

Introduction to NGS data formats, quality check and analytical tools

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Bioinformatics training,
July 2019

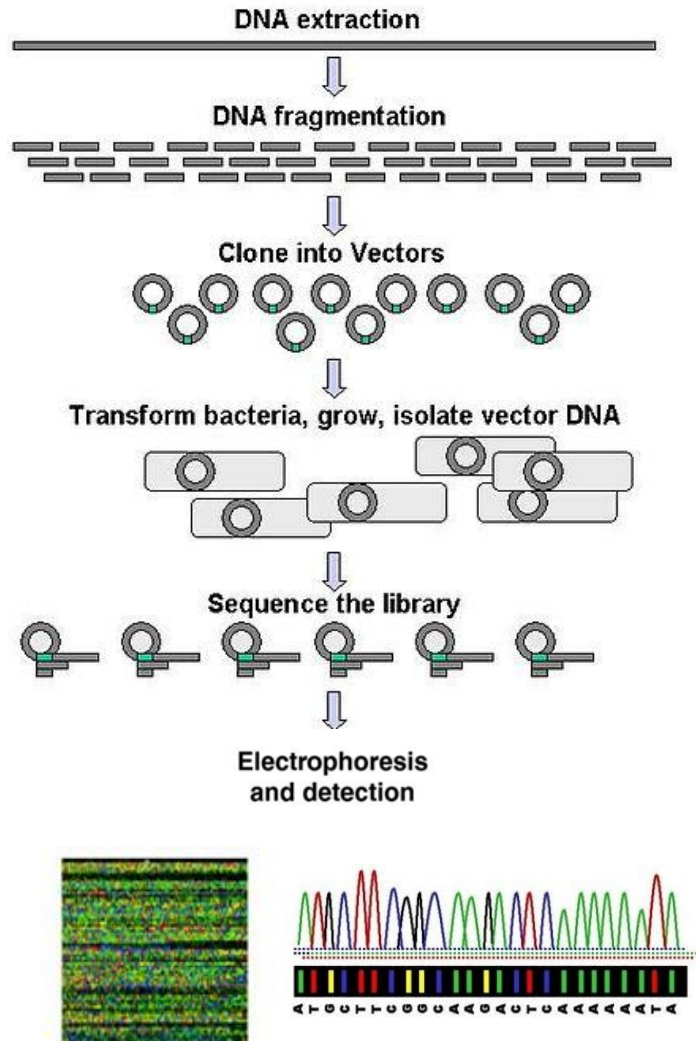


Istituto Superiore di Sanità, Dep. of Food Safety, Nutrition and Veterinary Public Health
European Union and National Reference Laboratory for *E. coli*, Rome, Italy

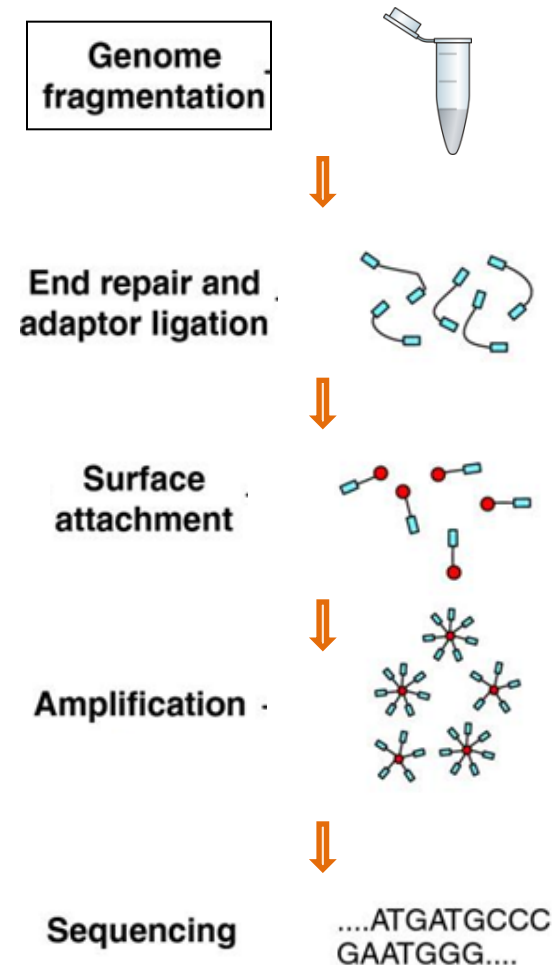


Conventional sequencing vs NGS

Conventional

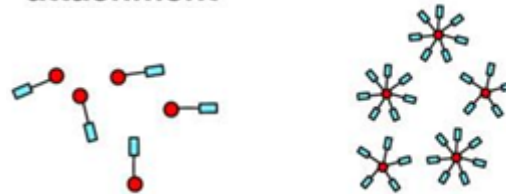


NGS Pipeline



Next generation sequencing

Surface attachment → Amplification



...

...



illumina MiSeq

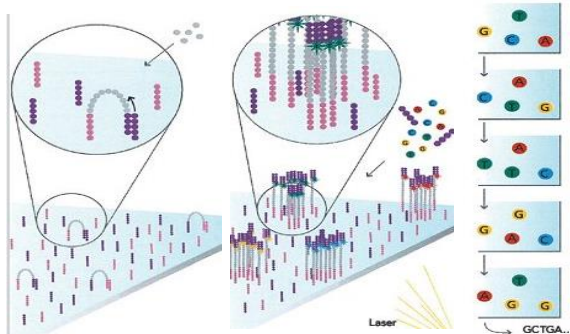
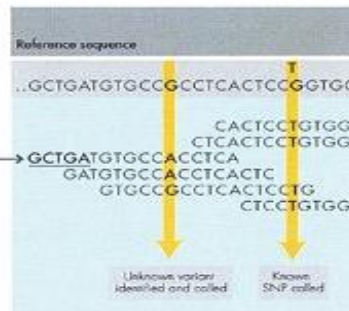


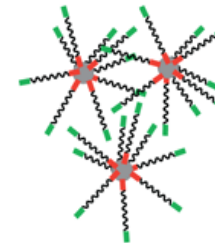
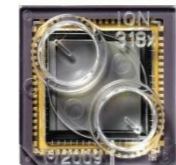
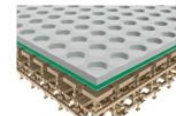
Image capture
Fluorescence detection



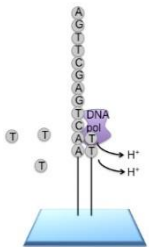
200bp-400bp short reads



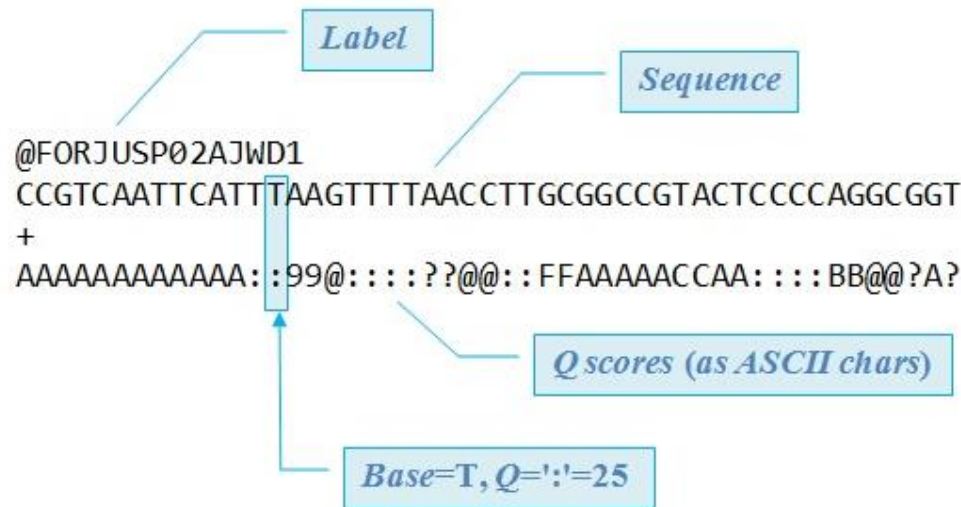
Ion Semiconductor Sequencing Chip



pH variation when incorporating nucleotides in the growing strand



.fastq files



Each .fastq file covering a 5 Mb genome at 50X weights about **500 MB**

Phred quality scores are logarithmically linked to error probabilities

Phred Quality Score	Probability of incorrect base call	Base call accuracy
10	1 in 10	90%
20	1 in 100	99%
30	1 in 1000	99.9%
40	1 in 10,000	99.99%
50	1 in 100,000	99.999%
60	1 in 1,000,000	99.9999%

Phred quality score

$$Q = -10 \log_{10} P$$

from 0 to 93 using ASCII characters 33 to 126

.fastq files

@

```
@X1L6C:01561:00672
AAATATCACCAATAAAAAAGCCTTAGTAAGTATTTTTCAGCTTTTCATTCTGACTGCAACGGGCAATATGTCTCTGTGTG
GATTAAGAGAGTGTCTGATAGCAGCTTCTGAAGTGGTTACCTGCCGTGAGTAAATTTTATTGACTTAGGTCAC
TAAATACCTTAACCAATATAGGCATAGCGCACAGACAGATAAAAAATTACAGAGTACACAACATCCATGAAACGCATTAGCA
CCACCATTACCACCACCATCACCATTACCACAGGTAACGGTGC GGCTGACGCGTACAGGAAACACAGAAAAAAGCCCGCA
CCTGACAGTGGGGCTTTTTCGACCAAGGTAACGAGGTAACAACCATGCG
```

@

```
+
CC:9::FBC<CD7:88888(>><C<CCCC<CCBBAAB/A@A8888,,<@;AABBB=?;B98992:B<
CGBBCCGDC??BCC;BB<ADEED*CCCCAACCCBCABBDDB>B?A;999;@8=>199A7>9::CBCH:B:>>>)999)
77037;<7==5=@@BCC:C@BBB9B<E<D9>>><6ADCBBAABBB@@@DDCBA@==+=.//?B<?AEB::6;DCD>
C;;;-:9:BC<BBCCC9?><AA;AG<CB>GD@B;;;A<AE;AA<B?>@9@C<BB<?>?BB;BBBAAAA::BAB099/9>
@=====(<<?)99997>>CCEBA>>=>2373333&3:99-33(3--717--43606704/47761
```

@

```
@X1L6C:01104:03031
AGAAGCTGCTATCAGACACTTTTTTTAATCCACACAGAGACATATTGCCCGTTGACGTGAGAATGAAAAGCTGAAAAATA
CTTACTAAGGCGTTTTTTTATTGGTGATTTTTTTCAATATCATGCAGCAAACGGTGCAACATTGCCGTGTCTCGTTGCTC
TAAAAGCCCCAGGCG
+
```

@

```
@AC=BCCC??B?@CBB@??>>>?8??>DAABEBCBABCAAA:@@+9:8>;<;/.
98283988*44449;;9/88:29:>>5;7833333&399298:6/./DCDDCC';>:ACBDAABB??9::+9<
1444@:77-3<03368:8755888;;9833)3777'--'--
@X1L6C:03659:02717
```

@

```
GCTTCTGAAGTGGTTACCTGCCGTGAGTAAATTTTATTGACTTAGGTCACTAACTTTAACCAATATAGGCATA
GCGCACAGACAGATAAAAAATTACAGAGTACACAACATCCATGAAACGCATTAGCACACCATTACCACCACCATCACCATTAC
CCACAGGTAACGGTGC GGCTGACGCGTACAGGAAACACAGAAAAAAGACCCGCCACTGACCAAGTGC
```

```
+
??9?BB@<CAA;A8@?:?@5::BCCCEC;C=CCC8CEJ8DE;AACF>CC?DDCCCB:B@??9?;B=B=CAA@?;?BCG
CCCCCBABBBBCCDDAA2:4;@???CAB@AAA9@AB?C;;;C;CDCCC>ECCAA<AC<CB>DC<AB=CD=C9::A4::>
CC;@@@A?CI@DDAFKDDD:A@CBCDC:::99199+8;4746@CA?)<444/3:4934333-3888//
@X1L6C:02011:02071
```

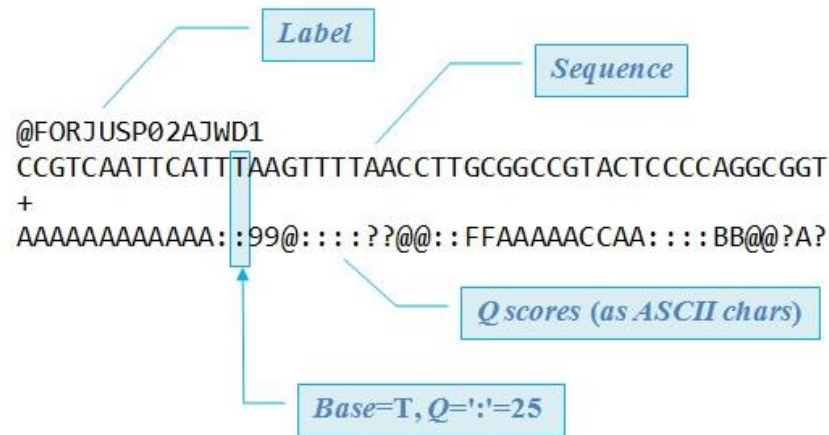
```
TTAAATTTTATTGACTTAGGTCACTAACTTTAACCAATATAGGCATAGCGCACAGACAGATAAAAAATTACAGAGTACA
CAACATCCATGAAACGCATTAGCACACCATTACCACCACCATCACCATTACCACAGGTAACGGTGC GGCTGACGCGTACAG
GAAACACAGAAAAAAGCCCGCACCTGACAGTGC GGCTTTTTTTTCGACCAAGTAACG
+
=0>>>19;;,,;7=CCDADC;?:::,,5;;=4>273:<@BBCF=CDH;@;MMFEED@?>>>:::~5/55<
;;@:::BC=BCBB<B@@@D<@B;3:::9@<BB=BD=AC;@B:??3::CAC=CD;;=BBAB>CC;AA;BAAA9AD@>>
>>955>4?94999855555&4<>2;;661499888...88/56666666$;6/.5:8(..+'++
@X1L6C:01333:03005
GCAATGCCAGGCGAGGCATGTACAGCTACGTACGTCTGAGCATCGATCGATGTACAGCTACGTACGTCTGAGCATCGATCGA
TGTACAGCTACGTACGTCTGAGCATCGATCGATGTACAGCTACG
+
```

```
555/55/(/(/(/(/8:9:<=><?@:98A??676<;;@:555555554444;=4443333;383338<68>>
68=333111831111111113933644588?==<76992---2+++0/
```

...and so on

Quality check

Output of NGS
sequencers



Input for
quality check

.fastq file

Sequencing errors would impact every following application

Unreliability of following results (and difficulty to detect the existence of problems!)

Parameters to control

- Phred score
- GC content distribution over all sequences
- Distribution of undetermined bases (N)
- Distribution of nucleotides
- ★ • Length of the reads
- ★ • Coverage

Adoption of corrective actions is possible to minimize some of these problems

Coverage (depth)

Reads mapped on a reference genome



Ref seq

COVERAGE

Mapped reads

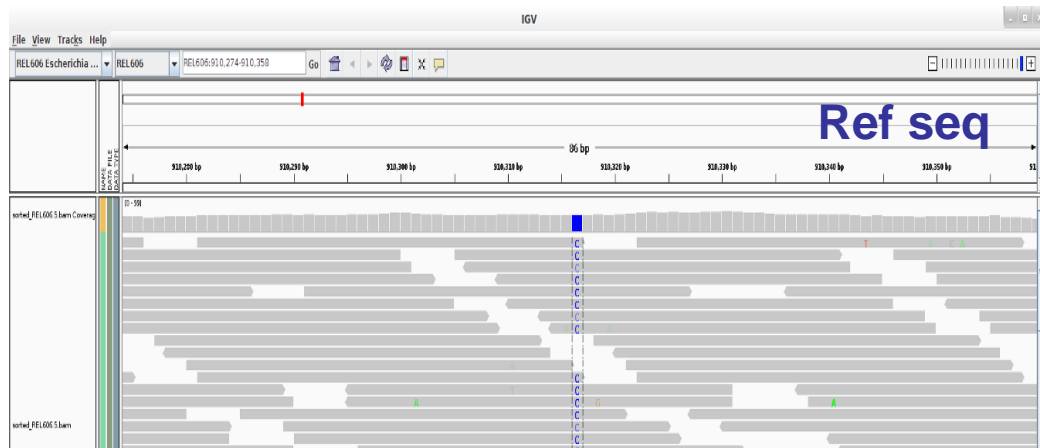
Coverage assessment:

Total length sequenced in Mb / expected genome size (5 Mb)

Count of reads mapping on housekeeping genes (e.g. MLST)

Alignment (mapping)

Alignment of the sequencing reads on a reference sequence or on a database of reference sequences



Possibility to directly inspect the **presence/absence of a target sequence** and the presence of **SNPs at interesting positions** by opening the bam file with a graphic viewer (e.g. IGV)

QNAME	FLAG	RNAME	POS	MAPQ	CIGAR
ME2UT:01383:01267	0	gad:3:EF547388	1285	0	113M18I4M
ME2UT:02555:01592	16	gad:4:CP001925	1123	0	27M1I248M39I4M
ME2UT:02231:01820	0	gad:5:CP001846	87	1	138M
ME2UT:01605:00255	16	gad:5:CP001846	399	1	51M
ME2UT:01345:02031	16	gad:5:CP001846	685	1	176M
ME2UT:03330:02136	16	gad:5:CP001846	1050	1	6M1I38M
ME2UT:01475:02165	0	gad:6:BA000007	1	0	3M31I47M1D130M
ME2UT:01488:00709	16	gad:6:BA000007	1	0	4M32I55M1I149M
ME2UT:01943:01152	16	gad:6:BA000007	13	1	196M1I50M1I10M

Possibility to convert the output in a sam file (tabular) to extract interesting info and sequences

Assembly

Short sequencing reads

.fastq file

```
@HWI-ST700693:238:B0224ACXX:1:1101:1218:1982
NACACTTGCTTTGGTGACAGCGGGGCATCCTCAAGC
+
#1=DDDDHHAFF?GEFGIIIIIIIIIIIIIIIFI
@HWI-ST700693:238:B0224ACXX:1:1101:1161:1986
NGATTTTGACCTCTCCAGTTTCCTCTTAACACTTTG
+
#1=BDFFFHGHGJJJJIJHIJJJJJJJJJJJJJJ
@HWI-ST700693:238:B0224ACXX:1:1101:1193:1989
NTATCCAGCCTGCGGTGCTACTTGGTGGAAGAGGAT
+
#1=DDFFFHGHGJJJFJGHJJJJJJIEGECHDFHCC?
@HWI-ST700693:238:B0224ACXX:1:1101:1440:1981
NTCAAGAATCCAAGTGGGGCCAGCATAATGTACGCT
+
#1=DDFFFHGHDFDAEGIIIFGIICGGHGBFGEFDHI
@HWI-ST700693:238:B0224ACXX:1:1101:1367:1983
NATTAGAACAGATCGCTACTTCGCCCGAAGATACAT
+
#4BDFFFHHHHHJGIIJJJJJJJJJJJJJJJJJJ
@HWI-ST700693:238:B0224ACXX:1:1101:1395:1988
NTGGAACGTTTTTAAACGCGGAGACAGCGTGAGAT
+
#1=DDFFFHCFHJJJJJJJJJJJJJJJJJJJJJJ
@HWI-ST700693:238:B0224ACXX:1:1101:1632:1994
NCTTTGCTGTATTGACCGTTTGTAGATTTGAATCTT
+
#4=DDFFFHBBHHHIGIJJFHIJFGGGIGIHIJJII
@HWI-ST700693:238:B0224ACXX:1:1101:1632:1989
NTCTATGAATGTTCAAGCGGTAGCTGAGGAGAGTCC
+
```



Partially assembled genome (contigs)

.fasta file

```
>NODE 1 length 449 cov 4.835189
ATCTTTTCGCGCCTTCCAGCTCCAGCCATTGCGAACCGTTGCGCAGAAAACGGGCGTAATC
GGGTAAAGACATAGCGCGGTTTTGTACGGCGCATGACCTTCAAACATATCGCAGATTACACC
TTCATCCAGCGCGCGCGGGGCTTCCGAGGAAGCTGTGGGTAAAGCAGATTGTTTTCTGC
TTCCAGTGCCAGAAAATGGCGCTTCTGCTCCGGGCTAAGCACTGGGCTGGTGACAATTTG
CTGGCAACGTTGTTGCAGTGCATTTTCATGAGAAGTGGGCATCTTCTTTCTTTTATGC
CGAAGGTGATGCGCCATTGTAAAGAGTTTCGTGATGTTCACTTTGATCCTGATGCGTTTG
CCACCACTGACGCATTCAATTTGAAAGTGAATTTTGAACCAAGATCGCATTACAGTGATG
CAAACCTGTAAAGTAGATTTCTTAATTGTGATGTGATCGAAGTGTGTTGCGG
>NODE 2 length 309 cov 4.686084
ACTGGTCAGTGCGGGTATCTTGACAATGGCCGATTGGACGTCTGGCGGATAAGTTTGG
TCGACTGCTGGTGTTCGCTGTTCAAGTCTTTGTGTCATCTCGGCAGTATCGCGATGCT
TAGCCAGGCGCGCATGGCCCCAGCGTTATTCATCCTCGGTGCGCTTGCTTACGCTATA
TCCGGTGGCGATGGCATGGGCTTGCAGAAAAGTTGAACATCATCAACTGGTGGCGATGAA
CCAGGCCTTACTGTTGAGCTATACTGTGGGAAGTCTGCTTGGCCCGTCATTTACCGCTAT
GCTAATGCAGAAATTTCTCCGATAATTTATTGTT
>NODE 3 length 101 cov 3.346535
AGCGCATGAGCGCGCAGCGCGCGCTTACGTGGTGCATCAGCATGATGTTGGCCGGAGAG
TACAGAGACTCCCCTTCATCCATGATGCCCTCTTCCACAGCAGTTCTTCAATCATCACC
AGACC
>NODE 4 length 311 cov 3.610933
CATCAACGCTAAAAGCCAGATGACGCAGACCGCAAGCTTCCGGTGGCTGGGTGTTCCG
GCGGGAACGGAAATGAGAAAAGCTCAATCACATATTGCCCATTAAGCGCCAAATCCCCTT
TCCATGAGTCGCGCGCTTCCGATAGACTTCGCTTTCAGCGGTGAAACCAAGAATATCGC
AGTAGAAAAGCTTTGCTCACCGCATAATCCGTGCAATATCGCAATATGGTGAACTGTT
TTAAACCCAGCATAACTGCTCTCTTTATTTGTTAACAGCAGTTACTCGCCCGGAAGCCG
TCTGGCAAGTTATCCCGCATTTTATAGACTCGTA
>NODE 5 length 186 cov 4.973118
CGAAGATATAAGAAAGCGAACCAGAAAGATGCCGGAGAACTTCATCAATTCACCTG
CATTGAGCAGATTTTGCAGGTTCTCAATTAACCGGTAAATCCAGCCCCAACGTTGGTGTAT
AGAGGAATTTACGCCCGGATTTTCCGCCGATTAACGCAACTGATGGTAGTAATCCATCG
ACGAGGTGTTGGCCTTTTGTTCGGCTGA
```

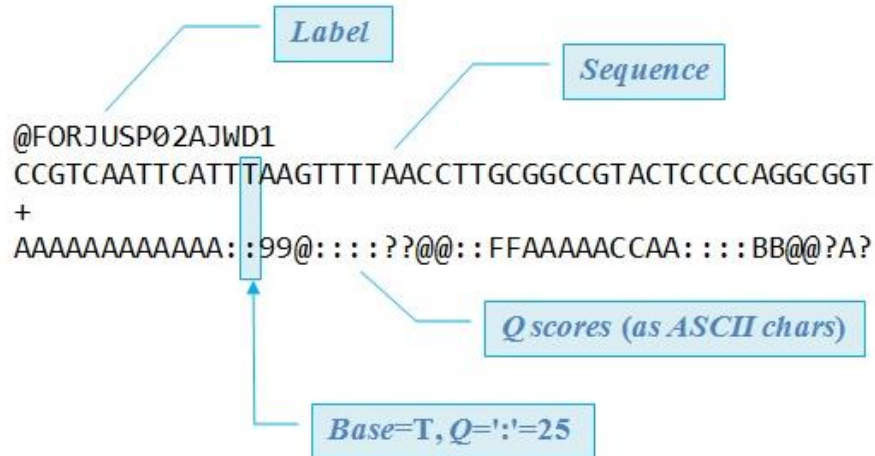
FastqSize \approx GenomeSize x Coverage x 2

At least 0.5 GB per genome

FastaSize for *E. coli* contigs

~5.5 MB

What should be trimmed out?



Phred quality scores are logarithmically linked to error probabilities

Phred Quality Score	Probability of incorrect base call	Base call accuracy
10	1 in 10	90%
20	1 in 100	99%
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40	1 in 10,000	99.99%
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60	1 in 1,000,000	99.9999%

Low quality positions

Adaptors and barcodes

Very short sequencing reads

What should be trimmed out?

FASTQ positional and quality trimming (Galaxy Version 0.0.1)

Is this library mate-paired?

Single-end

FASTQ file



No fastqsanger dataset available.

FASTQ format with Sanger-scaled quality values (Galaxy fastqsanger datatype)

Maximum length trimming

-1

Trim reads longer then this value (useful for Ion Torrent); -1 for no trimming

Left-side trimming

0

Number of bases to trim from 5' (left) end

Right-side trimming

0

Number of bases to trim from 3' (right) end

Minimum Phred quality score for right-side trimming

0

Starting from 3' (right) end, bases with quality less than this value will be trimmed

Average Phred quality score for right-side trimming

0

Starting from 3' (right) end, bases will be trimmed one-by-one until the average read quality reaches this value

Minimum length filtering

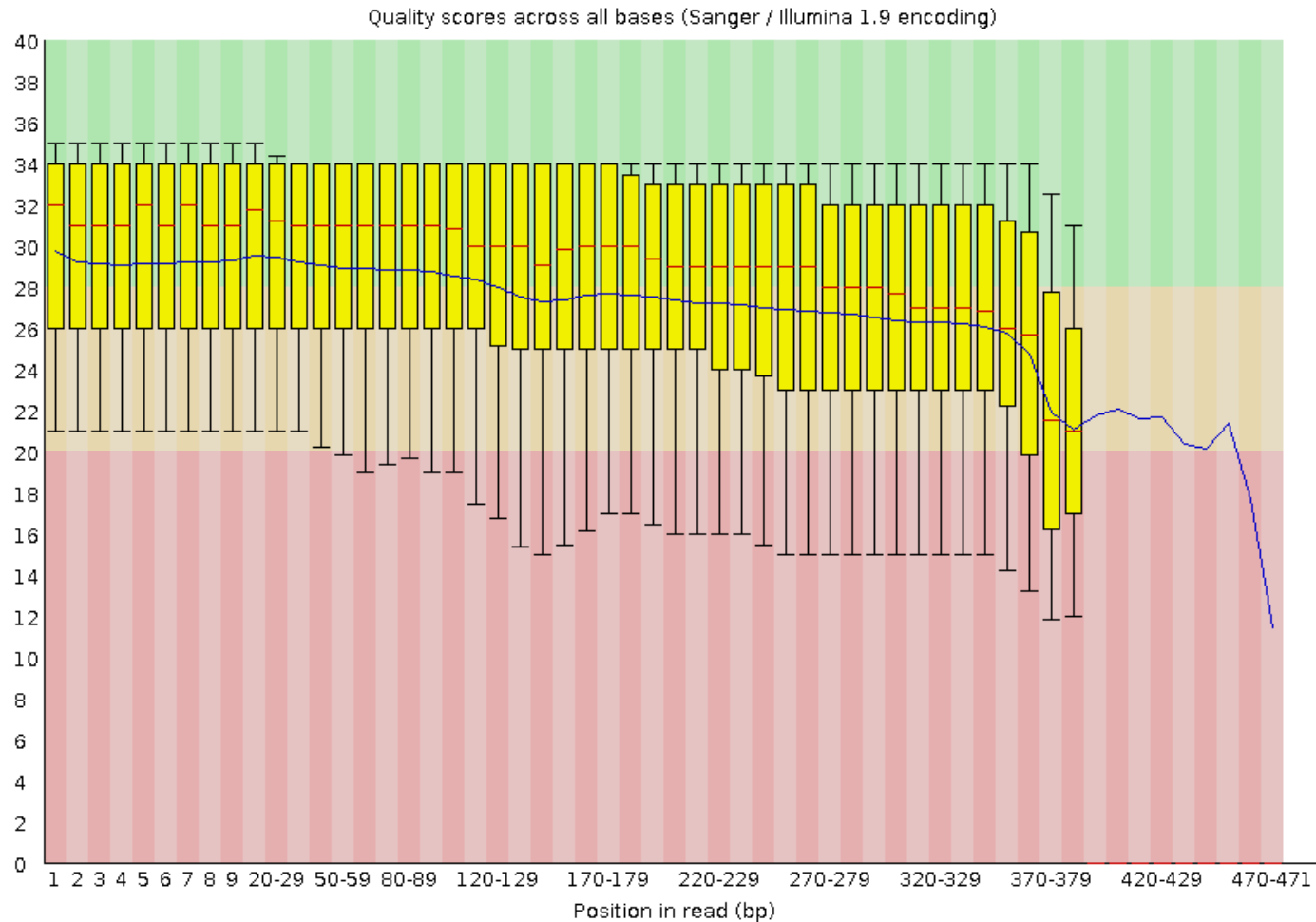
-1

Reads shorter than given length will be discarded; -1 for no filtering

✓ Execute

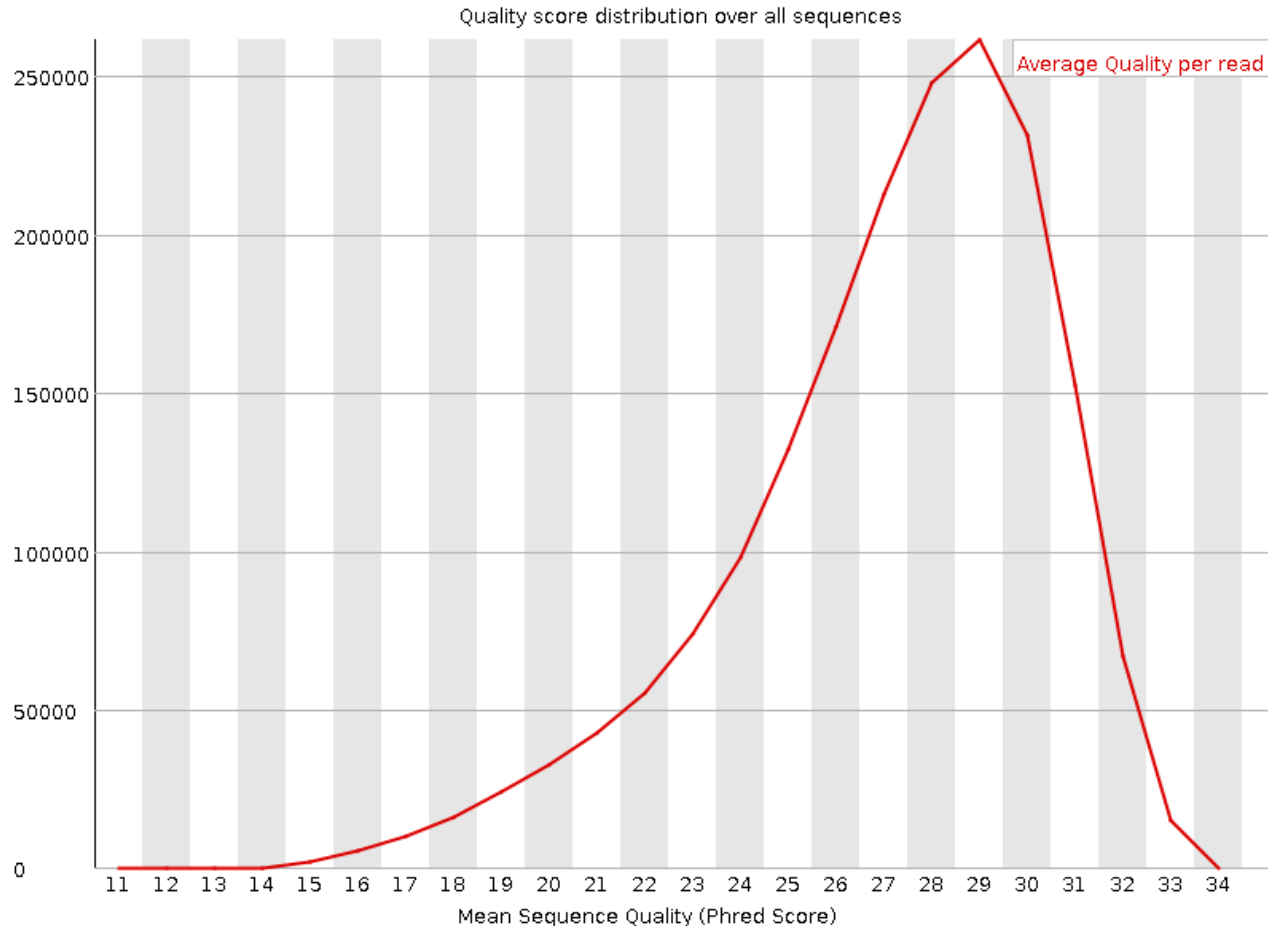
FastQC – quality check of aw data

❌ Per base sequence quality



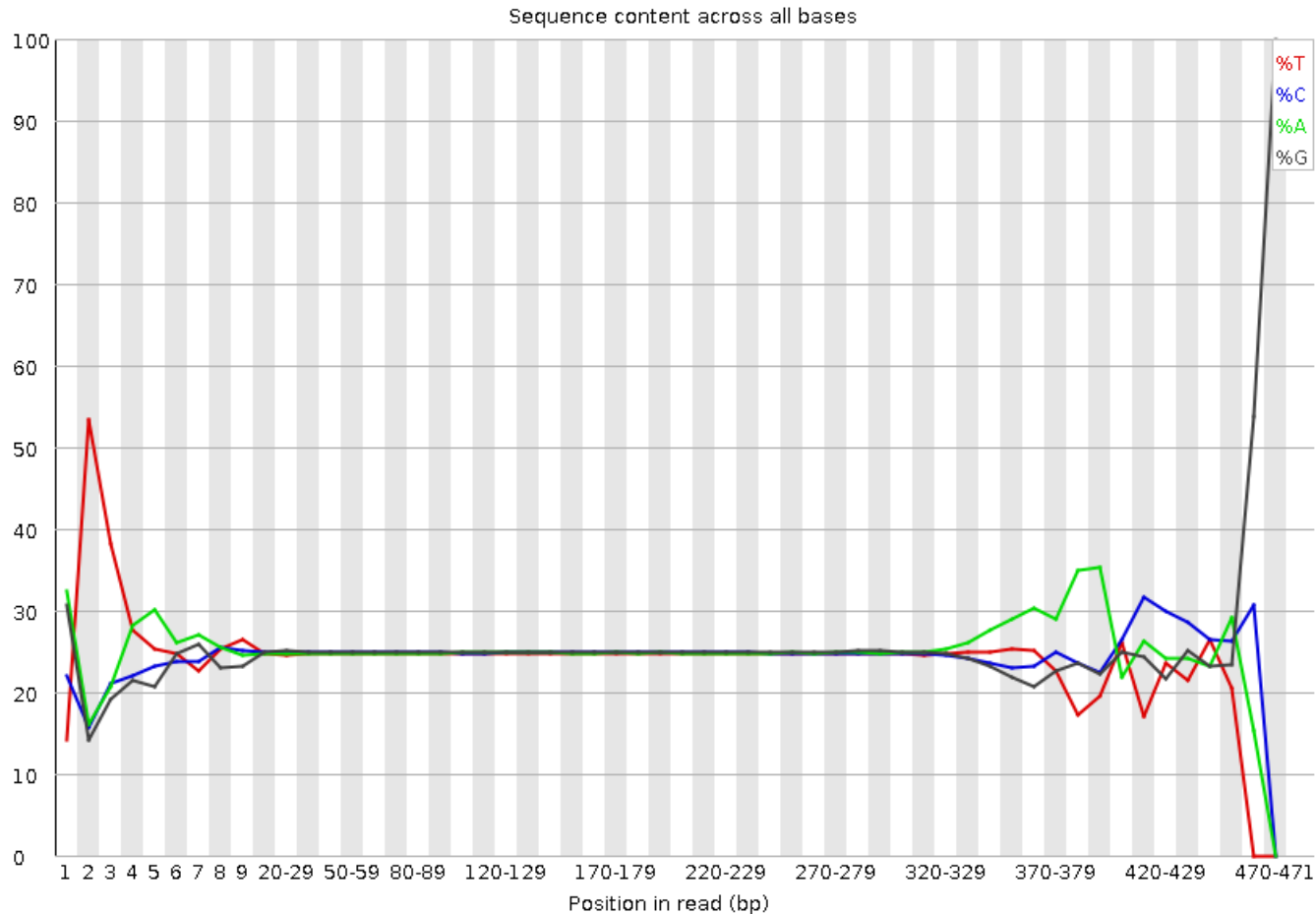
FastQC – quality check of aw data

✓ Per sequence quality scores



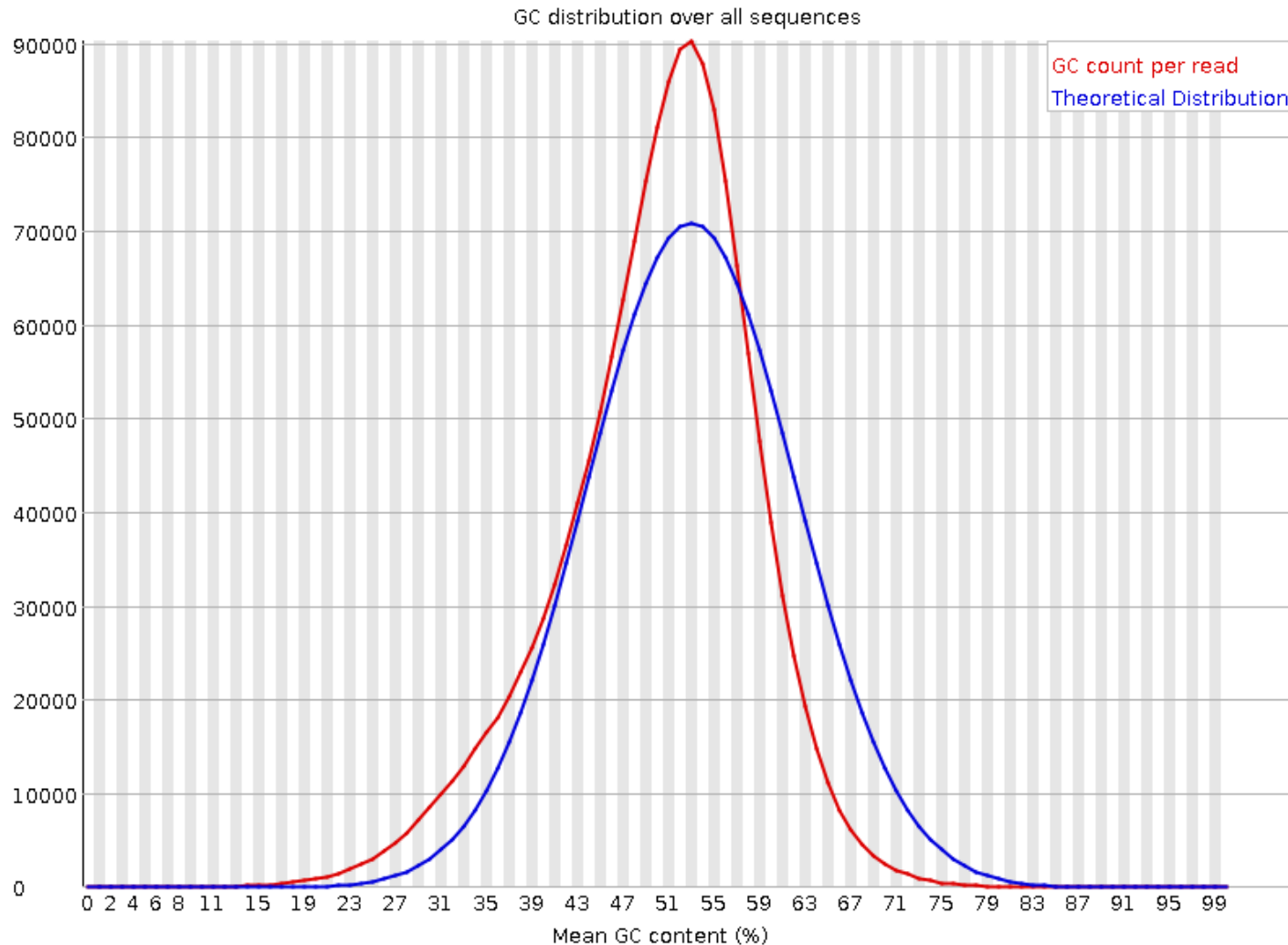
FastQC – quality check of raw data

❌ Per base sequence content



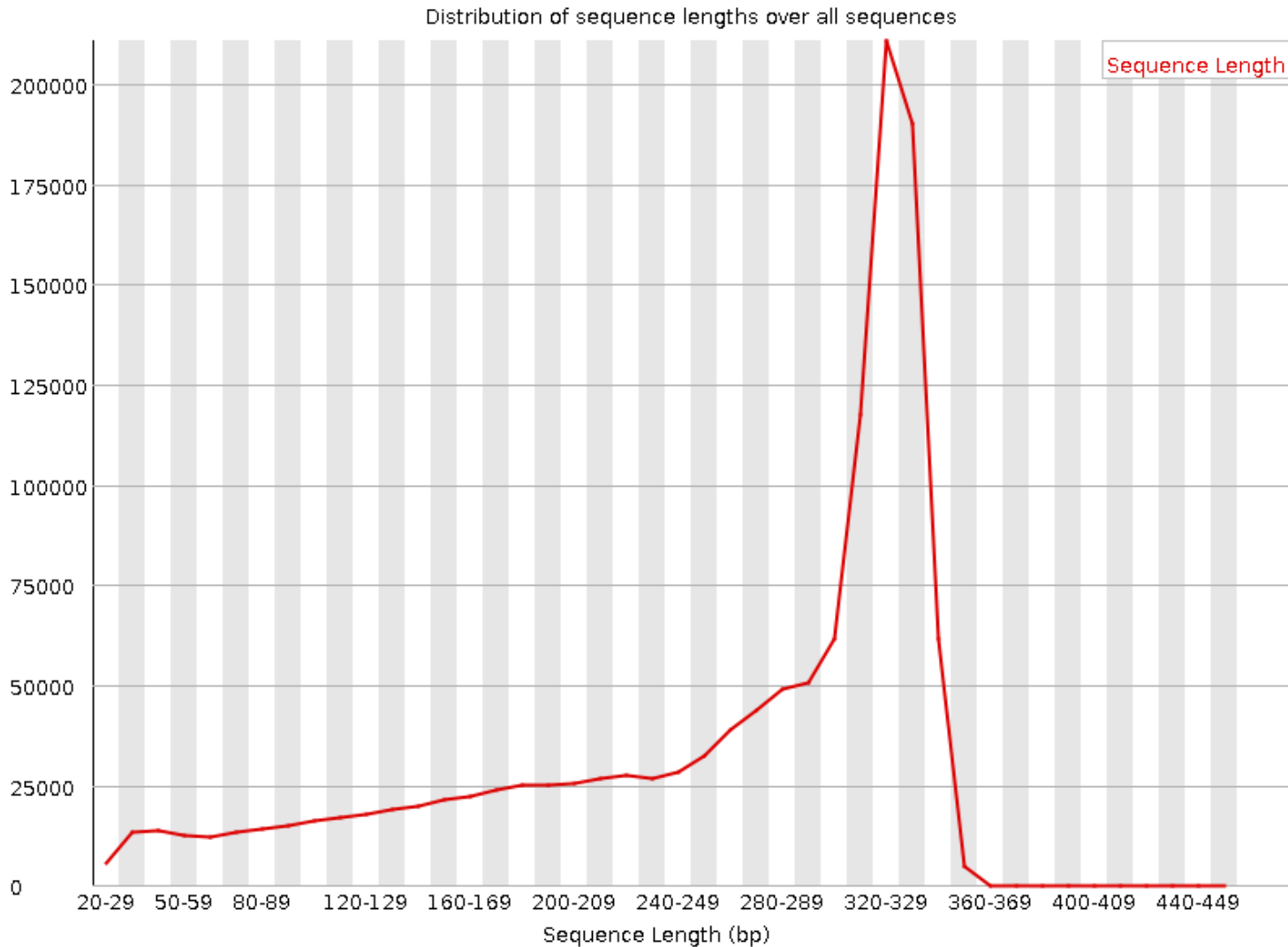
FastQC – quality check of aw data

Per sequence GC content



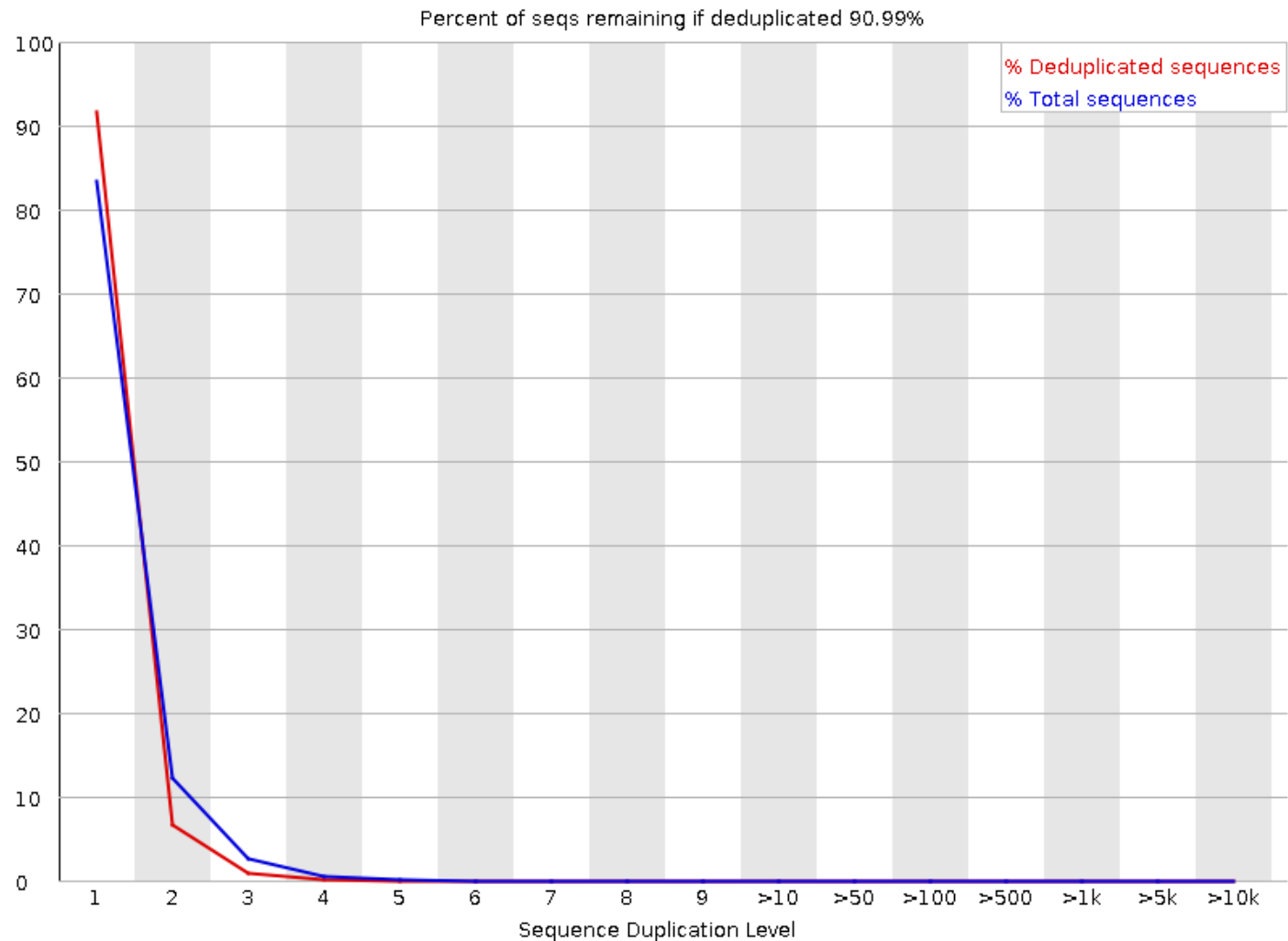
FastQC – quality check of aw data

Sequence Length Distribution



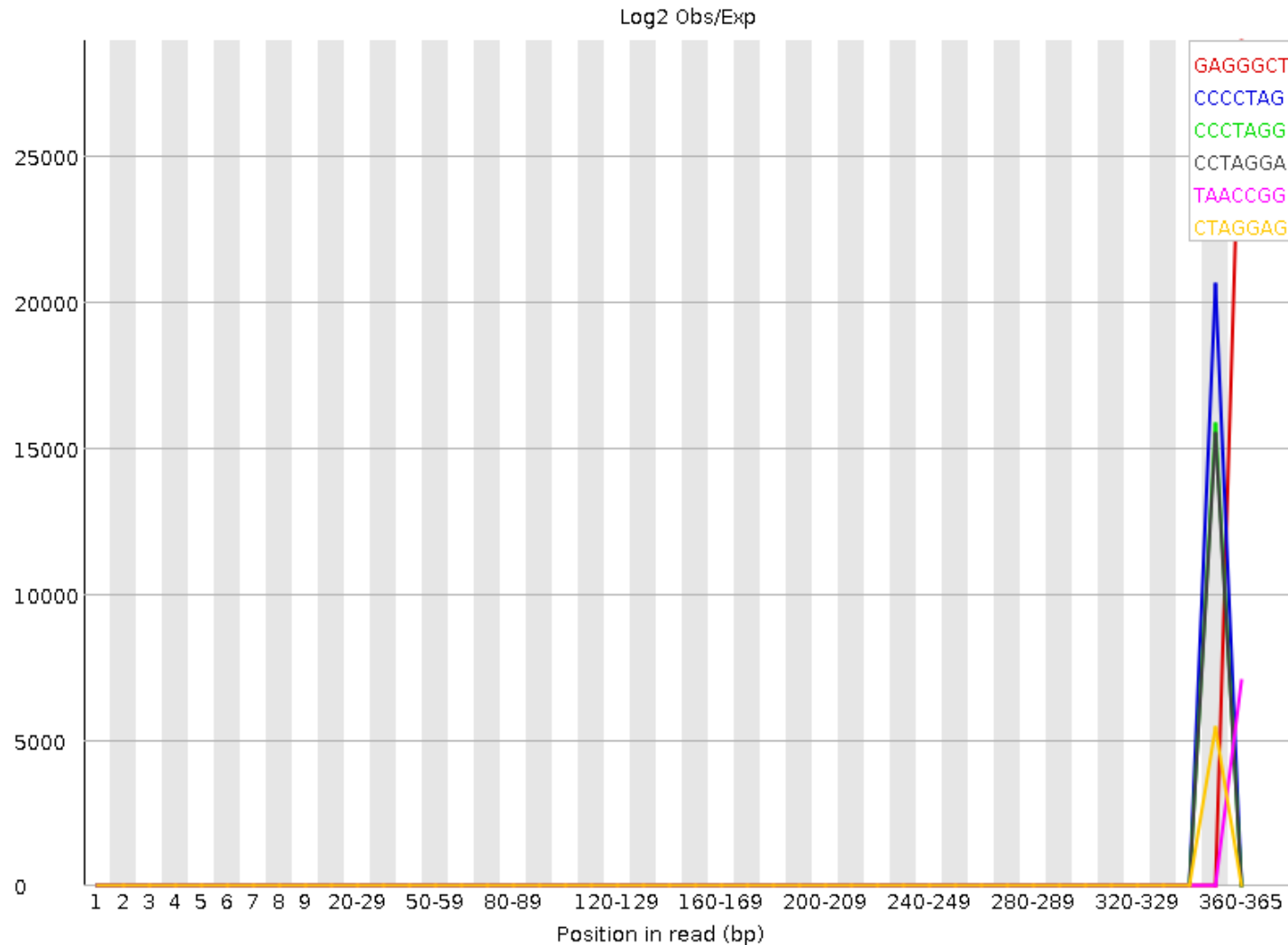
FastQC – quality check of aw data

✔ Sequence Duplication Levels



FastQC – quality check of raw data

Kmer Content



What should be trimmed out?

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Number of bases to trim from 3' (right) end

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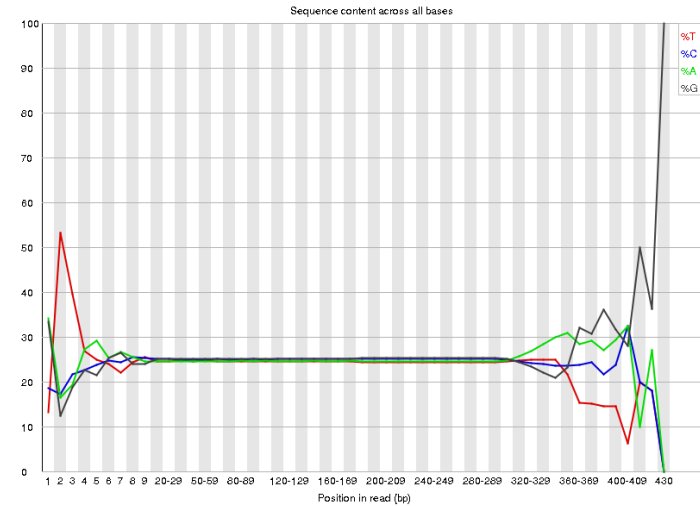
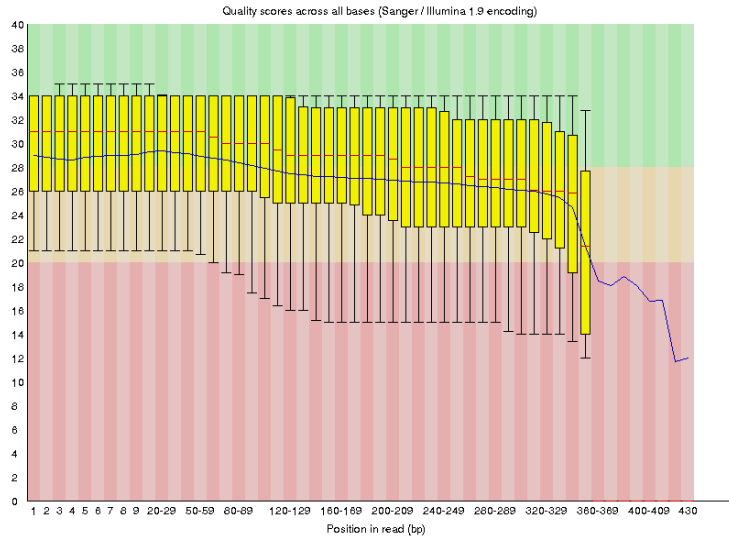
Minimum length filtering

-1

Reads shorter than given length will be discarded; -1 for no filtering

✓ Execute

Before trimming



After trimming

