J. Does cellular iron dysregulation play a causative role in Parkinson's disease? *Ageing Res Rev* 2004;3(3):327-43.
224. Schipper HM. Brain iron deposition and the free radical-mitochondrial theory of ageing. *Ageing Res Rev* 2004;3(3):265-301.
225. Hegde ML, Shanmugavelu P, Vengamma B, Rao TS, Menon RB, Rao RV, Rao KS. Serum trace element levels and the complexity of inter-element relations in patients with Parkinson's disease. *J Trace Elem Med Biol* 2004;18(2):163-71.
226. Forte G, Bocca B, Senofonte O, Petrucci F, Brusa L, Stanzione P, Zannino S, Violante N, Alimonti A, Sancesario G. Trace and major elements in whole blood, serum, cerebrospinal fluid and urine of patients with Parkinson's disease. *J Neural Transm* 2004;111(8):1031-40.
227. Olanow CW. Manganese-induced parkinsonism and Parkinson's disease. *Ann NY Acad Sci* 2004;1012:209-23.
228. Dobson AW, Erikson KM, Aschner M. Manganese neurotoxicity. *Ann NY Acad Sci* 2004;1012:115-28.
229. Martin R, McFarland HF, McFarlin DE. Immunological aspects of demyelinating diseases. *Annu Rev Immunol* 1992;10:153-87.
230. Steinman L. Multiple sclerosis: a coordinated immunological attack against myelin in the central nervous system. *Cell* 1996;85(3):299-302.
231. Noseworthy JH. Progress in determining the causes and treatment of multiple sclerosis. *Nature* 1999;399:A40-7.
232. Gilgun-Sherki Y, Melamed E, Offen D. The role of oxidative stress in the pathogenesis of multiple sclerosis: the need for effective antioxidant therapy. *J Neurol* 2004;251(3):261-8.
233. van der Goes A, Brouwer J, Hoekstra K, Roos D, van den Berg TK, Dijkstra CD. Reactive oxygen species are required for the phagocytosis of myelin by macrophages. *J Neuroimmunol* 1998;92:67-75.
234. Barnes PJ, Karin M. Nuclear factor-kappaB: a pivotal transcription factor in chronic inflammatory diseases. *N Engl J Med* 1997;336:1066-71.
235. Winyard PG, Blake DR. Antioxidants, redox-regulated transcription factors, and inflammation. *Adv Pharmacol* 1997;38:403-21.
236. Syburra C, Passi S. Oxidative stress in patients with multiple sclerosis. *Ukr Biokhim Zh* 1999;71(3):112-5.
237. Skukla UK, Jensen GE, Clausen J. Erythrocyte glutathione peroxidase deficiency in multiple sclerosis. *Acta Neurol Scand* 1997;56(6):542-50.
238. Jensen GE, Clausen J. Glutathione peroxidase and reductase, glucose-6-phosphate dehydrogenase and catalase activity in multiple sclerosis. *J Neurol Sci* 1984;63(1):45-53.
239. Karg E, Klivenyi P, Nemeth I, Bencsik K, Pinter S, Vecsei L. Nonenzymatic antioxidants of blood in multiple sclerosis. *J Neurol* 1999;246:533-9.
240. Greco A, Minghetti L, Sette G, Fieschi C, Levi G. Cerebrospinal fluid isoprostane shows oxidative stress in patients with multiple sclerosis. *Neurology* 1999;53:1876-9.
241. Calabrese V, Raffaele R, Cosentino E, Rizza V. Changes in cerebrospinal fluid levels of malondialdehyde and glutathione reductase activity in multiple sclerosis. *Int J Clin Pharmacol Res* 1994;14:119-23.
242. Langemann H, Kabiersch A, Newcombe J. Measurement of low-molecular-weight antioxidants, uric acid, tyrosine and tryptophan in plaques and white matter from patients with multiple sclerosis. *Eur Neurol* 1992;32:248-52.

243. Lu F, Selak M, O'Connor J, Croul S, Lorenzana C, Butunoi C, Kalman B. Oxidative damage to mitochondrial DNA and activity of mitochondrial enzymes in chronic active lesions of multiple sclerosis. *J Neurol Sci* 2000;177:95-103.
244. Vladimirova O, Lu FM, Shawver L, Kalman B. The activation of protein kinase C induces higher production of reactive oxygen species by mononuclear cells in patients with multiple sclerosis than in controls. *Inflamm Res* 1999;48:412-6.
245. Matute C, Perez-Cerda F. Multiple sclerosis: novel perspectives on newly forming lesions. *Trends Neurosci* 2005;28(4):173-5.
246. Matute C, Alberdi E, Domercq M, Perez-Cerda F, Perez-Samartin A, Sanchez-Gomez MV. The link between excitotoxic oligodendroglial death and demyelinating diseases. *Trends Neurosci* 2001;24(4):224-30.
247. Noseworthy JH, Lucchinetti C, Rodriguez M, Weinshenker BG. Multiple sclerosis. *N Engl J Med* 2000;343:938-52.
248. Pitt D, Werner P, Raine CS. Glutamate excitotoxicity in a model of multiple sclerosis. *Nat Med* 2000;6:67-70.
249. Smith T, Groom A, Zhu B, Turski L. Autoimmune encephalomyelitis ameliorated by AMPA antagonists. *Nat Med* 2000;6:62-6.
250. Gilgun-Sherki Y, Panet H, Melamed E, Offen D. Riluzole suppresses experimental autoimmune encephalomyelitis: implications for the treatment of multiple sclerosis. *Brain Res* 2003;989:196-204.
251. Smith KJ, Kapoor R, Felts PA. Demyelination: the role of reactive oxygen and nitrogen species. *Brain Pathol* 1999;9:69-92.
252. Stover JF, Pleines UE, Morganti-Kossmann MC, Kossmann T, Lowitzsch K, Kempfski OS. Neurotransmitters in cerebrospinal fluid reflect pathological activity. *Eur J Clin Invest* 1997;27: 1038-43.
253. Barkhatova VP, Zavalishin IA, Askarova LSh, Shavratskii VKh, Demina EG. Changes in neurotransmitters in multiple sclerosis. *Neurosci Behav Physiol* 1998;28:341-4.
254. Werner P, Pitt D, Raine CS. Multiple sclerosis: altered glutamate homeostasis in lesions correlates with oligodendrocyte and axonal damage. *Ann Neurol* 2001;50:169-80.
255. Cross AH, Misko TP, Lin RF, Hickey WF, Trotter JL, Tilton RG. Aminoguanidine, an inhibitor of inducible nitric oxide synthase, ameliorates experimental autoimmune encephalomyelitis in SJL mice. *J Clin Invest* 1994;93:2684-90.
256. Zhao W, Tilton RG, Corbett JA, McDaniel ML, Misko TP, Williamson JR, Cross AH, Hickey WF. Experimental allergic encephalomyelitis in the rat is inhibited by aminoguanidine, an inhibitor of nitric oxide synthase. *J Neuroimmunol* 1996;64: 123-33.
257. Brenner T, Brocke S, Szafer F, Sobel RA, Parkinson JF, Perez DH, Steinman L. Inhibition of nitric oxide synthase for treatment of experimental autoimmune encephalomyelitis. *J Immunol* 1997;158:2940-6.
258. Ding M, Zhang M, Wong JL, Rogers NE, Ignarro LJ, Voskuhl RR. Antisense knockdown of inducible nitric oxide synthase inhibits induction of experimental autoimmune encephalomyelitis in SJL/J mice. *J Immunol* 1998;160:2560-4.
259. Hooper DC, Bagasra O, Marini JC, Zborek A, Ohnishi ST, Kean R, Champion JM, Sarker AB, Bobroski L, Farber JL, Akaike T, Maeda H, Koprowski H. Prevention of experimental allergic encephalomyelitis by targeting nitric oxide and peroxynitrite: implications for the treatment of multiple sclerosis. *Proc Natl Acad Sci U S A* 1997;94:2528-33.
260. Hooper DC, Spitsin S, Kean RB, Champion JM, Dickson GM, Chaudhry I, Koprowski H. Uric acid, a natural scavenger of peroxynitrite, in experimental allergic encephalomyelitis and multiple sclerosis. *Proc Natl Acad Sci U S A* 1998;95:675-80.
261. Lehmann D, Karussis D, Misrachi-Koll R, Shezen E, Ovadia H, Abramsky O. Oral administration of the oxidant-scavenger N-acetyl-L-cysteine inhibits acute experimental autoimmune encephalomyelitis. *J Neuroimmunol* 1994;50:35-42.
262. Ruuls SR, Bauer J, Sontrop K, Huitinga I, 't Hart BA, Dijkstra CD. Reactive oxygen species are involved in the pathogenesis of experimental allergic encephalomyelitis in Lewis rats. *J Neuroimmunol* 1995;56:207-17.
263. Malfroy B, Doctrow SR, Orr PL, Tocco G, Fedoseyeva EV, Benichou G. Prevention and suppression of autoimmune encephalomyelitis by EUK-8, a synthetic catalytic scavenger of oxygen-reactive metabolites. *Cell Immunol* 1997;177(1):62-8.
264. Marracci GH, Jones RE, McKeon GP, Bourdette DN. Alpha lipoic acid inhibits T cell migration into the spinal cord and suppresses and treats experimental autoimmune encephalomyelitis. *J Neuroimmunol* 2002;131:104-14.
265. Morini M, Roccatagliata L, Dell'Eva R, Pedemonte E, Furlan R, Minghelli S, Giunti D, Pfeffer U, Marchese M, Noonan D, Mancardi G, Albin A, Uccelli A. Alpha-lipoic acid is effective in prevention and treatment of experimental autoimmune encephalomyelitis. *J Neuroimmunol* 2004;148:146-53.
266. Liu Y, Zhu B, Wang X, Luo L, Li P, Paty DW, Cynader MS. Bilirubin as a potent antioxidant suppresses experimental autoimmune encephalomyelitis: implications for the role of oxidative stress in the development of multiple sclerosis. *J Neuroimmunol* 2003;139:27-35.
267. Jensen GE, Clausen J. Glutathione peroxidase activity, associated enzymes and substrates in blood cells from patients with multiple sclerosis-effects of antioxidant supplementation. *Acta Pharmacol Toxicol* 1986;59:450-3.
268. Swank RL, Dugan BB. Effect of low saturated fat diet in early and late cases of multiple sclerosis. *Lancet* 1990;336(8706):37-9.
269. Mai J, Sorensen PS, Hansen JC. High dose antioxidant supplementation to MS patients. Effects on glutathione peroxidase, clinical safety, and absorption of selenium. *Biol Trace Elem Res* 1990;24:109-17.
270. Spitsin S, Hooper DC, Leist T, Streletz LJ, Mikheeva T, Koprowski H. Inactivation of peroxynitrite in multiple sclerosis patients after oral administration of inosine may suggest possible approaches to therapy of the disease. *Mult Scler* 2001;7(5):313-9.
271. LeVine SM. Iron deposits in multiple sclerosis and Alzheimer's disease brains. *Brain Res* 1997;760(1-2):298-303.
272. LeVine SM, Lynch SG, Ou CN, Wulser MJ, Tam E, Boo N. Ferritin, transferrin and iron concentrations in the cerebrospinal fluid of multiple sclerosis patients. *Brain Res* 1999;821(2):511-5.
273. Hulet SW, Powers S, Connor JR. Distribution of transferrin and ferritin binding in normal and multiple sclerotic human brains. *J Neurol Sci* 1999;165(1):48-55.
274. Melo TM, Larsen C, White LR, Aasly J, Sjobakk TE, Flaten TP, Sonnewald U, Syversen T. Manganese, copper, and zinc in cerebrospinal fluid from patients with multiple sclerosis. *Biol Trace Elem Res* 2003;93(1-3):1-8.
275. Levine SM, Chakrabarty A. The role of iron in the pathogenesis of experimental allergic encephalomyelitis and multiple sclerosis. *Ann NY Acad Sci* 2004;1012:252-66.
276. Behl C. Oxidative stress in Alzheimer's disease: implications for prevention and therapy. *Subcell Biochem* 2005;38:65-78.