

# Assessment of a real-time PCR for the identification and characterisation of Verocytotoxigenic *E. coli* (VTEC)

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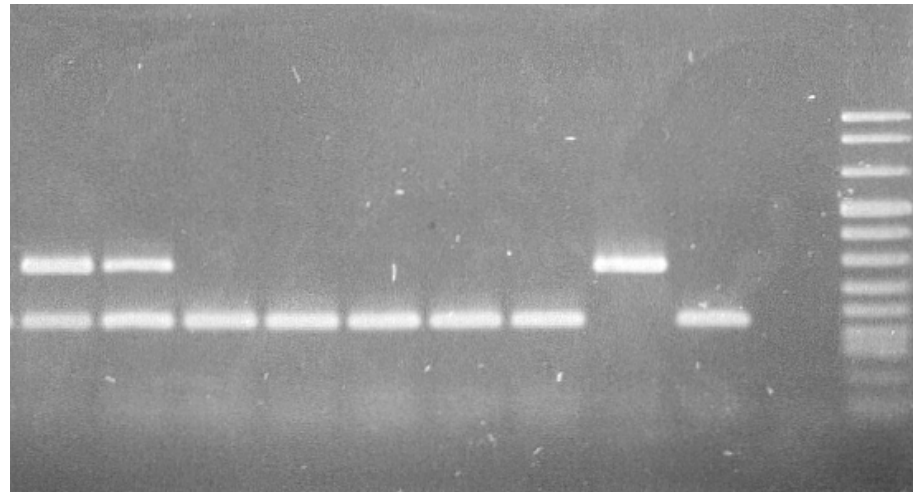


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# Previous block-based PCR for *vtx1* and *vtx2* genes

*vtx1*  
282bp

*vtx2*  
164bp



*vtx1* ↑  
*vtx2* ↑  
neg ↑  
controls

Detection of the *eae* (conserved) was by a separate block-based PCR

## Real-time PCRs for the identification of VTEC: guidelines from the Community Reference Laboratory (CRL) ([http://www.iss.it/binary/vtec/cont/VTEC\\_RealTime.pdf](http://www.iss.it/binary/vtec/cont/VTEC_RealTime.pdf))

- Vero cytotoxin genes (*vtx1* & *vtx2*)
  - Conserved region of intimin (*eae*)
  - *rfbE* encoding the O antigen of *E. coli* O157.
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- validation study using VTEC cultures
  - use of the assay in rapid identification of UK cases during the outbreak of VTEC O104 outbreak in Germany

# Organisation of the real time assays

- Two duplex PCR on 5-plex Rotorgene-Q (Qiagen)
- Duplex 1 = *vtx1* + *eae*; duplex 2 = *vtx2* + *rfbO157*
- Amplification parameters: 95°C for 5m, followed by 95°C for 15s and 60°C for 60s. The cycle threshold was 0.025 for *vtx1* and 0.05 for the others

	Duplex#1	Conc	Vol ul	Duplex#2	Conc	Vol ul	
	Fast blue	X1	12.5	Fast blue	X1	12.5	
	<i>eae</i> F primer	10uM	0.5	<i>vtx2</i> F primer	10uM	0.5	
	<i>eae</i> R primer	10uM	0.5	<i>vtx2</i> R primer	10uM	0.5	
YakY/ Joe	<i>eae</i> probe	1uM	0.5	<i>vtx2</i> probe	1uM	0.5	YakY/ Joe
	<i>vtx1</i> F primer	10uM	0.5	O157 F primer	10uM	0.5	
	<i>vtx1</i> R primer	10uM	0.5	O157 R primer	10uM	0.5	
Cy5	<i>vtx1</i> probe	1uM	0.5	O157 probe	1uM	0.5	FAM
	Water		7.0	Water		7.0	
	DNA target		2.5	DNA target		2.5	
	Total		25	Total		25	

# VTEC isolates for validation (545)

Source	Strain types	Number
England and Wales in 2010	O157	169
(i) UK and Ireland 2004-2010 (ii) UK and abroad pre-2004	Non-O157 <sup>1</sup>	310
VTEC EQA isolates from ring trials	Non-O157	66

<sup>1</sup> Includes clinically-important serogroups from human disease

# Concordance of previous block and real time methods

<i>vtx</i> genes	RT Agree	RT Disagree	Reason
Block O157	169 (100%)	0	
Block non-O157	372 (99%)	4 (1%)	<i>vtx1d</i> subtype

- 100% of the *E. coli* O157 strains were detected in the O157 *rfbE* PCR
- None of the *E. coli* of other serogroups was positive in the O157 *rfbE* PCR
- Real time *eae* PCR detected 100% isolates positive by block-based assay

## Ability to detect subtypes of *vtx* genes

<i>vtx</i> type	Subtype	Real time	Routine block
<i>vtx1</i>	<b>a</b> , b, c	Yes	Yes
<i>vtx1</i>	d	Yes	No
<i>vtx2</i>	<b>a</b> , b, <b>c</b> , <b>d</b> , e, g	Yes	Yes
<i>vtx2</i>	f	No	No

Most *vtx2f* strains produce intimin, so all *eae*-positive, *vtx*-negative strains can be tested with a specific *vtx2f*-PCR

## Application to faeces samples

214 specimens: cases with diarrhoeal symptoms and possible links to Germany for detection of VTEC O104 between 25<sup>th</sup> May and 24<sup>th</sup> June 2011

- Plated on MacConkey agar, Sorbitol MacConkey agar with and without cefixime and tellurite (CT-SMAC and SMAC)
- Enriched (6h and 18h) in modified tryptone soya broth (mTSB); DNA extracted from enrichment with Instagene (Bio-Rad Catalog# 732-6030)



# VTEC cases detected

**7 specimens positive by PCR for *vtx* genes confirmed by culture**

## VTEC O104

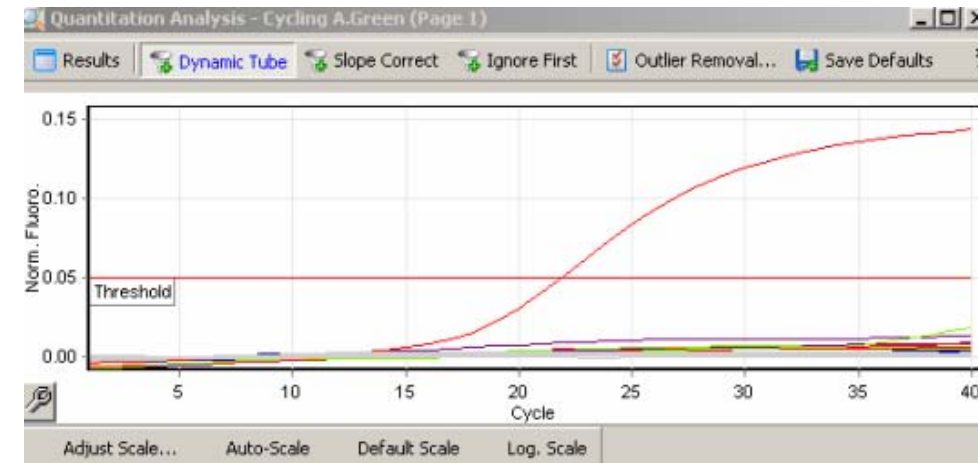
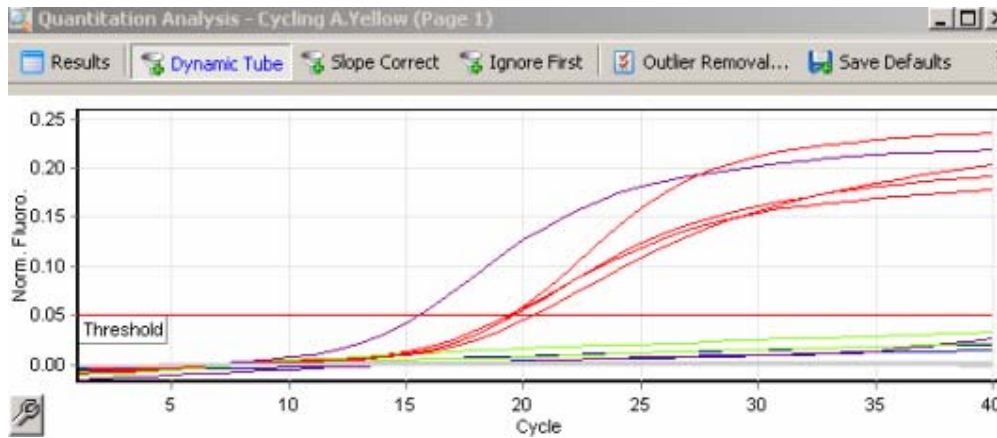
5 specimens positive for *vtx2*, negative *eae*: 8-24 h after receipt  
*vtx2* gene positive isolates obtained by colony picking: 24-30h after receipt

Biochemical ID and serotyping confirmed *E. coli* O104:H4  
PCR positive for the *aggR* gene of enteroaggregative *E. coli*  
Properties were consistent with the VTEC O104 strains isolated in Germany

## Other VTEC

1 isolate *E. coli* O103 (*vtx1*, *eae*).

1 isolate *E. coli* O157 (*vtx2*, *eae*, *rfbO157*)



O104: *vtx2* +ve  
 Negative for *vtx1*, *eae*,  
*rfbO157*

**In total 7 cases of VTEC O104:H4 were confirmed in the UK during the period of the outbreak**

# Conclusions and further work

- Confirmation of *vtx* genes in a wide range of VTEC
- Presence of *eae* gene in many clinically-important VTEC; enables routine detection of EPEC strains
- Molecular serotype for O157 < 4-h of culture receipt

During the VTEC O104 outbreak, the PCR was a rapid and reliable screening method for the detection of cases subsequently confirmed microbiologically by culture

- Generic detection of VTEC in faeces: current analysis of data on >500 samples
- Beginning evaluation of direct nucleic acid extraction from faeces on automated platform
- BUT isolation of VTEC from positive samples will still be time-consuming!!!