PT-05

DETECTION OF ECHINOCOCCUS SPP. WORMS IN THE INTESTINAL MUCOSA OF THE DEFINITIVE HOST

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Workshop of the National Reference Laboratories for Parasites, Rome, Italy, 28-29 October 2025





THE PT IS ACCREDITED IN A QUALITY SYSTEM ACCORDING TO ISO 17043 STANDARD



AIM: to test the competence of the appointed NRLs to detect worms of *Echinococcus* sp. in a matrix made by intestinal mucosa.

PT PANEL: consists in three tubes filled with homogenized intestinal mucosa spiked or not with worms of *Echinococcus* spp.





CALL FOR PARTICIPING

02-3rd

DEADLINE FOR SUBSCRIPTION

02 - 24th

PT SENDING

03 – 17th

DEADLINE FOR SUBMITTING RESULTS

03 – 24th

INDIVIDUAL REPORT

03 - 28th

FINAL REPORT PUBLISHED ON THE EURLP WEBPAGE

03 - 28th

CRITERIA FOR THE QUALITATIVE EVALUATION

• Each PT item is CORRECT if participant detected one or more *Echinococcus* spp. worms in spiked samples OR no worms in not spiked samples...

...OR INCORRECT in case of false positive or false negative results.

• The <u>FINAL EVALUATION</u> is only based on <u>qualitative</u> evaluation and is expressed as "<u>POSITIVE</u>" if the results of all samples are correct OR "<u>NEGATIVE</u>" if at least one result is incorrect.



Albania, Institute of Public Health

Austria NRLP, Austrian agency for health and food safety

Belgium NRLP, Institute of Tropical Medicine

Cyprus NRLP, Veterinary Services

Croatia NRL, Croatian Veterinary Institute

Czech Republic NRLP, State Veterinary Institute

Denmark NRLP, Statens Serum Institut, laboratory of parasitology

Estonia NRLP, Animal Health, Veterinary and Food Laboratory

Finland NRLP, Finnish Food Authority, Ruokavirasto

France NRL Echinococcus, ANSES, LRFS Nancy

Germany NRL Echinococcus, Friedrich-Loeffler-Institut fur Epidemiologie

Greece NRL Department of Parasitology-Parasitic Diseases Entomology & Bee Health

Hungary NRLP National Food Chain Safety Office, Laboratory of Parasitology, Fish and Bee Diseases

Iceland NRLP, Institute for Experimental Pathology Keldur

Ireland NRLP, Parasit section, Bact/Paras Division

Italy NRL, National reference laboratory for Echinococcosis, IZS della Sardegna

Latvia NRLP, Institute of food safety, animal health and environment

Lithuanian NRLP, National Food and Veterinary Risk Assessment Institute

Northern Ireland, AgriFood and Busciences Institute (AFBI)

Norway NRLP, Norwegian Veterinary Institute

Poland NRLP, National Veterinary Research Institute

Portugal NRLP, Instituto nacional de investigacao agraria e veterinaria

Republic of North Macedonia, Faculty of Veterinary Medicine

Romania NRLP, Institute for diagnosis and animal health

Slovakia NRLP, Veterinary and Food Institute in Bratislava

Slovenia NRLP, University of Ljubljana, Veterinary Faculty

Spain NRLP, Laboratorio Central de Sanidad Animal

Sweden NRLP, National Veterinary Institute

Switzerland, Institute of Parasitology Vetsuisse Faculty, University of Bern

The Netherlands NRL, National Institute for Public Health and the Environment

UK, NRL for Trichinella and Echinococcus, Animal and Plant Health Agency,

PARTICIPANTS (31)







- Carnivore intestines are collected and stored frozen at -80°C for one week.
- Faecal content is analysed by qPCR to exclude *E. multilocularis* infection.

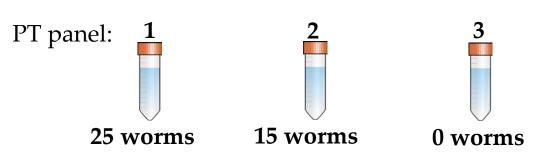


Finnish Food authority Ruokavirasto, Finland



- The mucosa of the small intestine of foxes is collected, cleaned and sent to ISS.
- Mucosa is homogenised (with 70% ethanol; ratio 2:1), aliquoted, and spiked (double check).







Worms have been previously deactivated (-80°C) by the providers.

Piwet, Poland University of Zurich, Switzerland Anses, France

SUMMARY OF RESULTS

Number of participant laboratories submitting results	31/31 (100%)
Number of participants that passed the PT	26 /31 (83.87%)
Number of participants that failed the PT	5/31 (16,13%)

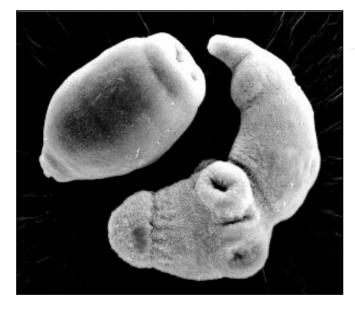


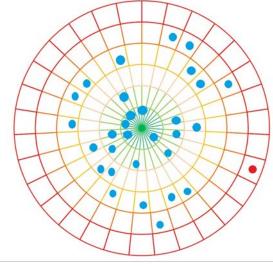
2. False negative, 0 -> 15

3. False positive, $1 \rightarrow 0$

4. False positive, $8 \rightarrow 0$

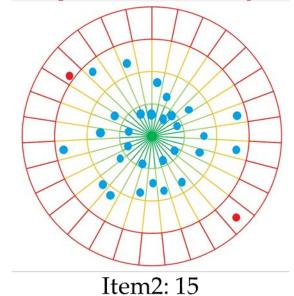
5. False positive, 8 -> 0 & false negative, 0 -> 15





Item1: 25

Mean of detection: 12



Mean of detection: 11

1 over-estimation (21) Without it, mean=10 Item3: 0 Mean of detection: 0,7

ABOUT METHODOLOGY

- The most of the labs (26/31) used the Sedimentation and Counting Tecnique (SCT) with no relevant modification.
 - One lab (1/31) used the PBS instead of NaCl 0,9% (results were: 8, 10, 0 -> correct ✓)
 - One lab (1/31) directly diluted the samples drop by drop in water under the binocular microscope (results were: 14, 10, $0 \rightarrow \text{correct} \checkmark$)
 - Three labs (3/31) poured the samples through a sieve
 - -180 uM (results were: 21, 9, 0 -> correct \checkmark)
 - -150 uM (results were: 8, 10, 0 -> correct \checkmark)
 - 50 uM (results were: 18, 10, 0 -> correct \checkmark)



ADDRESSING COMMENTS

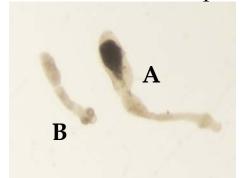
- One lab observed **all** the worms floating in both positive items (no modification to SCT).
- Two labs observed Taeniidae worms different from *Echinococcus*. That's true $\sqrt{}$



• Two labs observed at least two different kind of *Echinoccous* (different in size) . That's true ✓

Reasoning:

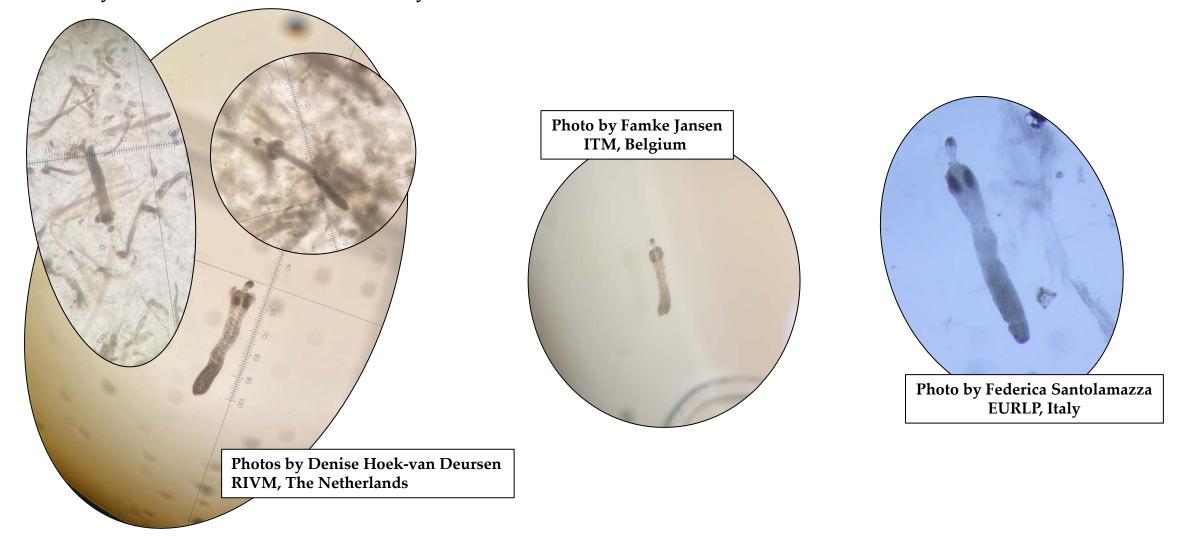
- -No single provision was sufficient for all items.
- -It is necessary to maintain homogeneity across items.
- -The three different batches were used in the same proportion, giving preference to the batch with larger worms. For example, out of 25 worms: 15 from Batch A, 5 from Batch B, and 5 from Batch C.



• Two labs observed Amobotaenia-like worms. That's true √



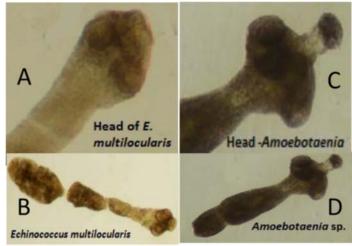
• This year, the mucosa was naturally infected with cestode worms similar but different from *Echinococcus*.



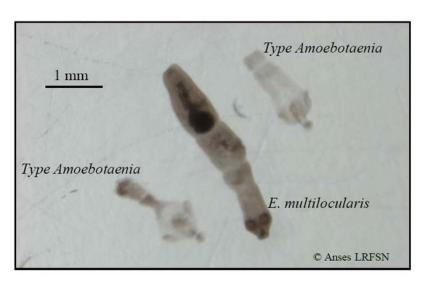
• Similar occurrence in 2022-PT05.



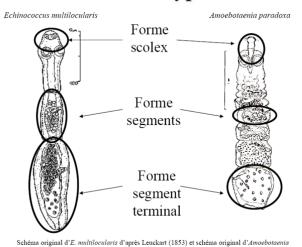
At the time, we studied this new finding a bit.







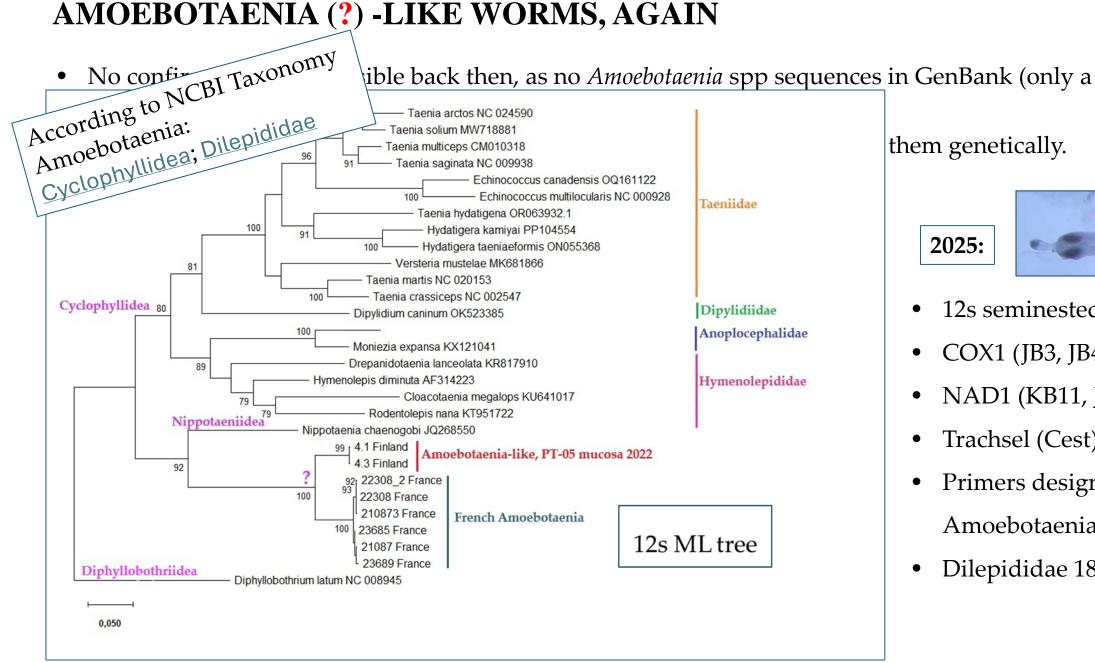
Comparaison entre *E. multilocularis* et 2 cestodes de type *Amoebotaenia*



paradoxa d'après Petavy et Deblock (1980).

Anses LRFSN LNR Echinococcus spp. Technopôle agricole et vétérinaire Bâtiment H 54220 MALZEVILLE

Courtesy of F. Janssen



them genetically.

2025:

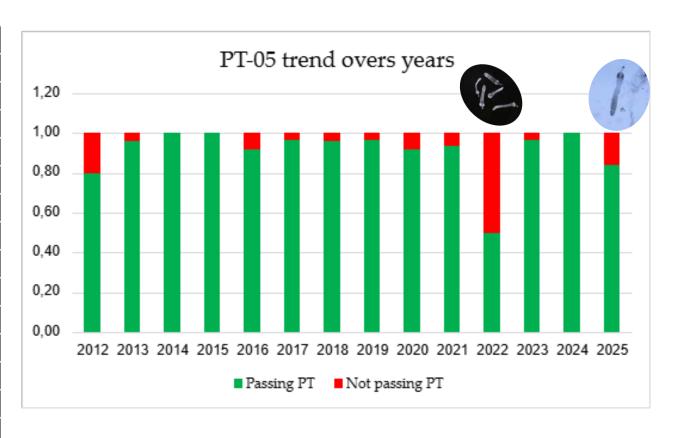


- 12s seminested: neg
- COX1 (JB3, JB4): neg
- NAD1 (KB11, JB12): neg
- Trachsel (Cest): neg
- Primers designed on old Amoebotaenias: neg
- Dilepididae 18S: neg

2025 HEATMAP OF RESULTS AND TREND OVER YEARS

EM1	6	Ø	0
EM2	4	5	6
EM3	15	1 0	0
EM4	9	5	6
EM5	14	10	0
EM6	11	Ø	6
EM7	17	1	0
EM8	10	Ø	6
EM9	6	4	6
EM10	18	12	0
EM11	0	Ø	6
EM12	23	11	8
EM13	18	10	0
EM14	15	11	0
EM15	12	6	6
EM16	4	6	6

EM17	7	4	6
EM18	8	10	0
EM19	3	6	8
EM20	1	a	6
EM21	21	9	6
EM22	20	11	0
EM23	17	Ø	6
EM24	10	4	6
EM25	25	21	0
EM26	20	9	6
EM27	7	6	6
EM28	9	4	6
EM29	18	12	0
EM30	11	6	1
EM31	8	6	6



2022: 14/28 false positive results

2025: 3/31 false positive results

1/3 did not participate 2022 PT

Increased awareness?

CONCLUSIONS

- In 2025, 26 out of 31 laboratories (84%) successfully passed PT-05.
- Detection performance appears robust, even when methods other than SCT are used.
 - The presence of natural infections with cestodes other than *Echinococcus* resulted in fewer errors compared to previous rounds, reflecting the increasing proficiency of NRL personnel.



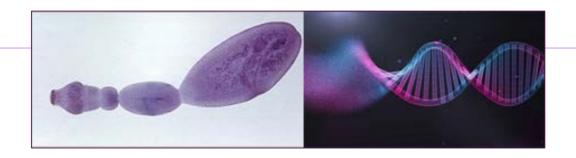
PT-08

European Union Reference Laboratory for Parasites

Unit of Foodborne and Neglected Parasitic Diseases

Department of Infectious Diseases
ISTITUTO SUPERIORE DI SANITÀ

Molecular identification of *Echinococcus* granulosus, *Echinococcus* multilocularis and *Taenia* spp



XX Workshop of National Reference Laboratories for Parasites 28th and 29th October 2025

Istituto Superiore di Sanità

Federica Santolamazza, Azzurra Santoro

European Union Reference Laboratory for Parasites (EURLP);
WHO Collaborating Centre for the Epidemiology, Detection and Control of Cystic and Alveolar Echinococcosis;
ISTITUTO SUPERIORE DI SANITÀ (Rome, Italy)



Due date to submit results: April 11, 2025
Individual report sent to participants within: April 30, 2025
Final report available on EURLP website from: May 31, 2025



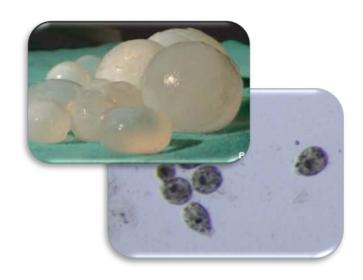
UPGRADE

Aim of the PT

Evaluation of laboratories competence in molecular identification of

Echinococcus granulosus sensu lato,

Echinococcus multilocularis and Taenia spp









spp.-2025

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PT timing 2025



February 24th





https://www.iss.it/en/web/iss-en/eurlp-proficiency-testing

March 17th



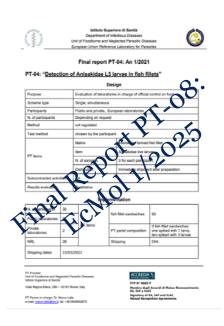
April 11th



April 30th



May 31st





European Union Reference Laboratory for Parasites

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Preparation of samples PT08-2025

The PT has been organized following the NRL request (2025)

The panel consists of 4 tubes:

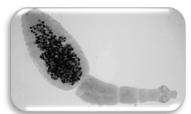
3 tubes containing DNA extracted from canine faecal stool **spiked** with:

- E. granulosus s.s. cyst
- E. multilocularis worms
- Taenia hydatigena proglottid









Negative: Anisakis DNA



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Detection method PT08-2025

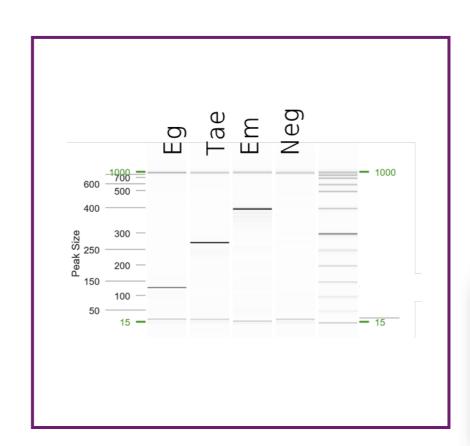
The samples were identified at species level through multiplex PCR

E.g. = *E. granulosus* (117 bp)

E.m. = *E. multilocularis* (395 bp)

Tae= Taenia spp (267bp)

Negative= Anisakis spp







Multiplex PCR (Trachsel et al. 2007)

In 30 µl total reaction with 0.2 mM of each primer (Cest1, Cest2, Cest3, Cest4, Cest5)

Dilution 1:10 and 1:100

COX1 PCR and sequencing (366bp)



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Detection method PT08-2025

Homogeneity was ensured by providing participants with aliquots of the same DNA preparation.



The tubes were plugged and sealed using plastic parafilm, individually coded.



Each PT panel was inserted in polystirene box with ice pack

All panels were delivered within 24-36 hours.







spp.-2025

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Evaluation criteria

The PT evaluation is qualitative, and no statistical analysis of the results is applied

Lab code	Expected	Observed	Result (correct/incorrect)	Evaluation (positive/negative)
	E. granulosus	E. granulosus	correct	
EGM X	Negative	Negative	correct	Positive
	Taenia spp.	Taenia spp.	correct	Fositive
	E. multilocularis	E. multilocularis	correct	
EGM X	E. granulosus Negative Taenia spp. E. multilocularis	E. granulosus Taenia spp. Negative E. multilocularis	correct incorrect correct	Negative
EGM X	E. granulosus Negative Taenia spp. E. multilocularis	E. granulosus Negative <mark>Taenia spp</mark> . Negative	correct correct incorrect correct	Negative

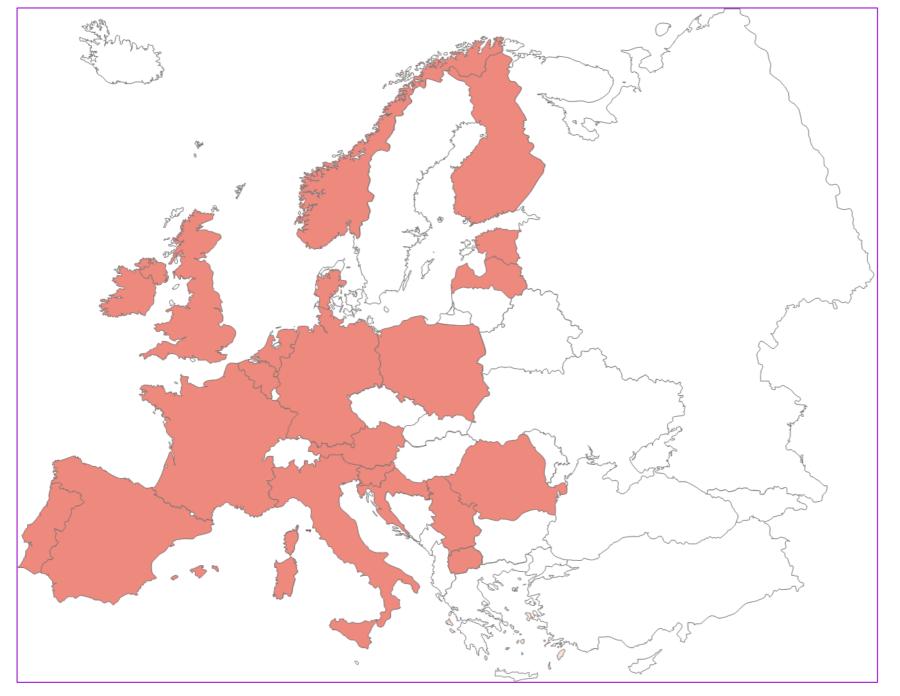
The result is "correct" if the PT items are correctly identified The result is "incorrect" if the PT items are incorrectly identified

The PT is considered "POSITIVE" if the results of all samples are "correct" The PT is considered "NEGATIVE" if at least one result is "incorrect"



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Participants (22)





Austria

Belgium

Croatia

Denmark

Estonia

Finland

France

Germany

Ireland

Italy

Italy (2° lab)

Latvia

Norway

Poland

Portugal

Rep. North Macedonia

Romania

Slovenia

Spain

Switzerland

The Netherlands

UK

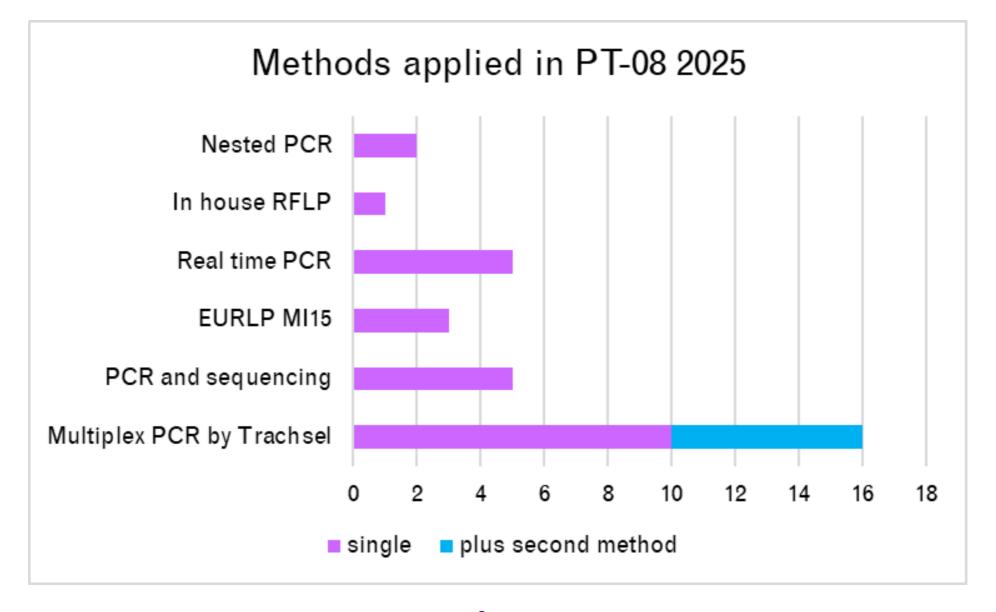




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Methods

The EURLP recommends the use of the multiplex PCR by Trachsel *et al.* however, any other suitable molecular-based method is accepted





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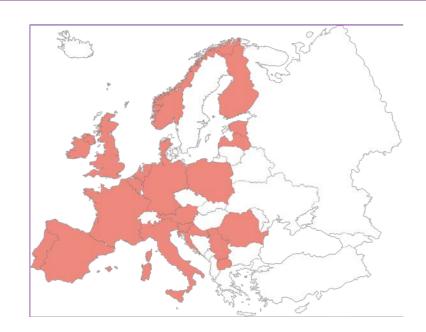




Participants N=22

Participants passed N=20

Participants failed N=2



Sample1 (E. granulosus s. l.): 22 labs (100%) out of 22 obtained a positive evaluation

Sample 2 (negative): 21 labs (95% 1. false positive -> the lab used the valuation Multiplex PCR by Trachsel, 2007

Sample 3 (*Taenia spp.*): 21 labs (9 2, false negative -> the lab used

the Multiplex by Trachsel, 2007 and the nad1 PCR & sequencing.

e evaluation

Sample 4 (E. multilocularis): 22 The lab reported that they did not include primers for Taenia spp.

a positive evaluation

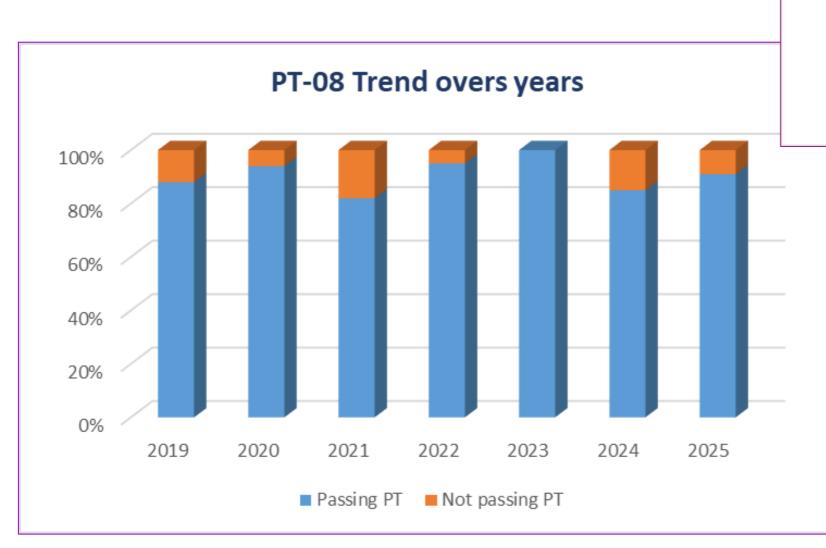
Carefully read emails, procedure and instructions!





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Suggestions, based on this year's experience:

Species identified during the Proficiency Test:

defined by:

Procedure

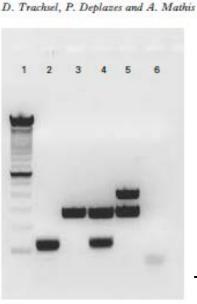
PT items

<u>Description</u>. The PT panel consists of four tubes containing: DNA extracted from faecal material contaminated with <u>Echinococcus granulosus sensu lato</u> (s.l.), DNA extracted from faecal material contaminated with <u>Echinococcus multilocularis</u>, DNA extracted from faecal material contaminated with <u>Taenia spp.</u> and a <u>negative</u> control.

Mixed-DNA items?

Not so far.

However, this could be a possible upgrade of the PT. If implemented, the procedure will be updated.





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Conclusions:

Out of 22 laboratories, 20 successfully passed the PT.

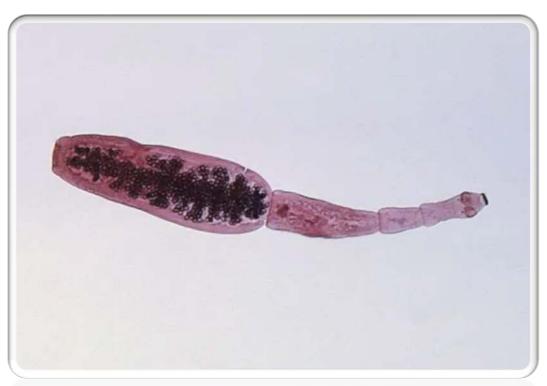
No specific errors were identified.

Overall, all the chosen methods appear to perform well, and NRL personnel demonstrate sufficient expertise to handle the PT-08, even when new challenges are introduced.



Thanks for your

attention



ACKNOWLEDGMENTS

Federica Santolamazza Alessia Possenti

